Loss-of-function mutants and overexpression lines of the Arabidopsis cyclin CYCA1;2/TAM exhibit different defects in prophase-I meiocytes but produce the same abnormal meiotic products

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ABSTRACT

Cyclins are known regulators of both meiotic and mitotic cell cycle progression in diverse organisms. In Arabidopsis, loss-of-function mutations in the A-type cyclin CYCA1;2/TARDY ASYNCHRONOUS MEIOSIS (TAM) gene lead to the production of abnormal meiotic products including triads and dyads. To investigate the effect of overexpression of TAM on meiosis, we generated Arabidopsis plants that harbored the ASK1:TAM transgene that increased TAM expression. 14 of 36 T₁ plants, representing five of six independent transformations, produced dyads or a mixture of dyads, triads and tetrads in male meiosis, indicating that the transgene had a dominant-negative effect. 34 of the 36 T₁ plants with or without the abnormal male meiotic products produced shriveled seeds, suggesting that the seed phenotype was more sensitive to the overexpression of TAM than, and independent of, the meiotic phenotype. To understand how overexpression of TAM produced the same meiotic products as the loss-of-function mutants, we conducted detailed analysis of spread prophase-I chromosomes in the wild type (WT), tam-1, and an ASK1:TAM line. We found that both the synapsed and unsynapsed chromosomal regions in zygotene were thicker in tam-1 and the ASK1:TAM line than in the WT, but in pachytene, synapsed chromosomes in tam-1 and the ASK1:TAM line were thinner and thicker than in the wild type, respectively. In diplotene, only the synapsed chromosomal regions in the ASK1:TAM line continued to be thicker than in the wild type while the desynapsed chromosomal regions among the three genotypes and the synapsed chromosomal regions between the wild type and tam-1 were not statistically different. We also found that the pericentromeric heterochromatin regions in tam-1 and tam-2 formed a tight cluster at the pachytene and diplotene stages in approximately 40 % of the male meiocytes, much higher than the 6% found in the wild type and the ASK1:TAM line. Because the levels of TAM affects chromosomal morphology in specific phases in prophase I, we examined the expression and localization of a prophase-I marker protein ASY1 by immunolocalization in male meiocytes in the wild type, tam-1, tam-2, and two ASK1:TAM lines with different expression levels of TAM. The timing of ASY1 expression in the tam mutants appeared to match that in the wild type, but ASY1 in the null mutant tam-2 was not found to be colocalized with the chromosomes as in the wild type and the partial loss-of-function mutant tam-1. The level of ASY1 was severely reduced in the highly expressed ASK1:TAM line while moderately reduced in the relatively less overexpressed ASK1:TAM line. Our results indicate that the reduction and increase in the activity of TAM differentially affect chromosomal morphology and the action of ASY1 in prophase I. Based on these results, we propose that either the different meiotic defects trigger a common cell cycle checkpoint, or the different defects later lead to a common defect such as missing ASY1 on the chromosomal axes, which further causes the production of the same abnormal meiotic products in the tam mutants and ASK1:TAM lines.

Differential effects of *tam-1 and ASK1:TAM2* on chromosomal thickness in zygotene-to-diplotene male meiocytes (Figs. 3 and 4)



Figure 3. Subtle differences in chromosome thickness among Col, tam-1 and ASK1:TAM2. (A-**C**) Zygotene, pachytene, and early diplotene chromosomes in Col, respectively. (D-F) Zygotene, pachytene, and early diplotene chromosomes in *tam-1*, respectively. (G-I) Zygotene, pachytene, and early diplotene chromosomes in ASK1:TAM2, respectively. The quantitative differences in chromosome thickness was summarized in Fig. 4. Thin arrows, unsynapsed zygotene or desynapsed diplotene chromosome regions. Thick arrows, synapsed chromosome regions. Bar = $25 \mu m$.

WHAT IS KNOWN ABOUT THE ROLE OF TAM IN MEIOSIS

• TAM is predominantly expressed at the pachytene stage in male meiosis but it regulates cell cycle progression at the pachytene and post-pachytene stages; tam mutants produce abnormal meiotic products such as triads and dyads.

•. TAM acts in the same pathway as OSD1, TDM1, and SMG7; OSD1 is an APC/C inhibitor, TDM1 has no known functional domain, and SMG7 functions in nonsense-mediated RNA decay.

• An epistasis order for the mutants of these genes is, starting from the most epistatic, osd1, tdm1, smg7, and tam.

CURRENT FINDINGS

Overexpression of TAM has a dominant effect on meiosis and seed development (Fig. 1 and Table 1)



Figure Thicknesses of synapsed, 4. unsynapsed, and desynapsed chromosomes in male meiocytes in Col, tam-1, and ASK1:TAM2. Shown are average thicknesses ± standard errors. *The values are statistically different from the corresponding values of Col (t-test, p < 0.01, $11 \le n \le 78$). Also note that the synapsed chromosomes in Col and ASK1:TAM2 underwent a thickening process whereas pachytene synapsed chromosomes in *tam-1* underwent a thinning process. Zyg, zygotene, with the left and right and unsynapsed synapsed being chromosomes. Pac, pachytene. Dip, diplotene, with the left and right being synapsed and desynapsed chromosomes.

Pericentromeric heterochromatin regions tend to cluster in *tam-1* and *tam-2* (Fig. 5 and Table 2)





Figure 1. Male meiotic products in the wild type (CoI) and T_1 of ASK1;TAM2. (A) Normal tetrads in CoI. (B-D) A dyad, a triad, and a tetrad in ASK1; TAM2, respectively. Bar = 25 μ m.

Table 1. Male melotic products in ASKT. TAW at the T ₁ generation		
Experiment	Number of T ₁ s	Meiotic products
1	9	Tetrads
	5	Dyads
2	6	Tetrads
	3	Dyads
	1	Mixture of dyads, triads, and tetrad
3	4	Tetrads
4	2	Tetrads
	2	Dyads
5	1	Tetrads
	1	Dyads
6	1	Dyads
	1	Mixture of dyads, triads, and tetrad

Table 1 Male maintie products in ACK1, TAM at the T as

Figure 5. Spread chromosomes at the pachytene or diplotene stage showing scattered and clustered pericentromeric heterochromatin regions. (A) Col. (B) tam-1. (C) ASK1:TAM2. (D) Octaploid tam-2. Large arrows, scattered and elongated pericentromeric heterochromatin regions. Small arrows, conjoined telomeric regions containing rDNA on chromosomes 2 and 4. Arrowheads, clustered pericentromeric heterochromatin regions. Bar = $25 \mu m$.

Table 2. Percentage of meiocytes with a single cluster of pericentromeric heterochromatin regions from pachytene to early diplotene

	22°C	28°C
WT	6 (n = 53)	6 (n = 122)
tam-1	23 (n = 96)	40 (n = 88)
tam-2	41 (n = 27)	Not determined
ASK1::TAM	6 (n = 85)	Not determined

Chromosomal loading and abundance of ASY1 are differentially affected in *tam* mutants and ASK1:TAM lines (Fig. 6)



34 of the above 36 plants produced variable amounts of shriveled seeds (Fig. 2)



Figure 2. Seeds from Col and ASK1:TAM lines. (A) Diploid Col. (**B**) Likely normal and trom an ASK1:TAM2 plant. (C) Mostly shriveled and likely diploid seeds from another ASK1:TAM2 plant. (**D**) Seeds from tetraploid Col. (**E**) Likely normal and tetraploid seeds from an ASK1:TAM1 plant. (E) A mixture of shriveled and normal-looking seeds from another ASK1:TAM1 plant. These seeds appeared larger than the tetraploid seeds on the left.

Figure 6. Immunolocalization of ASY1 in male meiocytes. Shown are DAPI and corresponding anti-ASY1 (green fluorescence) images. Note that ASK1:TAM3 has a much higher expression of TAM than ASK1:TAM2. Bar = 25 µm. The results show that ASY1 was not loaded onto the chromosomes in tam-2 and overexpression of TAM inhibited the expression of ASY1 in the male meiocytes.

CONCLUSIONS

• The reduction and increase in the activity of TAM cause different defects in prophase I.

The different defects either trigger a common cell cycle checkpoint or later lead to a common defect such as missing ASY2 on the chromosomal axes, which further causes the production of the same abnormal meiotic products in the tam mutants and ASK1:TAM lines.

Acknowledgements

This work was supported by grants from the Energy Center at the Environmental Institute, Oklahoma Center for the Advancement of Science and Technology. The ASY1 antibody was a gift from Chris Franklin at the University of Birmingham, UK.