Comparative Analysis of Opioid-Induced Microbiome Alterations in Rat Small Intestine, Cecum, and Colon

Gwen Reilly¹, Senait Assefa¹, Alejandro Torres², Songjukta Chakraborty¹, Shilpa Dange¹, Halla Hamdan¹, Ayomide Ishola¹, Serene Lim¹, Joe McCreary¹, Thomas Momanyi¹, Bhuvanesh Kumar Raju¹, Logan Swope¹, Dolores Vazquez Sanroman², and Gerwald Koehler¹

¹ Department of Biochemistry and Microbiology, OSU Center for Health Sciences ² Department of Anatomy and Cell Biology, OSU Center for Health Sciences

INTRODUCTION

Trillions of bacteria, archaea, fungi, and viruses comprise the animal microbiome and virome. The composition and diversity of these complex communities have profound impacts on the health of their human or animal host. Changes in the composition of these microbiome profiles are multifactorial such as diet, health, or medication. Embedded in these communities might be groupings such as individual bacterial species/strains or consortia that have key roles and could serve as biomarkers for host health and disease. Due to new sequencing techniques and data analysis approaches, the characteristics of healthy (eubiotic) and dysbiotic microbiomes are being discovered (Shreiner et.al 2015).

OBJECTIVES

To develop workflows and bioinformatic approaches to identify microbiome profiles within different gastrointestinal regions after oxycodone administration.

METHODS

We are using fecal samples to perform 16S ribosomal RNA gene sequencing at the hypervariable region V4 in order to profile and compare microbiota in the small intestine, cecum, and colon. Nextgeneration sequencing using an Illumina MiSeq system is employed to sequence 16S rRNA amplicons as described by Kozich and coworkers (Kozich et al., 2013). Demultiplexed sample sequence reads are quality controlled and analyzed using the CLC Genomics Workbench (Qiagen) or QIIME 2 (Bolyen et al., 2019). The SILVA ribosomal RNA database is used for sequence read classification (Quast et al., 2013) and dedicated bioinformatics workflows are employed to determine microbial composition, alpha and beta diversities, and differential abundance analyses. All animal procedures used in this study were performed under protocols approved by the OSU-CHS Institutional Animal Care and Use Committee (IACUC).



RESULTS

Biomarker Discovery Workflow



Fig 1: Bioinformatics Workflow. This flowchart summarizes the bioinformatic approaches used to analyze this data.

Alpha Diversity



Fig 4: Alpha diversity analysis. The phylogenetic diversity of microbiota is significantly reduced in the small intestine (a). Differences in the total number of OTU's between the treatment types (saline and oxycodone administration) in the colon microbiota are shown (b). The total number of OTU's in the cecum are not significantly different between male and female rats. This graph illustrates that there are differences between the male and female samples used (c).

Beta Diversity



Fig 5: Beta diversity analysis. The microbiota at the different sites showed differences in beta diversity (a). Sex differences in Bray-Curtis dissimilarity were seen in cecal samples (b). Oxycodone administration affected the microbial composition in cecal samples (Bray-Curtis) OXY, oxycodone, SAL, saline (c).



Relative Abundance-Phylum Level







Fig 2: Relative abundance at the phylum level. Taxonomic classification revealed differences in relative abundance of phyla between the small intestine, cecum, and colon. The phylum Bacteriodetes appears to be less abundant in most small intestinal samples compared to the cecum and colon samples.



Genus Level Abundance Tables

Fig 3: Relative abundance of genera at the three intestinal sites. The small intestinal microbiota are remarkably different to cecal and colon microbiotas at the genus level

LEfSe Analysis



Fig 6: Biomarker discovery. LEfSe analysis of the rat cecum microbiota in the different treatment groups of oxycodone and saline for first treatment and saline and naloxone for the second treatment. This provides analysis of the bacterial taxa affected by each treatment that could provide potential biomarkers of health and disease in the gut microbiome. OXN, oxycodone-naloxone; OXS, oxycodone-saline; SAN, saline-naloxone; SAS, saline-saline;

OKLAHOMA STATE UNIVERSITY CENTER FOR HEALTH SCIENCES

LIMITATIONS

Microbiota profiling using 16S rRNA gene sequencing is limited due to

- 16S rRNA primer biases
- Absolute versus relative abundance
- RNA operon number differences in bacteria and archaea
- Functional profiles can at best only be predicted
- Elucidation of causal relationship require in-depth functional studies of microbiota-host interactions



CONCLUSION

Analysis of animal microbiotas by 16S rRNA gene sequencing has provided invaluable insights into the role of the microbial communities in health and disease. Correlation of these profiles with host metadata will aid in the identification of potential biomarkers of oxycodone use in the different host organ environments that could lead to novel diagnostic targets and preventative or therapeutic interventions. However, these analyses must be reinforced with functional evaluations to elucidate causal relationships.



REFERENCES

Bolyen E, Rideout JR, Dillon MR, Bokulich NA, Abnet CC, Al-Ghalith GA, Alexander H, Alm EJ, Arumugam M, Asnicar F, et al.: Reproducible, interactive, scalable and extensible microbiome data science using QIIME 2. Nature Biotechnology 2019, 37:852-857.

Quast, C., Pruesse, E., Yilmaz, P., Gerken, J., Schweer, T., Yarza, P., Peplies, J., and Glockner, F.O. The SILVA ribosomal RNA gene database project: improved data processing and web-based tools. Nucleic Acids Res 2013, *41*, D590-596.

Kozich, J.J., Westcott, S.L., Baxter, N.T., Highlander, S.K., and Schloss, P.D. Development of a dual-index sequencing strategy and curation pipeline for analyzing amplicon sequence data on the MiSeq Illumina sequencing platform. Applied and Environmental Microbiology 2013, *79*, 5112-5120. Segata, N., Izard, J., Waldron, L., Gevers, D., Miropolsky, L., Garrett, W.S., and Huttenhower, C. Metagenomic

biomarker discovery and explanation. Genome Biol 2011, *12*, R60. Shreiner AB, Kao JY, Young VB. The gut microbiome in health and in disease. Curr Opin Gastroenterol. 2015

Jan;31(1):69-75. doi: 10.1097/MOG.0000000000000139. PMID: 25394236; PMCID: PMC4290017.

ACKNOWLEDGEMENTS



The research results presented here were made possible in part thorough funding to GK and DVS from the Oklahoma Center for Adult Stem Cell Research (OCASCR), a program of TSET. We are thankful to Dr. Rob Allen, OSU-CHS Department of Forensic Sciences, for allowing us access to the Illumina MiSeq and Ion Torrent PGM sequencing systems.