

Generation of a gene model for *Aedes aegypti* perilipin-1 gene and a probe to investigate number and size of the transcripts in mosquito tissues

John Adams

Biochemistry and Molecular Biology

Abstract

The mosquito *Aedes aegypti* is a known vector of disease for humans. Lipid metabolism is of key importance for survival and reproduction. Perilipin-1, PLIN1, is an important protein in the mobilization and storage of lipids in insects. Though the genome of *A. aegypti* has been sequenced, the gene structures must be confirmed by independent methods. Similarly, the nature of the transcripts from the gene must also be confirmed. From the available genomic and transcriptomic information, a cDNA clone was produced and used to obtain antibodies for western blot assays. Preliminary studies in which protein size was ascertained showed a protein much larger than was predicted by the reported sequences and the clones produced in the lab. Further studies which included cloning and sequencing led to the development of cDNA predicting a protein of the size found in the western blot. Using this information and the newly sequenced *Aedes albopictus* genome, a structural model of the PLIN1 gene has been constructed and a northern blot probe designed and produced to determine the number and size of the transcripts.

Introduction

Aedes aegypti is a vector associated with transmission of disease such as dengue, yellow fever, and chikungunya virus which affects millions of people annually¹. There are many possible targets of insecticides which may someday include agents which inhibit lipid metabolism. However, a thorough understanding of lipid metabolism must be ascertained before this can occur. Of critical importance to understanding this biological process is characterizing the proteins associated with it. One of these proteins is PLIN1, which may also be referred to as lipid storage droplet protein 1 (LSD1). This protein is found in association with cytosolic lipid droplets, which in insects are made up of large accumulations of triglycerides^{2,3}. Furthermore, triglycerides are the primary means by which fatty acids are stored. Fatty acids are the main source of energy for embryonic development and are a major component of eggs. Therefore, understanding the mechanisms by which fatty acids are stored and mobilized is of critical importance³. Previous studies conducted by the lab using the model insect *Manduca sexta* indicate that PLIN1 plays a central role in regulating fatty acid storage and mobilization. The genetic expression of PLIN1⁴ and the modulation of lipase activity via phosphorylation⁵ appear to be a critical factors in regulating the mobilization and storage of fatty acids. This protein has also been demonstrated to play a role in regulating fatty acid metabolism during wing development in *Drosophila melanogaster*⁶. The present study is part of effort directed to elucidate the function of PLIN1 in *Aedes aegypti*. To this end, an *Aedes aegypti* PLIN1 cDNA cloned in the lab was used to develop an RNA probe to determine the size and number of transcripts at various stages of the life cycle of *Aedes aegypti* by northern blot assays. We have

also generated a structural model of the PLIN1 gene and compared it to the gene in the recently sequenced *Aedes albopictus* genome.

Materials and Methods

Preliminary studies. According to vectorbase.org, which hosts the genome information of *Aedes aegypti*, the Lsd1 gene has 2 predicted transcripts. Lsd1 A (AAEL005951-RA) of length 714 base pairs and Lsd1 B (AAEL005951-RB) of length 1598 base pairs that encode proteins of 24kDa (211 aa) and 28kDa (245 aa), respectively (both transcripts are annotated as partial mRNA). At the time of these studies an additional transcript of 906 base pairs, AET-5405, encoding a protein of 34kDa (301 aa) protein was also reported in NCBI. This sequence was used to clone a cDNA of PLIN1 and express the recombinant protein in *E. coli*. The protein was then used to produce an antibody useful for western blotting. The immuno-blots of samples of *A. aegypti* fat body homogenates revealed a protein of 46kDa suggesting that AET-5405 was also a partial mRNA sequence. ESTs from NCBI were then used to search for missing regions of the previous sequences. This information was then used to design primers that would yield cDNA of the appropriate length. Cloning and sequencing with these primers yielded a cDNA product of 1548 base pairs with a predicted protein size of 46kDa. A multiple sequence alignment is shown in appendix.

PLIN1 Gene Model. The results of the cDNA sequencing were run in BLAST to find possible locations of exons using the Aag2 cell line genome assembly created by the Adino Lab at University of California, San Francisco and Pacific Biosciences (<https://www.vectorbase.org/organisms/aedes-aegypti/aag2/aag2>). *Aedes albopictus* is closely related to *A. aegypti* and shows a single transcript for Lsd1 AALF025165 of 1305 bp encoding a protein with a weight of 49 kDa. To compare the transcripts, the similarity of the proteins was ascertained using the ExPASy local alignment tool. In order to further determine similarity, the cDNA sequence was queried against the *A. aegypti* genome which was followed by a query of the *A. albopictus* PLIN1 transcript against the *A. albopictus* genome. A possible genetic map was then made using the hits from the cDNA query in which the approximate size of the exons was postulated using the size of the hits and the size of the introns were calculated as the difference in one hit and the next.

Preparation of the RNA Probe. To determine regions of overlap between the 1548bp cDNA, AET-5405, Lsd1 A, and Lsd1 B, a multiple alignment was constructed (Fig 1) using the clustal omega tool found on the European Bioinformatics institute (<http://www.ebi.ac.uk/>). This information was then used to design the primers needed to make the antisense northern blot RNA-probe of approximately 700bp. The following primers were designed: forward primer AELsd1C-F (5'-TGCCAGC-AGCAATCGGATGACTCTCATAAAACG-CAC-3') and the reverse primer AELsd1C-R (5' TAATACGACTCACTATAGGGTCCATTGGCGCTCATCG). The reverse primer contains a T7 promoter (5'-GTCCATTGGCGCTCATCG) to be used in the

generation of the antisense RNA probe. The location of the primers in the cDNA sequence is shown in Figure 1.

Results

Gene Structure. To determine the location of introns and exons in the PLIN1 gene, the 1548bp cDNA cloned in the lab was used to run a BLAST search against the genomic sequence of *A. aegypti* reported in vector base. The search revealed 7 exons (Table 1). The first exon contains only 4 coding bases and the 5'-UTR. Comparison of the *A. aegypti* and *A. albopictus* predicted proteins yielded 363 overlapping amino acids with 97.5% similarity (Fig 1). Since the sequence of PLIN1 from *Aedes albopictus* is almost identical to that from *A. aegypti*, we also compared the predicted gene structure of *A. aegypti* with that reported for *A. albopictus* (Table 1). The comparison shows that both species of *Aedes* have similar gene structures (seven exons of similar size) and, as expected, some differences in the length of introns and untranslated regions (not shown).

A sketch of the generated gene model of *A. aegypti* PLIN1 is shown in figure 2 with a model of *A. albopictus* in figure 3.

Table 1. Summary of cDNA search against *A. aegypti* genome and *A. albopictus* transcript against genome.

cDNA against <i>A. aegypti</i> genome	Alignment Length	% Identity	Location (Aag2 cell line genome sequence)	<i>A. albopictus</i> PLIN1 transcript against <i>A. albopictus</i> genome	Alignment Length	% Identity	Location (GCA_001876365.2 assembly)
Hit 1 3937249 5' ATG	138bp	97.8% Identity	3937115-3937252	Hit 1 5' ATG	54bp	100%	1008612-1008665
Hit 2	127bp	100%	3937448-3937574	Hit 2	127bp	99.2%	1008898-1009024
Hit 3	196bp	95.96%	3937646-3937843	Hit 3	191bp	95.3%	1009098-1009288
Hit 4	215bp	97.67%	3939175-3939389	Hit 4	209bp	97.1%	1019564-1019722
Hit 5	439bp	97.9%	3952495-3952935	Hit 5	447bp	97.5%	1020266-1020712
Hit 6	114bp	98.2%	3953034-3953147	Hit 6	116bp	97.4%	1034832-1034947
Hit 7 3953426 TAG 3'	332bp	97.3%	3953293-3953628	Hit 7 TGA 3'	229bp	99.1%	1035083-1035311

Figure 1. Local alignment of protein sequence of *A. aegypti* and *A. albopictus*

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>>Albopictus 434 bp (434 aa)

Waterman-Eggert score: 2205; 590.7 bits; E(1) < 2.7e-173
94.5% identity (97.5% similar) in 363 aa overlap (1-363:1-361)

      10      20      30      40      50      60
Aegypt MVHQKLRQNSGLPRMESISRVSIPVVETGLKTANTVYQKIKESNGLFNWGLETAEAIT
      .....
Albopi  MVHQKLRQNSGLPRMESISRFGSIPVVETGLKTANTVYHKVKESNGLFNWGLETAETIT
      10      20      30      40      50      60

      70      80      90      100     110     120
Aegypt YAFVDSLRPAAKLIIEGPLHRLDNFMCKSLDFVEQKVPSMYLPPEMMYWNTKEYMSDHLVK
      .....
Albopi  YAFVDSLRPAAKLIIEGPLHRLDNFMCKSLDFVEQKVPSMYLPPEMMYWNTKEYMSDRLVK
      70      80      90      100     110     120

      130     140     150     160     170     180
Aegypt PVLSRANSMKNLGHVVLESRVSNYAADRLDGALNVCDKYVDRYLPAEPEPAEDQTDSPNP
      .....
Albopi  PVLSRANSMKNLGHVVLESRVSNYAADRLDGALNVCDKYVDRYLPAE--PAGDCTDSLHP
      130     140     150     160     170

      190     200     210     220     230     240
Aegypt CQQQSDDSHKTHVIQTIHRGQLISRKLTRRLTFRTRQELTALKKQSTEAVHVVFYAAELI
      .....
Albopi  TQQQSDDSHKTHVIQTIHRGQQISRKLTRRLTFRTRQELTALKKQSTEAVHVVFYAAELI
      180     190     200     210     220     230

      250     260     270     280     290     300
Aegypt ATNPRLAMQKGVLEWQYLSADEPENQARPQTLEQLVLLTRESVRKVVHLINFTAGTVTK
      .....
Albopi  ATNPRLAMQKGVLEWQYLSADEPENQARPQTLEQLVLLTRESVRKVMVHLVNFTAGAVTK
      240     250     260     270     280     290

      310     320     330     340     350     360
Aegypt VPKTIRSQTRELLHHMMFATDRLIKA AHLENAKKATITEATGLMHRIQHTYEELQNQTNL
      .....
Albopi  VPRTIRVQTRELLHHMMFATDRLIKA AHLENAKNATITEATGLMHRIQHTYEELQNQTNL
      300     310     320     330     340     350

Aegypt ALE
      :.
Albopi ALQ
      360
```

Figure 2. Gene model of PLIN1 in *Aedes aegypti*

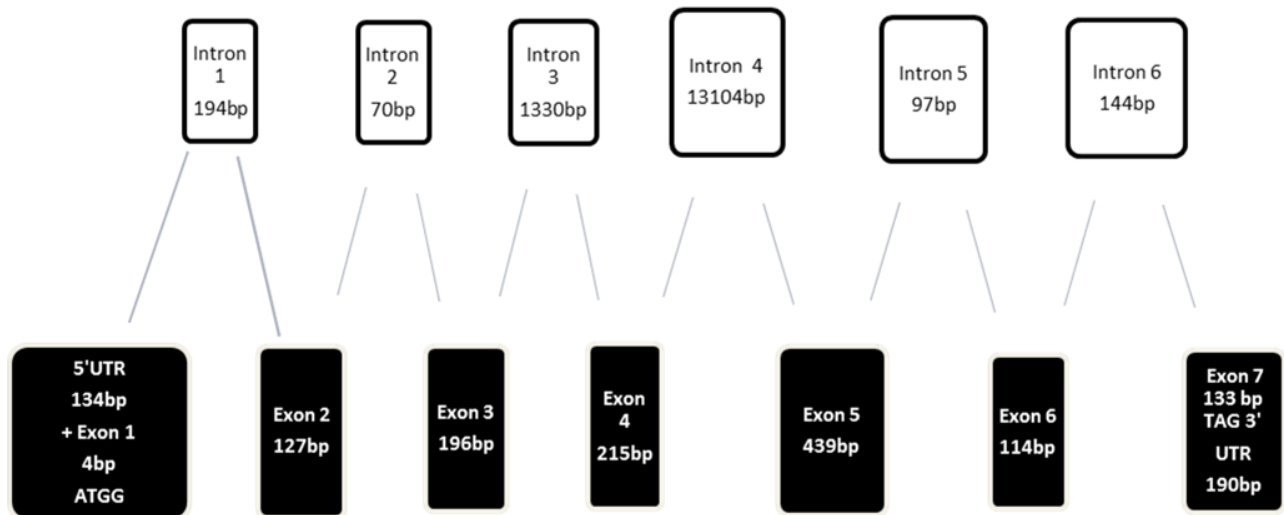
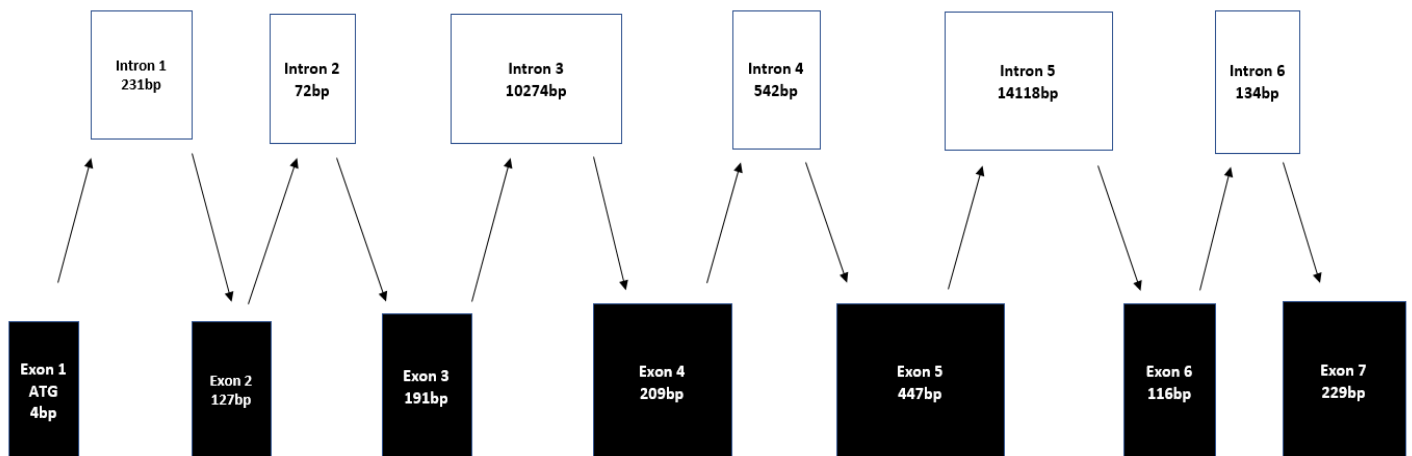


Figure 3. Gene model of PLIN1 in *Aedes albopictus*



Design and production of the anti-sense RNA probe. The sequence of the 1548 bp cDNA clone was used to obtain the RNA probe. The locations of the primers used to obtain the probe are highlighted in the figure 4. In order to generate sufficient quantities of cDNA (>20µg) to make the RNA probe, a total of 8 PCR reactions were conducted using the previously sequenced 1548bp cDNA as a template (Table 2). The PCR products were purified using an affinity column (Qiagen PCR Cleanup Kit) and the amount of purified PCR product obtained was estimated from the UV-absorption spectra. A picture of a representative agarose gel, used to confirm the presence and size of the PCR product is shown in figure 5. As expected we obtained a band of approximately 700 base pairs (Figure 5).

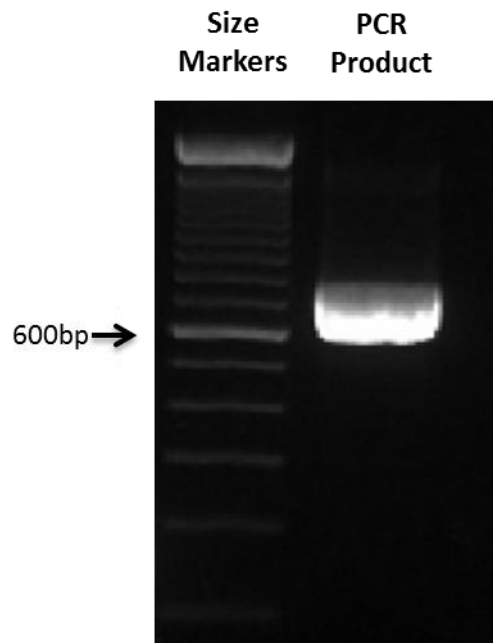
Figure 4. Aedes aegypti PLIN1 cDNA sequence. The locations of the forward and reverse primers used to obtain the RNA probe are highlighted in green and yellow.

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GATCGTTTTCTTTTTCCCGTGCGTTGAGCACTCAAAGTTTCCAATCAAGAGTTGTGTTTTCTCGTGCGTGTGTC
CCAAACCAGCTAAGGGTCATCGTCATTGTTCAAACAGCCAATACCTACGCTGCAACAATGGTGCATCAGAAGCTG
AAGCGCCAAAATTCGGGACTCCCGCGGATGGAATCCATATCGCGGGTCGGTAGCATTCCGGTCTGGAAACCGG
ACTGAAAACCGCCAACACGGTTTTATCAGAAGATTAAGGAAAGTAATGGATTGTTAACTGGGGACTCGAGACGG
CTGAAGCAATCACATACGCCTTCGTGGATTCACTGCGTCCGGCTGCTAAGCTGATCGAAGGACCGCTGCATCGTTT
GGATAATTTTATGTGTAAGGCTTGGACTTTGTGCAACAGAAAGGTCCTTCAATGTATCTTCTCCGGAGATGATGT
ACTGGAACACCAAGGAATACATGTCGGATCACCTGGTCAAGCCGGTGCTGAGCCGTGCCAACTCGATGAAGAAC
CTCGGTCATGTAGTGCTAGAGTCCCGAGTGCGAACTACGCAGCGGATCGTCTGGATGGGGCTTAAATGTGTGC
GATAAATATGTGGATCGATACCTCCCTGCGGAGCCAGAACCCGCCGAAGATCAAACCGATTACCTAATCCTTGCC
AGCAGCAATCGGATGACTCTATAAAACGCACGTTATCCAAACCATTACCGGGGACAATAATATCTCGCAAAC
GACACGTCGATTAACGTTTCGCACCCGTCAGGAATTGACCGCCTCAAGAAGCAGAGTACCGAAGCCGTACACGT
GGTGTCTACGCAGCCGAACTGATTGCCACCAACCCACGGCTGGCCATGCAGAAGGGCGTCAACTGTGGCAAT
ATCTTAGCGCTGACGAACCGGAAAACCAAGCCCGCCCCAAACGCTAGAACAACCTGGTGGTCTACTGACGAGA
GAATCCGTCCGTAAGGTTGTACATCTGATCAACTTACCGCTGGAACAGTAACGAAGGTCCCAAGACGATTGTT
CGAAACGCGCAATTGCTGCACCACATGATGTTTGCCACCGATCGATTGATAAAGGCTGCCACTTGGAACCG
CTAAGAAAGCAACGATTACCGAAGCGACTGGACTGATGCATCGAATTCAACACACCTATGAAGAAGTGCAAAATC
AAACCAATTTGGCTTTGGAACGTTAGCAGTGTTCTTTCCGGCCGCTGGAAGCGGAGAAGATCACCACCACCG
ACAATCCCCGGCGACGGATCCAAAACCGAGCGCACCATAATCCAATGCATACTAGCATCAATGGCGTTTATTAGCG
TACGAACGCGCGAGTTGCTGAACAGACGGATCCGCGAGGGCACCCCGCTCGAACCCGTAACGTTATTAAGAT
GAGCGCCAATGGACC AATGGACGTGAATGAGATGTTTTTTAAATAAGTTCTGAAGCATAAAACATTTGTGGTGT
AATAATGTTACTACTTGTGATAATAATAAATTTTCATATGATGTAC
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Table 2. Estimates of PCR product yields

	ng/ μ l	μ l	ng
PCR #1	62.6		
PCR #2	41.5	49	2035
PCR #3	25.6	49	1253
PCR #4	16.6	49	813
PCR #5	37.2	49	1821
PCR #6	37.3	49	1830
PCR #7	40.1	49	1963
PCR #8A	36.4	49	1786
PCR #8B	55.3	49	2709

Figure 5. Purity and size of PCR product. The PCR was analyzed by electrophoresis in an agarose gel, labelled with ethidium bromide and photographed under UV light. The left lane shows the used to make cDNA for the antisense RNA probe. 100 base pair ladder with dark band at 600 base pair



Discussion

The preliminary studies conducted involving immunoblotting indicate that the predicted transcripts from vectorbase and NCBI are incomplete. Comparison between the PLIN1 protein sequence predicted by the 1548 bp cDNA and that predicted by the *A. albopictus* genome were found to be highly similar. Further comparison between the predicted exons of the cDNA the *A. albopictus* exons supports the proposed genetic model. These findings also indicate that the genomic sequence for both *A. aegypti* and *A. albopictus* is correct.

The purified PCR product should be sufficient to produce enough anti-sense RNA probe for northern blot assays. However, this will remain uncertain until the northern blot has been conducted. These assays will be done in ovaries and fat body at various times of the *Aedes aegypti* life cycle to investigate the size and number of transcripts of PLIN1.

References

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Appendix

Multiple alignment of cDNA sequenced in the lab and the partial sequences, AET-5405, Lsd1A and Lsd1B. The primers for the cDNA are highlighted in green and yellow with the region covered by the probe highlighted in turquoise.

cDNA	GATCGTTTTCTTTTTTCCCGTGCCTTTGAGCACTCAAAGTTTCCAATCAAGAGTTGTGTT	60
DQ440406.1	-----	0
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0
cDNA	TTCTCGTGCCTGTTGTCCCAAACCAGCTAAGGGTCATCGTCATTGTTCAAACAGCCAATA	120
DQ440406.1	-----	0
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0
cDNA	CCTACGCTGCAACAATGGTGCATCAGAAGCTGAAGCGCCAAAATTCGGGACTCCCGCGGA	180
DQ440406.1	-----	0
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0
cDNA	TGGAATCCATATCGCGGGTCGGTAGCATTCCGGTCGTGGAAACCGGACTGAAAACCGCCA	240
DQ440406.1	-----	0
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0
cDNA	ACACGGTTTATCAGAAGATTAAGGAAAGTAATGGATTGTTTAACTGGGACTCGAGACGG	300
DQ440406.1	-----	0
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0
cDNA	CTGAAGCAATCACATACGCCTTCGTGGATTCCTGCGTCCGGCTGCTAAGCTGATCGAAG	360
DQ440406.1	-----	0
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0
cDNA	GACCGCTGCATCGTTTGGATAATTTTATGTGTAAAAGCTTGGACTTTGTCGAACAGAAGG	420
DQ440406.1	-----	0
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0
cDNA	TCCCTTCAATGTATCTTCCCTCCGGAGATGATGTACTGGAACACCAAGGAATACATGTCCG	480
DQ440406.1	-----ATGATGTGCTGGAACACCAAGGAATACATGTCCG	34
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0
cDNA	ATCACCTGGTCAAGCCGGTGTGAGCCGTGCCAACTCGATGAAGAACCCTCGGTCATGTAG	540
DQ440406.1	ATCACCTGGTCAAGCCGGTGTGAGCCGTGCCAACTCGATGAAGAACCCTCGGTCATGTAG	94
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0
cDNA	TGCTAGAGTCCCGAGTGTGCGAACTACGCAGCGGATCGTCTGGATGGGGCTTTAAATGTGT	600
DQ440406.1	TGCTAGAGTCCCGAGTGTGCGAACTACGCAGCGGATCGTCTGGATGGGGCTTTAAATGTGT	154
AAEL005951-RA	-----	0
AAEL005951-RB	-----	0

cDNA	CGGATAAATATGTGGATCGATACCTCCCTGCGGAGCCAGAACCCGCCGAAGATCAAACCG	660
DQ440406.1	CGGATAAATATGTGGATCGATACCTCCCTGCGGAGCCAGAACCCGCCGAAGATCAAACCG	214
AAEL005951-RA	-----ATGGCACACGGATCAAAAAGCCAAAAGCAGCAACTTCACAAGGCT	46
AAEL005951-RB	-----ATGGCACACGGATCAAAAAGCCAAAAGCAGCAACTTCACAAGGCT	46
	* * * * *	
cDNA	ATTACCTAATCCTT CGCCAGCAGCAATCGGATGACTCTCATAAAAACGCAG GTTATCCAAA	720
DQ440406.1	ATTACCTAATCCTT CGCCAGCAGCAATCGGATGACTCTCATAAAAACGCAG GTTATCCAAA	274
AAEL005951-RA	CGGCACCTAATCCTT CGCCAGCAGCAATCGGATGACTCTCATAAAAACGCAG GTTATCCAAA	106
AAEL005951-RB	CGGCACCTAATCCTT CGCCAGCAGCAATCGGATGACTCTCATAAAAACGCAG GTTATCCAAA	106

cDNA	CCATTCACCGGGGACAACCTAATATCTCGCAAACCTGACACGTCGATTAACGTTTCGCACCC	780
DQ440406.1	CCATTCACCGGGGACAACCTAATATCTCGCAAACCTGACACGTCGATTAACGTTTCGCACCC	334
AAEL005951-RA	CCATTCACCGGGGACAACCTAATATCTCGCAAACCTGACACGTCGATTAACGTTTCGCACCC	166
AAEL005951-RB	CCATTCACCGGGGACAACCTAATATCTCGCAAACCTGACACGTCGATTAACGTTTCGCACCC	166

cDNA	GTCAGGAATTGACCGCCCTCAAGAAGCAGAGTACCGAAGCCGTACACGTTGGTGTCTACG	840
DQ440406.1	GTCAGGAATTGACCGCCCTCAAGAAGCAGAGTACCGAAGCCGTACACGTTGGTGTCTACG	394
AAEL005951-RA	GTCAGGAATTGACCGCCCTCAAGAAGCAGAGTACCGAAGCCGTACACGTTGGTGTCTACG	226
AAEL005951-RB	GTCAGGAATTGACCGCCCTCAAGAAGCAGAGTACCGAAGCCGTACACGTTGGTGTCTACG	226

cDNA	CAGCCGAACCTGATTGCCACCAACCCACGGCTGGCCATGCAGAAGGGCGTCGAACTGTGGC	900
DQ440406.1	CAGCCGAACCTGATTGCCACCAACCCACGGCTGGCCATGCAGAAGGGCGTCGAACTGTGGC	454
AAEL005951-RA	CAGCCGAACCTGATTGCCACCAACCCACGGCTGGCCATGCAGAAGGGCGTCGAACTGTGGC	286
AAEL005951-RB	CAGCCGAACCTGATTGCCACCAACCCACGGCTGGCCATGCAGAAGGGCGTCGAACTGTGGC	286

cDNA	AATATCTTAGCGCTGACGAAACCGGAAAACCAAGCCCGCCCAACCGCTAGAACAACCTGG	960
DQ440406.1	AATATCTTAGCGCTGACGAAACCGGAAAACCAAGCCCGCCCAACCGCTAGAACAACCTGG	514
AAEL005951-RA	AATATCTTAGCGCTGACGAAACCGGAAAACCAAGCCCGCCCAACCGCTAGAACAACCTGG	346
AAEL005951-RB	AATATCTTAGCGCTGACGAAACCGGAAAACCAAGCCCGCCCAACCGCTAGAACAACCTGG	346

cDNA	TGGTCTACTGACGAGAGAATCCGTCGGTAAGGTTGTACATCTGATCAACTTCACCGCTG	1020
DQ440406.1	TGGTCTACTGACGAGAGAATCCGTCGGTAAGGTTGTACATCTGATCAACTTCACCGCTG	574
AAEL005951-RA	TGGTCTACTGACGAGAGAATCCGTCGGTAAGGTTGTACATCTGATCAACTTCACCGCTG	406
AAEL005951-RB	TGGTCTACTGACGAGAGAATCCGTCGGTAAGGTTGTACATCTGATCAACTTCACCGCTG	406

cDNA	GAACAGTAACGAAGGTCCCAAGACGATTTCGTTTCGCAAACCGCGGAATTGCTGCACCACA	1080
DQ440406.1	GAACAGTAACGAAGGTCCCAAGACGATTTCGTTTCGCAAACCGCGGAATTGCTGCACCACA	634
AAEL005951-RA	GAACAGTAACGAAGGTCCCAAGACGATTTCGTTTCGCAAACCGCGGAATTGCTGCACCACA	466
AAEL005951-RB	GAACAGTAACGAAGGTCCCAAGACGATTTCGTTTCGCAAACCGCGGAATTGCTGCACCACA	466

cDNA	TGATGTTTGGCCACCGATCGATTGATAAAGGCTGCCCACTTGGAACCGCTAAGAAAGCAA	1140
DQ440406.1	TGATGTTTGGCCACCGATCGATTGATAAAGGCTGCCCACTTGGAACCGCTAAGAAAGCAA	694
AAEL005951-RA	TGATGTTTGGCCACCGATCGATTGATAAAGGCTGCCCACTTGGAACCGCTAAGAAAGCAA	526
AAEL005951-RB	TGATGTTTGGCCACCGATCGATTGATAAAGGCTGCCCACTTGGAACCGCTAAGAAAGCAA	526

cDNA	CGATTACCGAAGCGACTGGACTGATGCATCGAATTCACACACCTATGAAGAAGTGCAAA	1200
DQ440406.1	CGATTACCGAAGCGACTGGACTGATGCATCGAATTCACACACCTATGAAGAAGTGCAAA	754
AAEL005951-RA	CGATTACCGAAGCGACTGGACTGATGCATCGAATTCACACACCTATGAAGAAGTGCAAA	586
AAEL005951-RB	CGATTACCGAAGCGACTGGACTGATGCATCGAATTCACACACCTATGAAGAAGTGCAAA	586

cDNA	ATCAAACCAATTTGGCTTTGGAACGTTTAGCAGTGTTCCTTTCCGGCCGCTGGAAGCGG	1260
DQ440406.1	ATCAAACCAATTTGGCTTTGGAACGTTTAGCAGTGTTCCTTTCCGGCCGCTGGAAGCGG	814
AAEL005951-RA	ATCAAACCAATTTGGCTTTGGAACGTTTAGCAGTGTTCCTTTCCGGCCGCTGGAAGCGG	646
AAEL005951-RB	ATCAAACCAATTTGGCTTTGGAACGTTTAGCAGTGTTCCTTTCCGGCCGCTGGAAGCGG	646
	***** * * * * *	
cDNA	AGAAGATCACCAACCGACAATCC-CCGGCAGCGGATCCAAAACCGAGCGCACCATAAT	1319
DQ440406.1	AGAAGATCACCAACCGACAATCC-CCGGCAGCGGATCCAAAACCGAGCGCACCATAAT	873
AAEL005951-RA	CAT--TAAACCATCATAAAAAATATAATGCCAAAAGTGTCAATGCTAATGCATGTTAA-	703

AAEL005951-RB	AGAAGATCACCACCACCGACAATCC-CCGGCGACGGATCCAAAACCGAGCGCACCATAAT ***** ** * ** * ** * ** * ** * ** *	705
cDNA	CCAATGCATACTAGCATCAATGGCGTTTTATTAGCGTACGAACGGCCGAGTTGCTGAACAG	1379
DQ440406.1	CCAATGCATACTAGCATCAATGGCGTTTTATTAG-----	906
AAEL005951-RA	-----ATTAAATTTA-----	714
AAEL005951-RB	CCAATGCATACTAGCATCAATGGCGTTTTATTAGCGTACGAACGGCCGAGTTGCTGAACAG * * * * *	765
cDNA	ACGGATCCGCGAGGGCACCCTCGAACCCGTACTCAACGTCGATGAGCGCCAATGGAC	1439
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	ACGGATCCGCGAGGGCACCCTCGAACCCGTACTCAACGTCGATGAGCGCCAATGGAC	825
cDNA	CAATGGACGTGAATGAGAT----GTTTTTTTAAAATAAGTTCTGAAGCATAAAACATTTG	1495
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	CAATGGACGTGAATGAGATATATGTTTTTTTATAATAAGTTCTGAAGCATAAAACATTTG	885
cDNA	TGGTGAATAATGTTACTACTTGTGATAAATAATAAATTCATATGATGTAC-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	TGGTGAATAATGTTACTACTTGTGATAAATAATAAATTCATATGATGTACAAAATA	945
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	GTCTTAACTAATCTCAATTGATTGAGAACTCTATCTCTTCTTCTTAAATCATCGGTGTCC	1005
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	AAATGGGTGGTCCCCCGAATCCGACGGTCGAGTAAGAAACACATTCGAAGCAAACCTCT	1065
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	CATCGATTTGCTCGCATTTGCCAAATCGGAAATGTTGTCACATCTTGGGATTCGGTTC	1125
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	CAGCAGAAGATTCTCTGATTCCGCTTGCAAGGTCAGCTTCTCCAACGGAATTCACACGG	1185
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	GTAGGTCTTCTTCGGCAATCTGGGGCATTTCTAAACCAAGATCGGCGTAAACCTCACTCA	1245
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	ACCGGGTGATATTTTATCGGCAGTTTCATCGGCTTCGCCAGCAGACGTTTCAGCTGTT	1305
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	CAGTGTCCGGACGAACCTCCGCGCTTTTTCCTCGATGTCCTTCAAGGAGTTTTGGTTGT	1365
cDNA	-----	1548
DQ440406.1	-----	906

AAEL005951-RA	-----	714
AAEL005951-RB	CGGCGATGAAGCGTCCGGCTTGATCAAGTTTGTGGGGTTTTGCTGCACGAAGCCACACG	1425
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	CGAATTGGGCAGTTTTTTTGCCTAGTAGTAAGCCCAGCTTGGTGTCTACTTCGTAGGCATGTT	1485
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	CCTTAGCGGTACGGAACATTTTCAATCCCAGGAACAGAACTTAAAACGTGCGATCTGGAT	1545
cDNA	-----	1548
DQ440406.1	-----	906
AAEL005951-RA	-----	714
AAEL005951-RB	TTCACCGATTTCAACGTAAATGTTTACTGGAACGATCAAATCCAAACTGCGC	1598