



Detection, Conservation, and Augmentation of Naturally Occurring Beneficial Nematodes for Natural Pest Suppression

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The use of natural enemies such as spiders, predatory insects, and parasitoids to control insect pests is fairly well known among plant producers and home gardeners. One category of natural enemies that is less known, however, is the entomopathogens, or "insect pathogens." This group consists of microbial disease agents such as bacteria, viruses, fungi, and entomopathogenic nematodes (EPN) that make insects sick and ultimately kill them. Some of these disease agents are currently available as commercial insecticides, however, many occur naturally in the soil and simply require conservation or augmentation to provide control of soil-dwelling insect pests.

Entomopathogenic nematodes are considered insect pathogens rather than insect parasites because symbiotic bacteria that exist within the gut of these nematodes are released when the nematode enters an insect host and are ultimately responsible for the death of the insect. Entomopathogenic nematodes purchased commercially are likely to be non-native strains that may displace native EPN. By conserving and/or augmenting the native EPN species present in your system (e.g., garden, small or large production facility, or animal pasture), you can promote pest suppression of soil-dwelling insect pests with minimal disruption to the ecological balance of the soil community.

Background

Entomopathogenic nematodes are widely distributed throughout the world and have been isolated from many types of natural and managed habitats in a wide variety of soils. They are naturally occurring pathogens and specific to insects, and are not harmful to the environment, humans, or other vertebrates. Because EPN specifically target soil-dwelling insects and are not harmful to other animals, they are exempt from registration with the Environmental Protection Agency as a commercial insecticide.

Entomopathogenic nematodes are small roundworms, barely visible with the naked eye. The only free-living form of these nematodes is the infective juvenile (IJ), which requires an arthropod host to complete its life cycle. These nematodes function as a naturally occurring biological control agent of arthropods that live all or part of their lives in the soil. Two families of EPN, *Steinernematidae* and *Heterorhabditidae*, each contain many different species that exist commonly in Oklahoma soils. Infective juveniles from different species

exhibit different host-finding behaviors. Some hunt for an insect host on the surface, while others burrow into the soil looking for an arthropod host. Because of variations in host-seeking behavior, it is important to use the correct species of EPN against the specific type of insect pest targeted for control. Table 1 specifies which EPN species are most effective against different types of insect pests.

Life Cycle

Once an arthropod host is located, the IJ enters the host's body through natural openings, such as the mouth, anus, or spiracles (pores for gas exchange). Once inside the insect host, EPN release bacteria that are housed within their gut; the species of bacteria is unique to the family of nematodes and not known to exist anywhere but in the gut of these nematodes. These bacteria ultimately kill the host, usually within 48 hours, and preserve it for the duration of the nematode's life cycle. Completion of the life cycle may take 5 to 10 days for *Steinernematidae* species and 10 to 15 days for *Heterorhabditidae* species (Figure 1). Nematodes develop through two to three generations inside the dead insect before emerging as IJ's from the insect cadaver (Figure 2). The general size of the emerging IJ's can be examined under a microscope to aid in identification of EPN, as some species are distinctly large or small (Table 2). Tens of thousands of new IJ's may be released from one infected host.

The symbiotic bacteria, *Xenorhabdus* spp. (in *Steinernematidae*) and *Photorhabdus* spp. (in *Heterorhabditidae*), are virulent against a broad range of arthropod hosts. Once released by the nematode, the bacteria kill the host insect via bacterial infection and release a wide variety of compounds that act to preserve the insect cadaver in the soil. The bacteria also provide a nutrient source for the developing nematodes. All nematode-infected insect cadavers will have a distinct consistency (firm and rubbery) and stay intact for more than a week, while the nematodes complete their life cycle. Insects that die from something other than EPN infection will typically rot and disintegrate within a day or two of death.

Insects infected with EPN typically exhibit symptoms that are specific to a particular nematode species. For example, cadavers of the greater wax moth larva (waxworms), *Galleria mellonella*, infected with *Heterorhabditis* spp. of nematodes will turn a purplish color due to metabolites produced by their symbiotic *Photorhabdus* bacteria. Waxworms infected with

Table 1. Target pests for entomopathogenic nematodes.¹

<i>Pest insect</i>	<i>Common name</i>	<i>life-stage²</i>	<i>Commodity</i>	<i>Nematode sp.³</i>
COLEOPTERA				
Curculionidae	Billbugs	L	turf	Sc, Hb
	Root Weevils	L	berries, citrus, forest seedlings, hops, mint, ornamentals, sweet potato, sugar beets	Sc, Sk, Hb, Hi, Hm, Sr
Chrysomelidae	Flea beetles	L	mint, potato, sweet potato, sugar beets	Sc
Scarabeidae	Rootworms	L	corn, peanuts, vegetables	Sc, Sr
	White grubs	L	berries, field crops, ornamentals, turf	Hb, Sg, Hm
DIPTERA				
Agromyzidae	Leaf miners	L	ornamentals, vegetables	Sc
Ephydriidae	Shore flies	L	ornamentals, vegetables	Sf
Sciaridae	Fungus gnats	L	ornamentals, vegetables, mushrooms	Sf
Tipulidae	Crane flies	L	turf, ornamentals	Sc, Hm
Muscidae	Filth flies	A	animal rearing facilities	Sf, Hb
LEPIDOPTERA				
Noctuidae	Cutworms	L/P	corn, cotton, peanuts, turf, vegetables	Sc
	Armyworms	L	corn, cotton, peanuts, turf, vegetables	Sc
Pterophoridae	Plume moths	L	artichoke	Sc
Pyalidae	Webworms	L	cranberries, ornamentals, turf	Sc
Sessiidae	Crown borers	L	berries	Sc
	Stem borers	L	cucurbits, ornamentals, shrubs, fruit trees	Sc
Cossiidae	Carpenter worms	L	ornamentals, shrubs	Sc
	Leopard moth	L	apple, pear	Sc
Carposinidae	Peach borer moth	L	apple	Sc
ORTHOPTERA				
Gryllotalpidae	Mole crickets	N,A	turf, vegetables	Sc, Ss, Sr
BLATTODEA				
Blattellidae	German cockroach	N,A	apartments, structures	Sc
SIPHONAPTERA				
Pulicidae	cat fleas	L/P	pet/vet	Sc
NEMATODA				
	Plant-parasitic nematodes same	L/P	turf	Sc

¹ Lacey, L.A. and H.K. Kaya, eds. 2007. Field Manual of Techniques in Invertebrate Pathology.

² L= larva; P= pupa; N= nymph; A= adult

³ Sc = *Steinernema carpocapsae*; Sf = *S. feltiae*; Sk = *S. kraussei*; Sr = *S. riobrave*; Ss = *S. scapterisci*; Hb = *Heterorhabditis bacteriophora*; Hi = *H. indica*; Hm = *H. megidis*

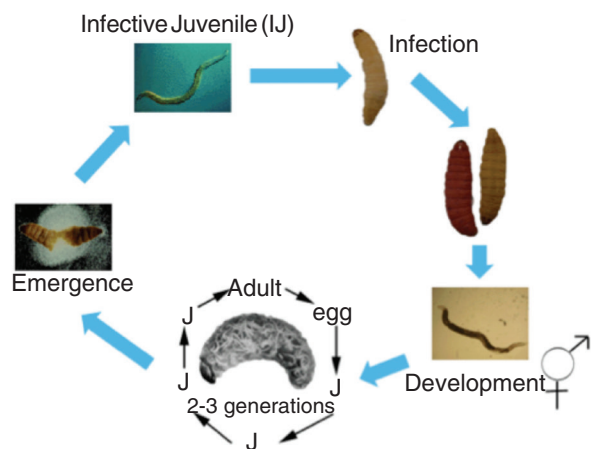


Figure 1. Life cycle of *Steinernematidae* (light brown) and *Heterorhabditidae* (purple) using the greater wax moth, *Galleria mellonella*, as the arthropod host.

Steinernema carpocapsae often exhibit a beige color, while those infected with *Steinernema glaseri* or *Steinernema feltiae* will turn dark brown (Figure 3). Again, the color change is due to the symbiotic bacteria associated with each nematode. Table 2 lists more examples of the symptomatic appearance of insect cadavers.

Conservation of Naturally Occurring Entomopathogenic Nematodes

Entomopathogenic nematodes are sensitive to a variety of stressors. Application of nematicides, some broad-spectrum insecticides, and intense tillage are examples of management practices that can negatively impact beneficial nematodes. Thus, reducing these disturbances can help conserve EPN as well as arthropod predators and parasites. Another conservation strategy is to use soils rich in organic matter, since they tend to be less compacted and have more pore spaces between soil particles; EPN tend to congregate within these



Figure 2. *Heterorhaditidae bacteriophora* infective juvenile (IJ) spilling out of a *Galleria mellonella* cadaver into water.



Figure 3. Healthy *Galleria mellonella* or “waxworm” (left), waxworm infected with *Steinernema carpocapsae* (middle), and waxworm infected with *Steinernema glaseri* (right).

pore spaces on or near the soil surface. Pore spaces may also form around plant roots, worm tunnels, or underneath mulch. Research has shown IJ's to be tolerant of short exposures (2 to 6 hours) of most agrochemicals including herbicides, fungicides, acaricides, and insecticides in solution. Applying these pesticides with adequate amounts of water will promote their survivability.

Table 2. Characteristics of common EPN and infected host cadavers, taken from Lacey and Kaya (2007).

<i>Nematode species</i>	<i>IJ length (µm)</i>	<i>Host cadaver color</i>
<i>S. carpocapsae</i>	558 (468-650)	Beige
<i>S. riobrave</i>	622 (561-701)	Beige
<i>S. feltiae</i>	849 (736-950)	Tan/walnut brown
<i>S. glaseri</i>	1130 (864-1448)	Grayish- dark brown
<i>S. kraussei</i>	951 (797-1102)	Tan/walnut brown
<i>H. bacteriophora</i>	588 (512-670)	Brickred to dark purple
<i>H. indica</i>	528 (479-573)	Dark red
<i>H. megidis</i>	768 (736-800)	Orange brown
<i>H. zealandica</i>	685 (570-740)	Pale mint green

How to Detect Entomopathogenic Nematodes

The use of a living organism to survey for a biological agent is termed “bioassay.” Waxworms are caterpillars of the greater wax moth and are often used to detect the presence of soil pathogens at a given site. Waxworms in their natural state are parasites of honey bee hives, feeding on the wax and normally never coming in contact with the soil. Thus, waxworms have no natural immunity to soil pathogens and will most likely become infected when they come in contact with EPN. Because symptoms of EPN infection include a color change, waxworm caterpillars are ideal organisms for bioassays. Waxworms may be purchased online through a variety of different vendors (simply perform an internet search for waxworms or *Galleria mellonella* and a variety of suppliers should be displayed). Occasionally, they may be purchased at bait shops or pet stores. Any extra waxworms not used for bioassay purposes should be placed in the freezer overnight (to ensure that they are dead) and then discarded.

Methods for Surveying

Entomopathogenic Nematodes in Soil

Entomopathogenic nematodes occur in soils worldwide and it is very easy to survey any area for their presence. . Species commonly collected in Oklahoma soils are listed in Table 3. Rather than examining soil samples for IJ's, which

Table 3. Entomopathogenic nematode species collected from soils in Oklahoma.

<i>Species</i>	<i>Associated Habitat(s)</i>	<i>Efficacy Against (Insect Pests)</i>
<i>Steinernema carpocapsae</i>	Pasture, prairie	Armyworms, black cutworm, billbugs, black vine weevil, clearwing borers, codling moth, corn earworm, crane flies, iris borer, mole crickets, leafminers, white grubs, shore flies
<i>S. diapresi</i>	Pasture, wheat	Natural host unknown
<i>S. riobrave</i>	Pasture	Armyworms, corn earworm, mole crickets, root weevils
<i>S. glaseri</i>	Pasture, prairie, wheat	Black vine weevil, white grubs
<i>S. feltiae</i>	Pasture, prairie, wheat	Armyworms, clearwing borers, codling moth, corn earworm, fungus gnats, leafminers, shore flies
<i>S. texanum</i>	Pasture	Natural host unknown
<i>Heterorhabditis bacteriophora</i>	Pasture, prairie	Billbugs, black vine weevil, clearwing borers, root weevils, corn rootworm, fungus gnats, grape root borer, iris borer, white grubs
<i>H. georgiana</i>	Prairie	Natural host unknown

Data provided by K. Risser, Graduate Research Assistant in the Dept. of Entomology and Plant Pathology at OSU.

are very tiny, the soil is baited with a living organism that will attract IJ's from the soil and then exhibit symptoms indicating infection.

Part I. Soil collection

- Soil should be collected from the site of interest, usually a site that is subject to management practices. This may be a garden, a crop field, a livestock pasture, or lawn.
- Collect soil when it is damp – not saturated and not completely dry.
- Using a trowel, collect your soil sample from the top 3 to 5 inches of soil; take several trowels full of soil from the surface to fill an approximately 12-ounce sample. A paper or plastic cup works well for measuring.
- Each 12-ounce soil sample should be placed into individual zipper-type freezer bags (freezer bags are thicker than sandwich bags, through which waxworms can chew and escape). Close the bag and knead gently to break up the soil.
- The bag should be labeled with the date and location sampled as well as a description of the sample area.
- EPN tend to be patchy in their distribution, so soil samples should be collected from areas that are subject to different applications (e.g., different types of plants, different tillage types, different soil types, etc.). Keep in mind that sampling designs may vary, depending on the type and size of habitat you are sampling (e.g., a diverse garden assemblage will be different than a row crop or pasture).

Part II. Bioassay for entomopathogenic nematodes

- Once the soil samples are collected and labeled, place five healthy waxworm caterpillars into each bag.
- Seal the bag almost all the way; leave a small opening for air to exchange in and out of the bag.
- Set the bags upright in a dark location at room temperature (approximately 75 F).
- After one week, carefully remove the waxworms from all of your soil samples and examine them for signs of infection (Table 2).
- Waxworms may still be alive, some may be dead and rotting (died from something other than EPN infection),

and some may be infected with fungi. If they are infected with EPN they will be dead but uncharacteristically firm and intact. Bacteria released by the nematodes preserve the dead insect and keep it firm for 8 to 12 days while the nematodes complete their life cycles. Infected cadavers will also likely be light tan, dark brown, purple, or pink.

- Each waxworm cadaver should be placed individually on a slightly moist piece of filter paper or paper towel; each cadaver should be placed in a separate dish to prepare for culturing EPN.

Culturing Native Entomopathogenic Nematodes and Augmenting Existing Populations in the Garden

- Place infected cadavers on a moist piece of paper towel in a covered dish (Figures 4 and 5).
- Place the dish loosely inside a plastic bag and seal it most of the way to hold in moisture, but not so much that air cannot get into the bag (Figure 6).
- Place the bagged dish in a dark place at room temperature and check it daily.

Todd Johnson, Agricultural Communications, Oklahoma State University

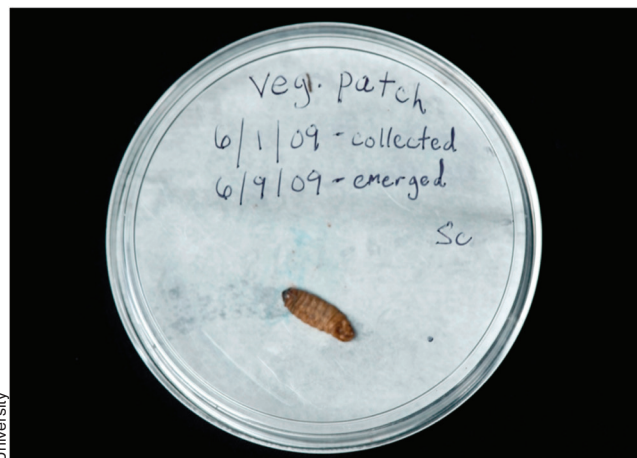


Figure 5. Infected waxworm cadaver on moist piece of paper towel with correctly labeled lid placed on the dish.

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Figure 4. Infected waxworm cadaver paced on a moist piece of paper towel (or filter paper) in a covered dish. The top of the dish is labeled with date and location of the soil sample that yielded the infected cadaver.

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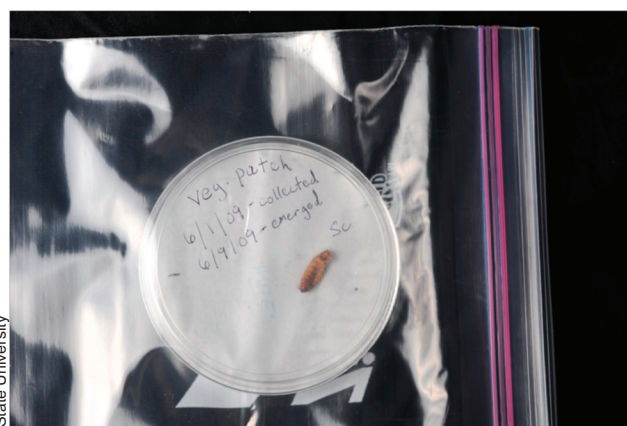


Figure 6. Covered dish containing infected waxworm cadaver, properly labeled with date and location information, placed inside a zipper-type freezer bag with the bag left partially open to allow airflow to the cadaver.



Figure 7. The bottom portion of the small covered dish has been inverted, with the moist piece of paper towel placed on the bottom of the dish. This portion of the dish is placed inside a larger covered dish containing a thin layer of water at the bottom. As new infective juveniles emerge from the infected cadaver, they will migrate down the small dish into the layer of water, where they can easily be poured into a storage flask.

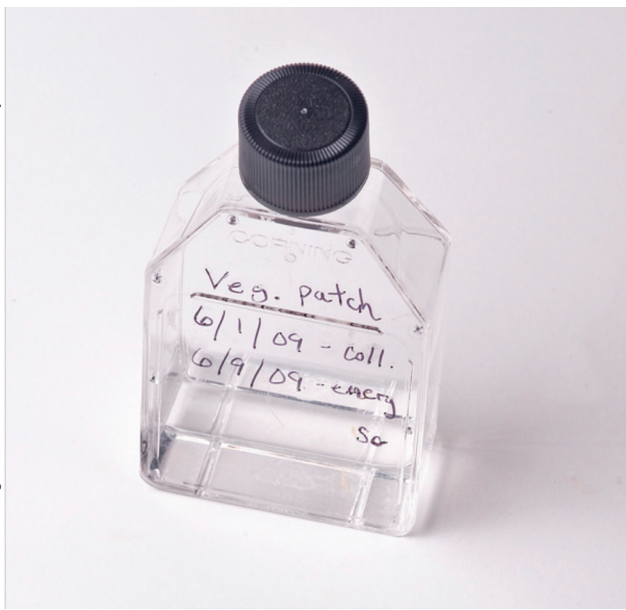


Figure 8. Newly emerged infective juveniles poured off the collecting dish may be stored in a water solution in a flask or bottle.

4. When IJ's begin to emerge, place the bottom petri dish into a larger "catch" dish that has a little water in it (Figure 7). For best results, very carefully remove the damp piece of paper towel containing the cadaver, flip the bottom dish upside down in the larger catch dish, and then place the piece of paper towel on the bottom/exposed surface of the flipped petri dish. Doing so will allow emerging infective juveniles to move directly into the water.
5. Nematodes will crawl into the water, where they may be poured off into a flask containing tap water that has been allowed to sit for at least 48 hours. The flask should be stored in the refrigerator with a loosened cap so nematodes have access to oxygen. The flask should not be filled completely, and nematodes should not be too concentrated in the flask (i.e., the water should not appear to be thick and cloudy). Use multiple flasks to accommodate these storage requirements. Be sure to label flasks with the same data on the covered dish from which the nematodes emerged (Figure 8).
6. Nematodes in solution may then be used to inoculate healthy waxworms by pouring a couple milliliters of the collected nematode solution over 5 to 10 waxworms which should be placed on a clean damp piece of paper towel in a clean pre-labeled petri dish. Repeat steps 1 to 5.
7. Nematodes in solution may be diluted and applied to the garden in a watering can.
8. Application of nematodes in solution should be done in the evening since EPN are sensitive to desiccation and intense solar radiation. Stored nematodes should be used within three weeks for optimal efficacy.

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