## **Supplementary Material**

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Fig. S1. The CCAP/bursicon cells grow but maintain a largely constant gross morphology during larval development. (A-C) CCAP/bursicon neurons from embryonic stage 17 (A), the second larval instar (B), and the wandering 3rd larval instar (C). (D) Quantification of soma size for CCAP/ bursicon cells of the genotypes shown in panels A-C. From embryonic stage 17 to the wandering 3rd instar larval stage, the soma size (cross-sectional area) of the CCAP/bursicon neurons was increased over 2.5-fold. Bars labeled with different letters are significantly different (P<0.0001, one way ANOVA; Tukey-Kramer post-hoc test). Scale bars: 20 µm (A), 50 µm (B), 100 µm (C).





■ GFP

Burs



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Fig. S3. Expression of foxo or InRact driven by burs-Gal4 altered metamorphic growth of the CCAP/ bursicon cells. (A) bursicon-Gal4driven co-expression of UASmCD8::GFP and UAS-foxo caused a significant reduction in soma and peripheral arbor growth and the number of visible somata (middle panels) at the pharate adult stage. In contrast, expression of *InR*<sup>act</sup> significantly increased soma size and the area covered by the peripheral axon arbor (lower panels). There was substantial overlap of the CD8::GFP (cyan) and anti-bursicon immunostaining (magenta) signals, and strong CD8::GFP expression persisted in the somata, suggesting that the reduction of peripheral axon arbor area with foxo overexpression was not due to the loss of cell markers. (B) The expression level of CD8::GFP driven by bursicon-Gal4 was higher than the one driven by ccap-Gal4. (C-E) Quantification of GFP fluorescence intensity (C), soma area (D, black bars), soma number (D, gray bars), and peripheral arbor area (E) of the CCAP/bursicon cells shown in genotypes in panels A and B. Means labeled with different letters were significantly different. A Student's t-test was performed on GFP intensity (P<0.001). One-way ANOVAs and Tukey-Kramer post-hoc tests were performed on soma size (P < 0.001). soma number (P < 0.001), and peripheral arbor area (P < 0.001). Scale bars: 20  $\mu m$  (A, CNS), 200  $\mu m$  (A, peripheral arbor), 50 µm (B).





Fig. S5. Overexpression of *foxo* disrupted metamorphic outgrowth of the CCAP/bursicon cells. Anti-bursicon immunostaining of the ventral nerve cord at 0, 24 and 48 hours after puparium formation (APF). In contrast to the controls (A, with only the *ccap-Gal4* driver), CCAP/bursicon cell-targeted expression of *foxo* (B) disrupted soma growth (solid arrows) and outgrowth of adult-specific neurites (solid arrowheads). A similar extent of pruning of the larval neurites was observed at the 24 hour time point in the two genotypes. Feathered arrows, lateral longitudinal tracks; feathered arrowheads, midline arbor. Scale bar: 100 µm.



Fig. S6. InR inhibited the effects of FOXO in the CCAP/bursicon cells. Cell-targeted expression of InR in the CCAP/bursicon neurons completely rescued the cell loss and cell growth phenotypes induced by *foxo* overexpression. (A,A') *ccap-Gal4* driver-only controls. (B,B') *UAS-InR*-only controls. (C,C') *ccap-Gal4; UAS-foxo*. (D,D') *ccap-Gal4; UAS-InR*. (E,E') *ccap-Gal4; UAS-InR, UAS-foxo*. Scale bars: 50 µm (A–E), 200 µm (A'–E').



Fig. S7. InR<sup>RNAI</sup> did not reduce soma or synapse growth in larval CCAP/bursicon cells. (A,B,A',B') Celltargeted expression of  $InR^{RNAi}$  with Dcr-2 in the CCAP/ bursicon neurons had no effect on the general morphology of the CCAP/bursicon neuron somata and CNS projections (A,A'), peripheral axon projections (B,B'), or neuromuscular junction (NMJ)-like endings (anti-bursicon immunostaining on w3<sup>rd</sup> larvae). (A,B) UAS-Dcr-2, CCAP-Gal4 controls. (A',B') UAS-Dcr-2, CCAP-Gal4/ UAS-InR<sup>RNAi</sup>. Scale bars: 100 µm (A,A'), 200 µm (B,B'). (C-E) InR<sup>RNAi</sup> had no significant effect on soma area (C) (P>0.05, Student's *t*-test), bouton number (D) (P>0.05, Student's *t*-test).



Fig. S8. Moderate effects of IIS on the metamorphic growth of Tv neurons. (A–C) Cell-targeted expression of  $InR^{act}$  or  $InR^{DN}$  in the Tv neurons changed the soma size (B,C) and the area covered by the peripheral axon arbor (B',C') (anti-bursicon immunostaining, stage P14 pharate adults). (A,A') *fmrf-Gal4* driver-only controls. Scale bars: 100 µm. (D,E) Altered InR activity significantly influenced the soma size of Tv neurons (D) and the peripheral axonal arbor area (E). One-way ANOVA; Tukey–Kramer *post-hoc* tests were performed on soma size (P<0.0001) and peripheral axon arbor (P<0.01). Means labeled with different letters are significantly different.

 Fig. S9. Reduced levels of DILP7 peptide in the dMP2 neurons of Odd>diln7<sup>RNAi</sup>, dicer-2 animals, (A,B) Stage P14 pharate adults express

B

OddGal4>DILP7<sup>RNAi</sup>, dicer2

DILP7<sup>RNAi</sup>, dicer2

Fig. S9. Reduced levels of DILP7 peptide in the dMP2 neurons of  $Odd > dilp7^{RNAi}$ , dicer-2 animals. (A,B) Stage P14 pharate adults expressing UAS-dilp7<sup>RNAi</sup> with UAS-dicer2 under the control of Odd-Gal4 displayed reduced levels of DILP7 peptide (B) compared to UAS-dilp7<sup>RNAi</sup>, UAS-dicer2-only controls (A) (anti-DILP7 immunostaining, stage P14 pharate adults). Scale bar: 25 µm.