SCHOOL OF FORENSIC SCIENCE The Degradation of Beta-Actin in Blood and Semen Stains

Elizabeth Beaton B.S., Robert Allen Ph.D., Jun Fu Ph.D.

INTRODUCTION

Piecing together a crime scene is one of the most significant components of any forensic investigation and estimating the age of biological stains at a scene can be crucial to the case. Several studies have demonstrated a correlation between the degradation kinetics for mRNA transcripts present in dried body fluid stains aged for varying amounts of time. The relationship between the age of a sample and the state of degradation of many transcripts can be exploited to estimate the age of an unknown crime scene stain. In this laboratory, preliminary work has demonstrated that the 5' and 3' ends of many mRNA transcripts degrade at different rates during aging of body fluid stains.

OBJECTIVES

The purpose of this study is to map the degradation of the entire ACTB transcript (~1800 nucleotides long) in dried blood and semen stains and to determine if variances in degradation sensitivity are uniformly distributed along the length of the molecule. We will also evaluate the kinetics of ACTB mRNA degradation in dried blood and semen stains to determine whether there are any tissue-specific differences in transcript degradation.

METHODS

We have designed qPCR primers from National Center for Biotechnology Information (NCBI) to span the entire length of the ACTB transcript to direct the amplification of ~100 base pair qPCR amplicons. Blood and semen RNA was extracted from collection cards. For each primer site, we separated the blood and semen stains into three categories as noted in **FIGURE 2** followed by real time qPCR using QuantStudio5. We have identified the amplification efficiencies for our collection of primers through qPCR.

RESULTS

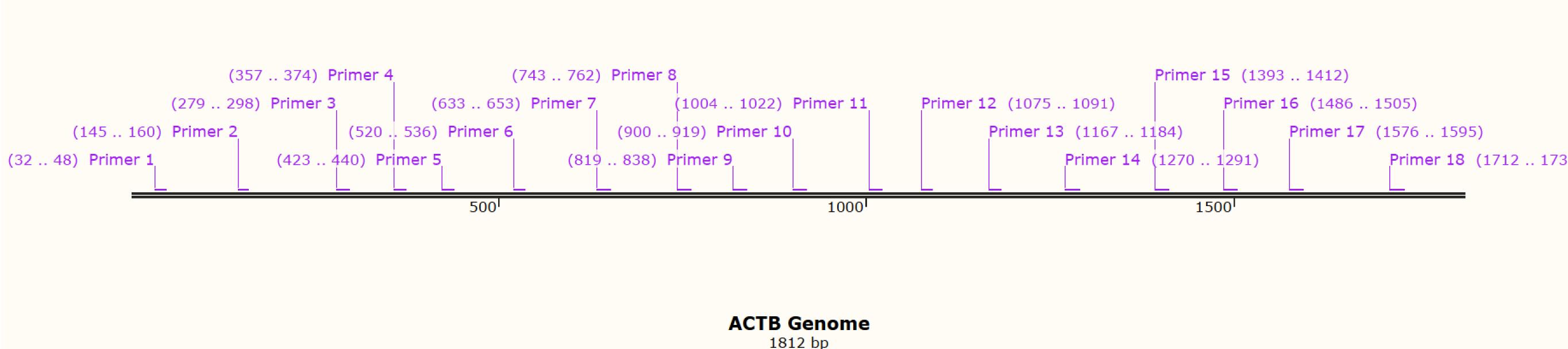


FIGURE 1: All 18 of our designed primers (pictured: forward primers) spanning along the 1812 nucleotide length of the ACTB genome.

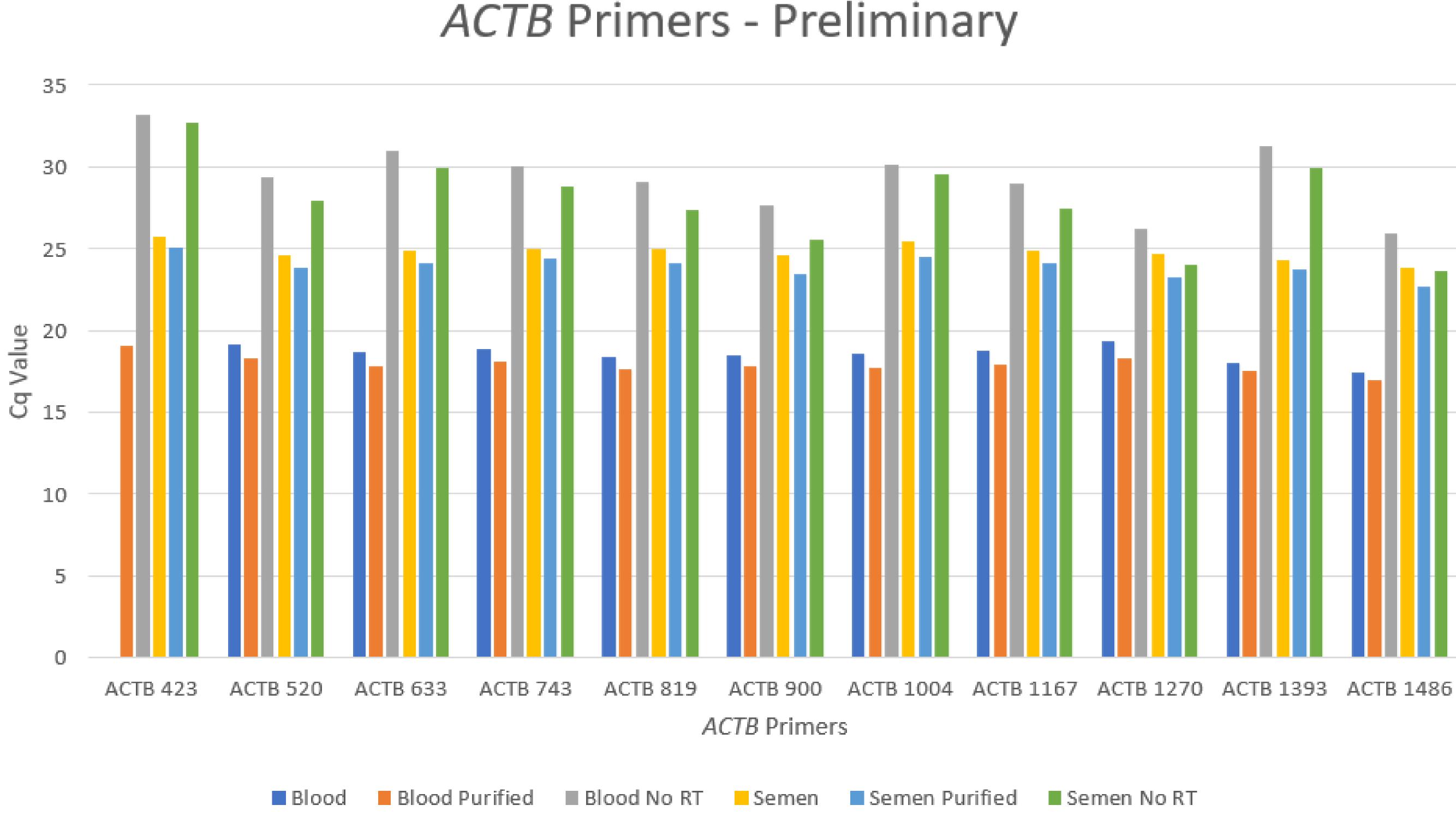


FIGURE 2: Preliminary testing of our first set of primers.





OKLAHOMA STATE UNIVERSITY **CENTER FOR HEALTH SCIENCES**

CONCLUSION

Now that we have our qPCR primers and have determined their amplification efficiencies, we can begin the study of ACTB degradation in our blood and semen stains stored for increasing periods of time up to eight weeks. In order to achieve the best results, we will continue to use reverse transcription and the addition of the inhibitor removal for further purification. The findings of this study will contribute to the knowledge that will be needed to apply this technology to estimate the age of biological evidence recovered from a crime scene.

REFERENCES

1. Weinbrecht, K. D., Fu, J., Payton, M., & Allen, R. (2017). Time-dependent loss of mRNA transcripts from forensic stains. Research and Reports in Forensic Medical Science, Volume 7, 1–12. https://doi.org/10.2147/rrfms.s125782

2. Bremmer, R. H., de Bruin, K. G., van Gemert, M. J. C., van Leeuwen, T. G., & Aalders, M. C. G. (2011). Forensic quest for age determination of bloodstains. Forensic Science International, 216(1-3), 1–11.

https://doi.org/10.1016/j.forsciint.2011.07.027

3. Fu, J., & Allen, R. W. (2018). A method to estimate the age of bloodstains using quantitative PCR. Forensic Science International: Genetics, 39, 103–108. https://doi.org/10.1016/j.fsigen.2018.12.004

4. Heneghan, N., Fu, J., Pritchard, J., Payton, M., & Allen, R. W. (2020). The effect of environmental conditions on the rate of RNA degradation in dried bloodstains. Forensic Science International: Genetics, 51, 102456. https://doi.org/10.1016/j.fsigen.2020.102456

ACKNOWLEDGEMENTS

Thank you to Dr. Allen and Dr. Fu for their assistance and guidance throughout the research process.