

A DIALLEL CROSS ANALYSIS OF FIBER STRENGTH  
IN FOUR VARIETIES OF UPLAND COTTON

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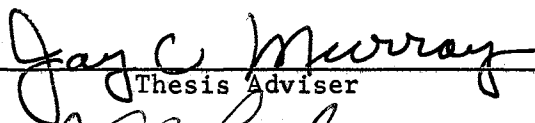
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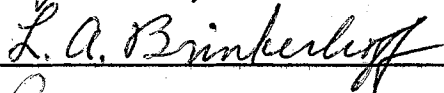
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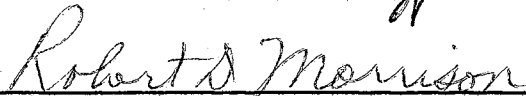
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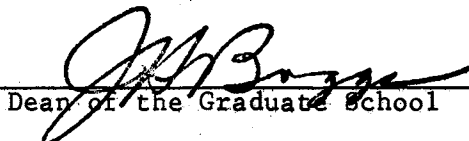
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## TABLE OF CONTENTS

Chapter	Page
I. INTRODUCTION . . . . .	1
II. REVIEW OF LITERATURE . . . . .	2
The Inheritance of Fiber Strength . . . . .	2
Heterosis and Combining Ability . . . . .	3
Environmental Effects . . . . .	4
III. MATERIALS AND METHODS . . . . .	7
Varieties . . . . .	7
Experimental Procedure . . . . .	7
Statistical Procedure . . . . .	9
A. Analysis of Variance . . . . .	9
B. Test of Heterosis and Inbreeding Depression . . . . .	9
C. Diallel Cross Analysis . . . . .	9
D. Diallel Cross Graph . . . . .	16
E. Test of Non-allelic Interactions . . . . .	18
IV. RESULTS AND DISCUSSION . . . . .	20
Analysis of Variance . . . . .	20
Heterosis and Inbreeding Depression . . . . .	22
Diallel Cross Graph . . . . .	23
Non-allelic Interactions . . . . .	27
Diallel Cross Analysis . . . . .	28
Discussion . . . . .	35
V. SUMMARY AND CONCLUSIONS . . . . .	39
LITERATURE CITED . . . . .	41

LIST OF TABLES

Table	Page
I.	Number of Subsamples . . . . . 21
II.	Analysis of Variance . . . . . 22
III.	Means of Parents ( $P_1$ , $P_2$ ), Mid-parent (MP), $F_1$ , $F_2$ , ( $F_1 - MP$ ) and ( $F_2 - F_1$ ) of Individual Crosses, in Grams/Grex . . . . . 23
IV.	Means of Parents and $F_1$ , in Grams/Grex . . . . . 24
V.	Means of Parents and $F_2$ , in Grams/Grex . . . . . 24
VI.	Estimates of $V_p$ , $V_r$ , and $W_r$ in the $F_1$ and $F_2$ generations . . 25
VII.	The $2L_2 - L_1$ Table . . . . . 27
VIII.	Means of Parents and $F_1$ in Replicate 2 and Replicate 4, in Grams/Grex . . . . . 29
IX.	Means of Parents and $F_2$ in Replicate 2 and Replicate 4, in Grams/Grex . . . . . 29
X.	Means of Parents and $B_1$ in Replicate 2 and Replicate 4, in Grams/Grex . . . . . 30
XI.	Estimates of the Second Degree Statistics, Used for the Estimation of the Diallel Cross Parameters, of $F_1$ , $F_2$ , and $B_1$ generations . . . . . 31
XII.	Estimates of Environmental Variances . . . . . 31
XIII.	Estimates of the Diallel Cross Parameters, Their Means, and Standard Errors . . . . . 33

LIST OF FIGURES

Figure		Page
1.	The Regression Lines of $W_r$ on $V_r$ with Varying Degrees of Dominance. After Dickinson and Jinks (7) . . . . .	17
2.	The $(V_r, W_r)$ Graph of the $F_1$ Family . . . . .	26
3.	The $(V_r, W_r)$ Graph of the $F_2$ Family . . . . .	26

LIST OF APPENDIX

Appendix		Page
A.	Regression Analysis of $W_r$ on $V_r$ on the $F_1$ Family . . . . .	45
B.	Regression Analysis of $W_r$ on $V_r$ of the $F_2$ Family . . . . .	46

## CHAPTER I

### INTRODUCTION

Cotton technologists have long recognized that the strength of raw lint cotton plays an important part in the production of high quality fabrics. Since there are varieties, which consistently produce fibers with relatively high tensile strength, it appears that the strength of the fiber of commercial cottons can be improved through breeding proper stocks.

In order to devise a breeding program for improving fiber strength, it is necessary to understand the inheritance, the heritability, and the nature of gene action governing this trait. Relatively little pertinent information concerning these subjects is available, hence, results of many more studies involving different groups of material and different methods of experimentation need to be accumulated to get a clearer picture of these phenomena.

It is the purpose of this study to provide some additional information that might contribute to a better understanding of the problem. Specifically, the objectives of this study are to obtain estimates of the diallel cross components of variance and other estimators derived therefrom, with respect to fiber strength, from a diallel cross of four commercial varieties of upland cotton grown in Oklahoma. Their breeding implications are also discussed.



## CHAPTER II

### REVIEW OF LITERATURE

#### The Inheritance of Fiber Strength

There is relatively little information available on the inheritance of fiber strength in upland cotton, Gossypium hirsutum, L. However, it has been fairly well established by early workers that fiber strength was a quantitative character being governed by a large number of genes.

Ware and Harrell (49) studied the  $F_1$ ,  $F_2$ ,  $F_3$ , and backcross generations of a cross between Florida Green Seed x Rowden, two stocks of upland cotton, and found that fiber strength was a quantitative character and there was slight dominance of the weak fiber.

From the data with crosses involving two varieties of upland cotton, AHA-50 x Half and Half, Self and Henderson (42) concluded that fiber strength was a quantitative trait probably governed by 4 or 5 pairs of gene. They also found that the heritability estimates for this trait were rather high.

In a rather indirect study, Ware (48) found that the X-ray diffraction patterns and the structure of cotton fiber were quantitative characters. Since there was a highly significant linear correlation between the diffraction angles and the strength of the fiber, fiber strength was also a quantitative character.

Tipton et al. (45) found in a recent study of two separate crosses of upland cotton strains, Cleveland Short Sympodia x Stardell 6-6, and Cleveland Short Sympodia x AHA-6-1-4, that fiber strength was a quantitative character presumably governed by 12 or 13 pairs of gene. They also found partial dominance of the low fiber strength and a high heritability estimate.

#### Heterosis and Combining Ability

Considerable interest exists among cotton breeders and geneticists on the expression of heterosis in upland cotton. Loden and Richmond (27) reviewed early works on heterosis in inter- as well as intra-specific crosses of cotton, pointing out findings ranging from no or little heterotic effects for certain characters in certain crosses to rather substantial in others.

More recent investigations on different characters (25, 32, 44, 46, 47, 50, 51) in crosses involving upland cotton varieties likewise produced variable results. Generally, in a group of materials, certain crosses exhibit substantial heterosis for certain characters as measured by comparing the  $F_1$  with the average performance of the parental lines. However, when such  $F_1$ 's are compared with the best of the parents, there are very few instances of significantly superior performances of the hybrids. Heterosis in cotton seems to be of primary importance only in yield, boll number, and boll size (32). Fiber traits occasionally showed small heterotic effects, but were generally intermediate between the parents.

There is rather meager information available on the relative magnitude of general and specific combining abilities for fiber characters

in upland cotton. Barnes and Staten (3) studied a diallel cross of seven western Acala strains, and found that specific combining ability was more important than general combining ability for fiber traits. However, from another set of diallel crosses involving south-eastern varieties, no specific combining ability was apparent. From a diallel cross among primitive and foreign strains of upland cotton, White and Richmond (51) reported a preponderance of general combining ability variance for fiber traits as compared to the specific combining ability variance.

From a diallel cross study of eight inbred lines of upland cotton, Miller and Marani (32) found a significant general combining ability variance for fiber strength and a non-significant specific combining ability variance.

Matzinger and Kempthorne (31) have shown that the general and specific combining ability variances are related to the nature of the gene action of the trait in the population under study. They reported that significant general combining ability suggests primarily additive gene action, whereas significant specific combining ability indicates the presence of dominant and epistatic effects of the character in question.

From the results of Miller and Marani (32) and of White and Richmond (51), fiber strength appears to be governed primarily by additive gene action, although the presence of heterotic effects (32) indicated the presence of dominant and epistatic effects.

#### Environmental Effects

Environment plays a rather important role in modifying the fiber characters, presumably due to the long maturity period of cotton fiber

(11).

Hancock (10) reported that, in general, dry and warm years tend to produce stronger and shorter fibers than do cooler and wetter seasons. However, during years of uniform rainfall distribution, the fibers tended to be both long and strong.

Pope and Ware (41) reported that fiber strength seems to be more affected by seasonal variation than do other characters of the fiber. Fiber strength appears to be significantly correlated with almost all climatic factors (12). Hesler, Lane, and Young (19) reported that the fiber development might be retarded if temperature is the limiting factor.

Spooner et al. (43) found that irrigation significantly increased fiber length, but did not significantly affect fiber strength, although there was a tendency toward lowering of strength at relatively high irrigation level.

Peebles, Den Hartog and Pressley (40) reported that somewhat weaker fibers were produced by close spacing, but the mean deficiency induced was very small and non-significant.

Limited information is available on the effect of fertilizer treatments on fiber traits. MacKenzie and van Schaik (28) found that nitrogen treatment did not affect significantly the fiber characters. Recently, Murray, Reed, and Oswalt (37) reported that various nitrogen, phosphorus, and potassium fertilizer treatments resulted in no significant differences in fiber traits.

Miller and his co-workers (33, 34, 35) have reported that fiber traits showed first and second order interactions between variety, location and year. However, in many cases the interaction components were very small relative to the variety variance component, and can be

considered unimportant.

In conclusion, fiber strength appears to be a quantitatively inherited character, governed by a large number of genes. Weak fiber appears to be partially dominant, and the heritability estimates are rather high. Presumably, the nature of gene action is primarily additive with small dominant and epistatic effects. Environment plays a part in modifying the phenotypic expression of this trait. The interactions between variety, location and year, in many cases, are small and can be considered unimportant in comparison to the varietal source of variation.

## CHAPTER III

### MATERIALS AND METHODS

#### Varieties

Four varieties of commercial upland cotton commonly grown in Oklahoma were included in this study, namely, Acala 4-42, Gregg, Lankart 57, and Stoneville 62. A short description of these varieties, as given by Brown and Ware (4), is as follows.

Acala 4-42 was developed by selection from the same general material as Acala 1517, which in turn was originally selected from Young's Acala.

Gregg was developed by selection from Macha, the latter was selected from Half and Half.

Lankart 57 is the most commonly grown variety in Oklahoma. It was developed from a Texas stormproof cotton called Lone Star.

Stoneville 62 was developed from Stoneville 2B by the Oklahoma Agricultural Experimental Station.

Under Oklahoma conditions, Acala 4-42 and Gregg produce relatively high strength fibers, while Stoneville 62 and Lankart 57 produce relatively weak fibers (36).

#### Experimental Procedure

All possible crosses of the four varieties were made during the summer of 1963 from a random sample of plants of each variety grown in

the field. These varieties can be considered "pure" and had been bulked for several years before. The seeds of the reciprocal crosses of any two parents were mixed together. The parental and  $F_1$  seeds were then grown in the greenhouse in rows of 10 to 12 plants each during the winter of 1963. The parental plants were selfed and crossed in all possible combinations to obtain selfed parental and  $F_1$  seeds. The  $F_1$  plants were also selfed and backcrossed to both parents in order to produce  $F_2$  and backcross seeds.

The experiment consisted of 28 entries, namely 4 parental, 6  $F_1$ , 6  $F_2$ , 6  $B_1$ , and 6  $B_2$  generations. They were grown in the summer of 1964.

The design of the experiment was a randomized complete block with 4 replications. Each entry was grown in a single row plot of 25 feet length. The spacing between the plants in the row was 24 inches, and the rows were 40 inches apart from each other. The plants in replicates 1 and 3 were directly sown from seeds, whereas those in replicates 2 and 4 were transplanted seedlings.

The plants were cultivated in the usual practice; irrigations were applied as needed; and the plants were sprayed for insect control.

Due to the differences in maturity among the plants in the populations, the plants were harvested twice during the season. Samples of 10 bolls were taken from plants which had 10 or more bolls. The seed cotton was ginned with a saw gin, and the lint was analyzed for their fiber properties. The strength of the fiber reported in this study was measured in  $T_0$  unit, namely, the strength of a bundle of fibers measured on a Stelometer with two jaws holding the bundle tightly appressed. The strength is expressed in grams/grex.

## Statistical Procedure

A. Analysis of Variance. Analysis of variance was conducted to test the differences in fiber strength among the entries, and to provide an estimate of error variance, which will be used in testing the presence of heterosis and inbreeding depression.

B. Test of Heterosis and Inbreeding Depression. To test the presence of significant heterotic effect the mean of the  $F_1$  of each cross was compared with the mean of each of its mid-parents (MP) value. By assuming that the mid-parents values are normally distributed and have equal error variance as the  $F_1$ , t-tests were conducted on each of the mean of  $(F_1 - MP)$  values.

Inbreeding depression was tested by comparing the mean of  $F_2$  and that of  $F_1$  of each cross. Similarly, the error variances of  $F_1$  and  $F_2$  values are assumed to be equal, and t-tests were conducted on each of the mean of  $(F_2 - F_1)$  values.

In addition, the direction of the average degree of dominance in the parents was determined from the sign of the mean of  $(F_1 - MP)$  values. On the assumption that these differences are normally distributed, the confidence limits of the mean of  $(F_1 - MP)$  were set.

C. Diallel Cross Analysis. The diallel cross analysis employed in this study is the one developed by Jinks and Hayman (23) and later elaborated by Hayman (13, 14, 15, 16) and Jinks (20, 21, 22), and recently used and discussed by Crumpacker and Allard (6), Johnson (24), White and Kohel (50), and Whitehouse, Thompson and Do Valle Ribeiro (52).

Hayman's model is based on several assumptions, each of which must hold true for the analysis to be valid. These assumptions are (6):



1. Homozygous parents
2. Diploid segregation
3. No reciprocal differences
4. No genotype - environment interaction
5. No non-allelic gene interaction
6. No multiple alleles
7. Uncorrelated gene distributions

The first four assumptions appear to be valid for G. hirsutum.

G. hirsutum can be considered as a mostly-self-pollinated crop. With controlled pollination, a homozygous condition can be more assured. The parental plants used in this study are samples of rather "pure" varieties, which had been bulked for several years, and selfed for one generation. Consequently, the parental plants are considered to be fairly homozygous.

Although G. hirsutum is an amphidiploid, according to Endrizzi (9) and Kimber (26), it segregates in a diploidal manner.

For most characters of cotton, many authors consider that there are no differences between reciprocal crosses. Recently, White and Richmond (51) reported no significant differences between reciprocal crosses for fiber strength and other fiber properties.

The fifth assumption can be tested using the method given by Hayman (14), which are presented later in this chapter.

The last two assumptions, in fact all of these assumptions, can be tested by the diallel cross or  $(V_r, W_r)$  graph (14, 20, 23). This test is presented later in this chapter.

When these assumptions are valid, the contribution of each locus to the family means in a diallel cross can be described in terms of the

genetic parameters  $\underline{u}$ ,  $\underline{v}$ ,  $\underline{d}$ , and  $\underline{h}$  (23, 29), where:

$u$  and  $v$  = respectively, frequencies of positive and negative alleles.

$$0 \leq u, v, \leq 1; u + v = 1$$

$$d = \text{additive effect. } 0 \leq d \leq c_1$$

$$h = \text{dominant effect. } c_2 \leq h \leq c_3; c_2 \leq 0, c_3 \geq 0$$

$c_1, c_2$  and  $c_3$  = constant values (for each gene).

Then a number of statistics can be calculated from the parental,  $F_1$ ,  $F_2$ , and backcross family means. For a number of genes, say  $k$ , the genetic contents of certain statistics, in terms of the genetic parameters, from the parental and  $F_1$  family are as follows (13, 23):

$$\text{Var.}^1 \text{ of parents} = V_p = V_{OLO} = 4 \sum u_i v_i d_i^2 \quad (a)$$

$$\text{Var. of the } r^{\text{th}} \text{ array}^2 = V_r = \sum_{i=1}^j u_i v_i (d_i - h_i)^2 + \sum_{j+1}^k u_i v_i (d_i + h_i)^2 \quad (b)$$

$$\text{Mean var. of arrays} = V_{1L1} = \sum \left[ u_i v_i \left\{ d_i^2 + h_i^2 - 2 d_i h_i (u_i - v_i) \right\} \right] \quad (c)$$

$$\text{Var. of array means} = V_{OL1} = \sum \left[ u_i v_i \left\{ d_i - h_i (u_i - v_i) \right\}^2 \right] \quad (e)$$

$$\text{Covar.}^3 \text{ of the } r^{\text{th}} \text{ array} = W_r = 2 \sum_{i=1}^j u_i v_i d_i (d_i - h_i) + 2 \sum_{j+1}^k u_i v_i d_i (d_i + h_i) \quad (f)$$

$$\text{Mean covar. of arrays} = W_{OLO1} = 2 \sum \left[ u_i v_i d_i \left\{ d_i - (u_i - v_i) h_i \right\} \right] \quad (g)$$

$i = 1, 2, \dots, k$ ;  $j$  = the number of negative alleles;  $j < k$

<sup>1</sup>Var. = variance.

<sup>2</sup>Array. = All crosses which have a common parent, including the parent itself.

<sup>3</sup>Covar. = Covariance of the  $r^{\text{th}}$  array and their non-recurrent parents.

The subscript L refers to the diallel cross mating system, and the subsequent number(s), beginning from 0 for the parents, refer to the generations under consideration. In variances of individual measurements, the number preceding L is the same as the one following, whereas in variances of means and in covariances, the number(s) preceding L refers to the generation(s) of the common parents. The summations are over all values of  $i$ , except for  $V_r$  and  $W_r$ .

Jinks and Hayman (23) defined the diallel crosses components of variance or the diallel cross parameters as:

$$D = 4 \sum u_i v_i d_i^2 \quad (p)$$

$$H_1 = 4 \sum u_i v_i h_i^2 \quad (q)$$

$$H_2 = 16 \sum u_i^2 v_i^2 h_i^2 \quad (r)$$

$$F = 8 \sum [u_i v_i (u_i - v_i) d_i h_i] \quad (s)$$

Taking into account the environmental variances, the expectations of the variances and the covariances of the family means, in terms of the diallel cross components of variance, are as follows:

For parental and  $F_1$  families (13, 20):

$$V_{OL0} = D + E_0$$

$$V_{OL1} = \frac{1}{4}D + \frac{1}{4}H_1 - \frac{1}{4}H_2 - \frac{1}{4}F + E_0 + (n - 2)E_1/n^2$$

$$V_{1L1} = \frac{1}{4}D + \frac{1}{4}H_1 - \frac{1}{4}F + E_0 + (n - 1)E_1/n$$

$$W_{OL01} = \frac{1}{2}D - \frac{1}{4}F + E_0/n$$

where:  $E_0$  and  $E_1$  are the environmental variances for the parental and  $F_1$  families.

$n$  is the number of parents in the diallel cross.

For parental and  $F_2$  families (14, 22):

$$V_{OL2} = \frac{1}{4}D + (1/16)H_1 - (1/16)H_2 - (1/8)F + E_0 + (n - 2)E_2/n^2$$

$$V_{1L2} = \frac{1}{4}D + (1/16)H_1 - (1/8)F + E_0 + (n - 1)E_2/n$$

$$V_{2L2} = \frac{1}{4}D + (1/8)H_1 + E_2$$

$$W_{0L02} = \frac{1}{2}D - (1/8)F + E_0/n$$

where:  $E_2$  is the environmental variance for the  $F_2$  family, and  $E_0$ ,  $E_1$ , and  $n$  are as defined above.

For the parental and backcross populations (22):

In the absence of non-allelic interactions, there exists a relationship between the  $F_2$  family mean and the means of the two reciprocal backcross families, namely  $\bar{F}_2 = \frac{1}{2}(\bar{B}_1 + \bar{B}_2)$ . Thus, in the absence of non-allelic interactions, the expected statistics for the means of the reciprocal backcross families are identical with those for the  $F_2$  family means (22).

Estimates for each set of these statistics can be calculated from the tables consisting of the means of the parents and  $F_1$ 's, the means of the parents and  $F_2$ 's, and the means of the parents and the average of the reciprocal backcrosses. These three tables will, respectively, be called the  $L_1$ ,  $L_2$ , and  $B_1$  tables.

Since the estimates of environmental variances can be obtained from replicated experiments, the estimates of the diallel cross parameters can be solved from each set of equations given previously. These equations can be solved by a method given by Mather (29) and Hayman (13). The normal equations of the diallel cross parameters are obtained and their coefficients are set in a square matrix that is symmetrical about the main diagonal. This matrix is then inverted to obtain the covariance matrix. The latter is used to provide estimates of  $D$ ,  $H_1$ ,  $H_2$ , and  $F$ . In this solution,  $V_{0L0}$ ,  $V_{0L1}$ ,  $V_{1L1}$ , and  $W_{0L01}$  are assumed to be independent and normally distributed.

To avoid these assumptions, Nelder (38) and Crumpacker and Allard (6) suggested that each replication be treated as a separate experiment with its own environmental component of variance. Then by assuming that the estimates of these parameters from each block are samples of normal population, the standard error of the mean of each of these parameters can be estimated from the variation of the block values around the overall mean. This is considered desirable, since the distribution of the sample means tends toward normality with increasing size of samples, even though the individual varieties may not be distributed normally. The estimates of the diallel cross parameters in this study are made according to Nelder's suggestion with separate estimates of environmental variances.

A number of important estimators that may be derived from the diallel cross parameters (22) among others are:

1.  $H_1/D$ , an estimator of the average degree of dominance, since  $H_1/D = \frac{\sum u_i v_i h_i^2}{\sum u_i v_i d_i^2}$ . This quantity is weighted in favor of the genes which have both alleles represented equally in the parents and have large  $h$  effects (6). With average partial dominance,  $H_1/D$  is expected to have a value between 0 - 1. The square root of  $H_1/D$  is a weighted measure of the average degree of dominance at each locus.
2.  $(V_{1L1} - E)/(W_{0L01} - E/n)$  is another estimate of the average degree of dominance. It is a weighted estimator in the same sense as  $H_1/D$ , and with partial dominance it will also fall between 0 and 1.
3. The sign of  $(\bar{F}_1 - \bar{MP})$ , an indicator of the average direction of dominance. The variation of the individual  $(\bar{F}_1 - \bar{MP})$  around the mean of  $(\bar{F}_1 - \bar{MP})$  was used to estimate the standard error for the mean of  $(\bar{F}_1 - \bar{MP})$ .

4.  $\frac{1}{2}H_2/H_1$ , an estimator for the average frequency of negative versus positive alleles in the parents, since  $\frac{1}{2}H_2/H_1 = (\sum u_i^2 v_i^2 h_i^2) / (\sum u_i v_i h_i^2) = \bar{u}\bar{v}$ . It has a maximum value of  $\frac{1}{4}$  when  $u = v = \frac{1}{2}$ . If the negative and the positive alleles are not equally distributed among the parents,  $uv < \frac{1}{4}$ . This estimator is weighted in favor of the genes with large  $h$  effects. No information is provided for the genes that have no dominant effects.
5. The parameter  $F = 8 \sum [u_i v_i (u_i - v_i) d_i h_i]$ . Its sign depends on the sign of  $(u_i - v_i) h_i$ , and the magnitude of each of these parameters. If no genes exhibit dominant effects, or if the dominant and the recessive alleles of each gene are distributed equally among the parents,  $F = 0$ . In the excess of dominant alleles  $F$  will be positive, whereas an excess of recessive alleles will cause  $F$  to be negative. Thus the sign of  $F$  is an indicator of the relative frequencies of dominant and recessive alleles in the parents. When the  $h$  effects of each gene are unequal, the sign of  $F$  will be weighted in favor of the genes with large  $h$  effects.
6.  $\underline{K}$ , the effective factors =  $(\bar{F}_1 - \bar{P}) / \frac{1}{2}H_2$ .

An effective factor has been defined by Mather (29) as the smallest unit that is capable of being recognized by the methods of biometrical genetics. It may be a group of closely linked genes or, at the lower limit, a single gene. The value of  $\underline{K}$  will be underestimated unless the  $h$  effects of all genes are equal in sign and size, and the distribution of the genes is uncorrelated (20, 29). It also gives no information on the genes with no dominant effects.

Most of these estimators are ratios, and the question of their biasness immediately arises. This problem is a troublesome one. However,

the approximate standard errors of these ratios can be estimated similar to Nelder's suggestion in estimating the standard error for the diallel parameters, namely, from the variation of the ratios calculated from each block around the over-all mean of the ratios. For example, the ratio of  $H_1/D$  is determined for each block and the block values are used to estimate the mean, and the standard error of the mean of  $H_1/D$ .

D. Diallel Cross Graph. Consider equations (b), (f), (p), and (q). The quantity  $W_r - V_r$  is equal to  $\frac{1}{2}(D - H_1)$  and is expected to be constant over all arrays, independent of  $r$ , if the basic assumptions are valid and environmental effects are negligible. Thus,  $W_r = \text{constant} + V_r$ , under these conditions. By plotting  $W_r$  on  $V_r$ , the regression of  $W_r$  upon  $V_r$  is a straight line with unit slope (7, 23).

Consider equations (a), (b), and (f). Jinks and Hayman (23) have shown that there is an inequality of  $W_r^2 \leq V_p V_r$ . This means that all  $W_r$  and  $V_r$  points on the  $(V_r, W_r)$  graph should lie inside the parabola  $W_r^2 = V_p V_r$ .

Thus, the regression of  $W_r$  on  $V_r$  in the diallel cross or  $(V_r, W_r)$  graph can be used to test the validity of the basic assumptions, supported by the fact that all values of  $V_r$  and  $W_r$  should lie inside the  $W_r^2 = V_p V_r$  parabola.

Furthermore, the  $(V_r, W_r)$  graph is also a means to make some genetic analysis. Consider again the equation  $W_r = \frac{1}{2}(D - H_1) + V_r$ . When  $V_r = 0$ , then  $W_r = \frac{1}{2}(D - H_1)$ . Thus on the  $(V_r, W_r)$  graph, the  $W_r$  intercept is an indicator of the average degree of dominance in the experimental materials. With partial dominance,  $H_1 < D$ , the  $W_r$  intercept will be positive, whereas in overdominance,  $H_1 > D$ , the  $W_r$  intercept will be negative. With average complete dominance,  $H_1 = D$ , the regression line

will pass through the origin. In the case of no dominance, all points of the  $(V_r, W_r)$  graph will estimate one single point,  $W_r = 2 V_r$  (6, 7, 23). Dickerson and Jinks (7) illustrated these situations with certain values of dominance (Fig. 1).

The positions of the array points along the line of regression of  $W_r$  on  $V_r$  depend upon the relative proportion of dominant and recessive alleles present in the common parent of each array (13, 20). Parents with a preponderance of dominant effects will have a low array variance and covariance, and will lie near the origin. Highly recessive parents will have a large array variance and covariance, and will lie on the part of the regression line, away from the origin. If the dominant effects of the genes are unequal, the position of an array point will be weighted in favor of genes with large dominant effects.

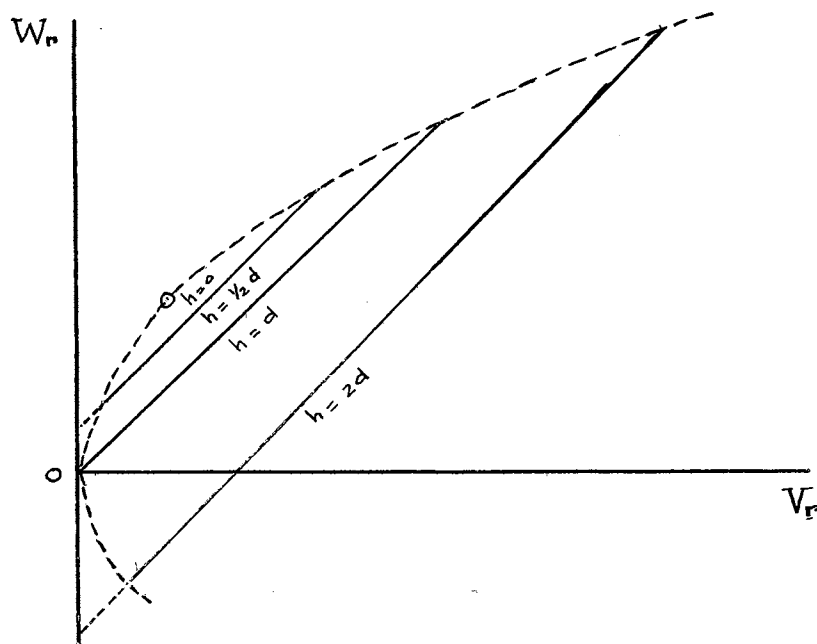


Fig. 1. The regression lines of  $W_r$  on  $V_r$  with varying degrees of dominance. After Dickinson and Jinks (7).



The analysis by means of the  $(V_r, W_r)$  graph can be applied and interpreted similarly to the  $F_2$  family (15, 23). Further, the graph of  $F_1$  array variances or covariances on the  $F_2$  variances or covariances should be on a straight line of slope  $\frac{1}{2}$  (15, 23).

Thus, the  $(V_r, W_r)$  graph provides a test of the validity of the basic assumptions ( $b_1 = 1$ ), the presence of dominance, and the average degree of dominance (the sign of  $b_0$ ), where  $b_1$  is the slope of the regression line and  $b_0$  is the  $W_r$  intercept.

Failure of any of the basic assumptions will cause deviation of the  $(V_r, W_r)$  points from the regression line with unit slope. With increasing proportion of heterozygosity in the parents, there will be a proportional shifting of the regression line upwards and to the left. The effect of this shift is to simulate a lower level of dominance. The deviation of the regression line from unit slope may be significant (7, 23). Similar bias may be due to allele frequencies correlation, or linkage (7). Non-allelic interaction, particularly complementary gene action, results in deviation of the points corresponding to the arrays of the interacting parents from the expected regression line. Hence, in such cases, the actual regression line will usually deviate from unit slope. It may cause extreme deviation of the corresponding points to the right and below the expected regression line (7, 13, 23).

E. Test of Non-allelic Interactions. One of the many recent explanations of heterosis is the joint action of favorable combinations of genes at different loci, that is non-allelic interactions or epistasis. Comstock and Robinson (5) pointed out that non-allelic interactions could inflate intra-allelic interactions. Jinks (21) suggested that apparent over-dominance may be due to epistasis. Henderson (18) suspected that

non-allelic interactions might be partly responsible for heterotic effects.

Due to the importance of non-allelic interactions, Hayman (14) introduced a method to test the significance of these interactions in the diallel cross experiment, when the  $F_2$  is included. It is a kind of Chi-square test for fit that can be calculated from the  $2L_2 - L_1$  table. This table is similar to the  $L_1$  or  $L_2$  table, and as the name suggests, it is constructed by subtracting each term of the  $L_1$  table from twice the term, in similar position, of the  $L_2$  table.

The Chi-square value for  $\frac{1}{2}n(n - 1)$  degrees of freedom is (14):

$$\text{Chi-square} = k_2 \left[ (n - 1)(V_{1LX} - V_{OLX}) + n(\bar{p} - \bar{x})/(1 + k) + (n - 1)(V_{OLO} - 4W_{OLOX} + 4V_{OLX})/(2 + k) \right]$$

Where:  $V_{OLO}$ ,  $V_{OLX}$ ,  $V_{1LX}$ , and  $W_{OLOX}$  are analogous to  $V_{OLO}$ ,  $V_{OL1}$ ,  $V_{1L1}$ , and  $W_{OLO1}$ , calculated from the  $2L_2 - L_1$  table.

$$k = nE_0/(8E_2 + E_1 - E_0)$$

$$k_2 = n/(8E_2 + 2E_1)$$

$E_0$ ,  $E_1$ ,  $E_2$ , and  $n$  are as defined before on page 12.

$\bar{p}$  = the mean of parents

$\bar{x}$  = the grand mean (of parents,  $F_1$ , and  $F_2$ ).

A significant Chi-square value indicates the presence of significant non-allelic interactions. It can be investigated in detail, whether it is manifested in the  $F_1$ , or  $F_2$  families. Then the epistatic crosses can also be classified into duplicate or complementary epistasis (14, 52).

## CHAPTER IV

### RESULTS AND DISCUSSION

In this experiment, the plants grown from seedlings survived better than those grown by direct seeding in the field. Many of the plants in replication 1 and replication 3 did not grow well, some had no or less than 10 open bolls during the harvest time. The number of observations or subsamples in replicate 1 varied from 0 to 13, whereas that in replicate 3 varied from 3 to 13. The smallest number of subsamples in replicate 2 and replicate 4 are 6 and 8, respectively. The complete number of subsamples for each entry in each replicate is given in Table I.

Due to the missing observation of the Gregg population in replicate 1, and the presence of very small (three) subsamples in replicate 3, only the analysis based on replicate 2 and replicate 4 is presented in this report. To avoid any effects of unequal subsamples in the diallel analysis, random samples (except for the entry that has the smallest number of subsamples) of 6 observations were taken from each population for statistical analysis.

#### Analysis of Variance

The analysis of variance based on these two replications is given in Table II. There are highly significant differences among the populations. Block effects, experimental and sampling errors are relatively small in comparison to the variation of the population. The differences

TABLE I  
NUMBER OF SUBSAMPLES

Generation	Entry	Replicate				Total
		1	2	3	4	
Parents	A	7	9	10	10	36
	G	0	12	3	10	25
	L	10	12	5	12	39
	S	9	12	8	12	41
F <sub>1</sub>	A x G	6	10	8	9	33
	A x L	12	12	9	11	44
	A x S	9	11	8	12	40
	G x L	13	12	10	12	47
	G x S	9	8	5	9	31
	L x S	9	9	9	11	38
F <sub>2</sub>	A x G	10	11	8	9	38
	A x L	9	11	5	11	36
	A x S	9	9	11	12	41
	G x L	5	13	8	11	37
	G x S	6	13	10	11	40
	L x S	11	12	5	9	37
B <sub>1</sub>	(A x G) x A	6	12	5	10	33
	(A x L) x A	7	10	6	11	34
	(A x S) x A	8	12	8	8	36
	(G x L) x G	10	6	7	8	31
	(G x S) x G	9	11	5	9	34
	(L x S) x L	9	10	9	12	40
B <sub>2</sub>	(A x G) x G	5	13	5	9	32
	(A x L) x L	6	12	5	9	32
	(A x S) x S	5	13	5	13	36
	(G x L) x L	2	12	8	10	32
	(G x S) x S	6	12	8	10	36
	(L x S) x S	8	12	3	10	33

Legend: A = Acala 4-42  
G = Gregg  
L = Lankart 57  
S = Stoneville 62

These abbreviations will be used in many instances in this report.

TABLE II  
ANALYSIS OF VARIANCE

Source of variation	d.f.	Sums of Squares	Mean Squares
Total	335	349.6630	
Replicate	1	0.3523	0.3523
Population	27	237.4092	8.7829**
Experimental error	27	12.6669	0.4691
Sampling error	280	99.2346	0.3544

\*\*Significant at 1% probability level.

among the populations are likely to occur, since two of the parents have high tensile strength, and the other two have low tensile strength.

#### Heterosis and Inbreeding Depression

The means of parents, mid-parents,  $\bar{F}_1$ ,  $\bar{F}_2$ ,  $(\bar{F}_1 - \bar{MP})$ , and  $(\bar{F}_2 - \bar{F}_1)$  are given in Table III. The estimate of the standard error of the difference between two means is 0.39. None of the calculated t - values of each of  $(\bar{F}_1 - \bar{MP})$  and of  $(\bar{F}_2 - \bar{F}_1)$  exceeds the tabulated t-value at 5% probability level. Thus, heterotic effect and inbreeding depression are not significant at 5% probability level.

All of the differences of the  $F_1$  and the mid-parent values have negative signs. The standard error of the mean of these differences is estimated to be 0.015. The 99% confidence limits on the mean of the differences are -0.11 and 0.01. Thus the sign of the average degree of dominance in the parents is negative, or the low tensile strength is dominant over the high tensile strength. This confirmed the findings of earlier workers (42, 45, 48).

TABLE III

MEANS OF PARENTS ( $P_1$ ,  $P_2$ ), MID-PARENTS (MP),  $F_1$   
 $F_2$ , ( $F_1 - MP$ ) AND ( $F_2 - F_1$ ) OF INDIVIDUAL  
 CROSSES, IN GRAMS/GREX

Crosses	$P_1$	$P_2$	MP	$F_1$	$F_2$	( $F_1 - MP$ )	( $F_2 - F_1$ )
A x G	4.00	4.10	4.05	4.02	4.04	-0.03	0.02
A x L	4.00	3.23	3.62	3.54	3.57	-0.08	0.03
A x S	4.00	3.47	3.75	3.64	3.50	-0.10	-0.14
G x L	4.10	3.23	3.67	3.61	3.72	-0.06	0.11
G x S	4.10	3.47	3.79	3.77	3.80	-0.02	0.03
L x S	3.23	3.47	3.35	3.34	3.50	-0.01	0.16
Mean			3.70	3.65	3.69	-0.05	0.04

In conclusion, no heterotic effects and inbreeding depression are detected, at 5% probability level, in individual crosses of the materials studied, although there are highly significant differences among the populations. The direction of the average degree of dominance in the parents is toward the low tensile strength, or on the average there is dominant effects of low tensile strength over high tensile strength.

#### Diallel Cross Graph

Before proceeding to the estimation of the diallel cross parameters, it is essential to see that the basic assumptions are valid. A method to test the validity of these assumptions is by analyzing the diallel cross or the ( $V_r$ ,  $W_r$ ) graph (7, 13, 23).

The means, overall replicates, of the parents,  $F_1$ , and  $F_2$  are given in Table IV and Table V, which are the  $L_1$  and the  $L_2$  tables, respectively. The means of parents are in the diagonal, and those of the crosses are in the off-diagonal positions in the respective table.

TABLE IV

MEANS OF PARENTS AND F<sub>1</sub>, IN GRAMS/GREX

Parent	A	G	L	S
A	4.00	4.02	3.54	3.64
G		4.10	3.72	3.80
L			3.23	3.35
S				3.47

TABLE V

MEANS OF PARENTS AND F<sub>2</sub>, IN GRAMS/GREX

Parent	A	G	L	S
A	4.00	4.04	3.57	3.50
G		4.10	3.72	3.80
L			3.23	3.50
S				3.47

Estimates of variance of parents ( $V_p$ ), variance of each array, ( $V_r$ ), and covariance of each array ( $W_r$ ) were calculated for the F<sub>1</sub> and F<sub>2</sub> generations from the L<sub>1</sub> and L<sub>2</sub> tables, respectively. For each of the array variance or covariance,  $r$  was substituted with  $\underline{a}$ ,  $\underline{g}$ ,  $\underline{l}$ , and  $\underline{s}$ , which stands for Acala 4-42, Gregg, Lankart 57, and Stoneville 62 arrays, respectively. The values of these estimates are given in Table VI.

The  $V_p$ ,  $V_r$ , and  $W_r$  values calculated from the L<sub>1</sub> and L<sub>2</sub> tables were plotted into the ( $V_r$ ,  $W_r$ ) graphs, respectively, in Figure 2 and Figure 3. In both graphs, all points lie inside the theoretical parabola  $W_r^2 = 0.1746 V_r$ . The slopes of the regression lines of  $W_r$  on  $V_r$  ( $b_1$ )

TABLE VI  
ESTIMATES OF  $V_p$ ,  $V_r$ , AND  $W_r$

Statistics	Generation	
	F <sub>1</sub>	F <sub>2</sub>
V <sub>p</sub>	0.1746	0.1746
V <sub>a</sub>	0.0605	0.0795
V <sub>g</sub>	0.0510	0.0337
V <sub>l</sub>	0.0303	0.0420
V <sub>s</sub>	0.0342	0.0242
W <sub>a</sub>	0.1027	0.1110
W <sub>g</sub>	0.0941	0.0765
W <sub>l</sub>	0.0725	0.0786
W <sub>s</sub>	0.0758	0.0423

in both graphs are not significantly different from one, and the  $W_r$  intercepts ( $b_0$ ) in both graphs are positive, but only in the F<sub>1</sub> generation is it significantly different from zero at 5% probability level. The complete regression analyses following the method given by Ostle (39) are in Appendix A and Appendix B.

From the results of the regression analysis, it seems likely that the basic assumptions are valid. However, the positions of the array points along the regression lines in figure 2 and figure 3 are not constant. This provides evidence that some of the basic assumptions are not strictly valid.

The position of the regression lines toward the hypothetical parabola in both graphs show that, on the average, the dominance effects are not complete. As previously shown, the direction of the average degree of dominance is negative; hence, there is partial dominance of the low over high tensile strength. This was also found by earlier workers



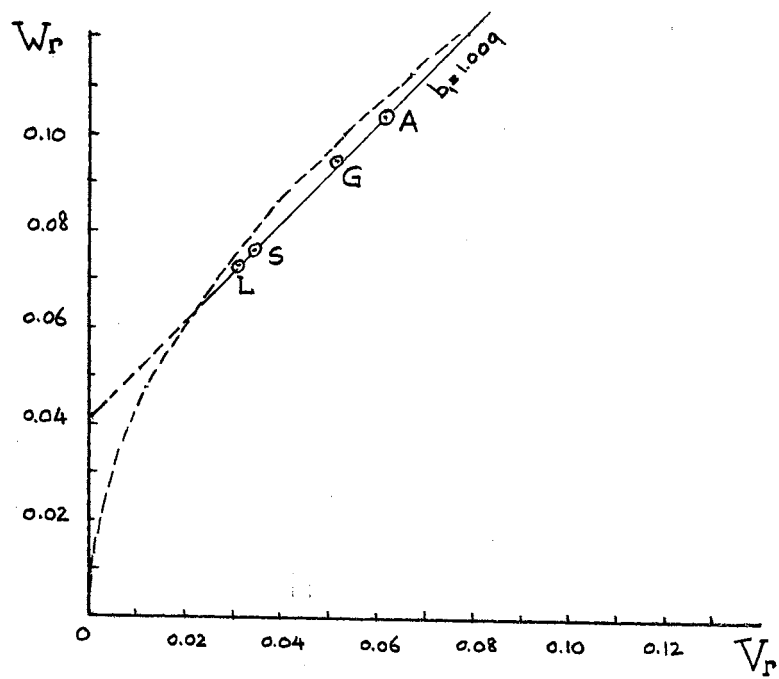


Fig. 2. The  $(V_r, W_r)$  graph of the  $F_1$  family.

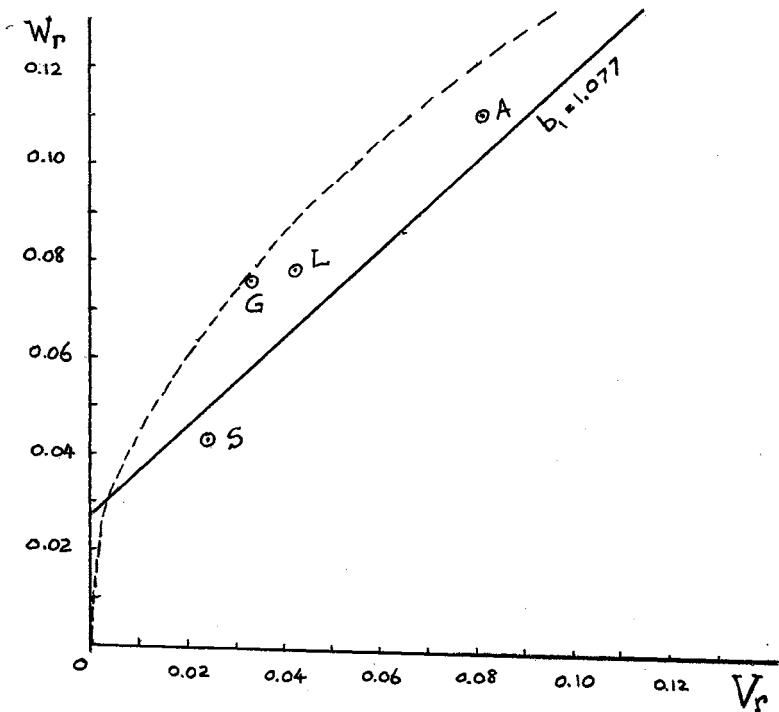


Fig. 3. The  $(V_r, W_r)$  graph of the  $F_2$  family.

(42, 45, 48).

The positions of the arrays along the regression line in figure 2, suggest that Lankart 57 and Stoneville 62 are parents with a preponderance of dominant alleles, and Acala 4-42 and Gregg are highly recessive parents. However, in figure 3 Gregg, Lankart 57, and Stoneville 62 do not maintain their positions. The downwards shifting of Gregg and Stoneville 62, suggests that these two parent probably contribute non-allelic interactions (7, 13, 23). The positions of Acala 4-42 and Lankart 57 shift upwards and to the right; an indication that these two parents are probably not quite homozygous. The position of Acala 4-42 in the  $F_1$  and  $F_2$  generations are always on the top of all arrays, farthest from the origin. Therefore Acala 4-42 is the parent that has the least dominant effects.

#### Test of Non-allelic Interactions

Due to the evidence of the presence of non-allelic interactions, as shown from Figure 2 and Figure 3, it is necessary to test the significance of these interactions. The method given by Hayman (12) was used here. Table VII is the  $2L_2 - L_1$  table.

TABLE VII

THE  $2L_2 - L_1$  TABLE\*

Parent	A	G	L	S
A	4.00	4.06	3.60	3.36
G		4.10	3.81	3.83
L			3.23	3.65
S				3.47

\*See text for explanation.

The calculated Chi-square value for 6 degrees of freedom is 0.94. The tabulated value at 5% probability level for 6 degrees of freedom is 12.6. Thus, non-allelic interactions exhibited by Gregg and Stoneville 62 arrays are not significant at the 5% probability level. This is also supported by the results of the regression analysis that the slopes of the regression lines in both generations do not deviate significantly from one. Since there are no significant non-allelic interactions, no further test was conducted to find the type of interactions.

#### Diallel Cross Analysis

Evidence from the regression analysis of  $W_r$  on  $V_r$ , in both  $F_1$  and  $F_2$  generations and the result of the Chi-square test for non-allelic interactions suggests that the basic assumptions are, to a certain extent, valid. Hence, the estimation of the diallel cross parameters can be justified.

As suggested by Nelder (38), by Crumpacker and Allard (6), and by Hayman (17), each replicate was treated as separate experiment, and the estimates of  $D$ ,  $H_1$ ,  $H_2$ , and  $F$  from each replicate are regarded as samples of the true parameters. In addition, the estimates of these parameters calculated from the  $F_2$  and backcross generations were also regarded as such samples, since these estimates are also estimates of the true parameters in the parents.

The sets of parent and  $F_1$  generation means, of the parent and  $F_2$  generation means, and the parent and the average of reciprocal backcross means for each replicate are presented in Table VIII, Table IX, and Table X. In most cases, the differences between the means of any generation in the two replicates are not apparent. This was expected,

TABLE VIII

MEANS OF PARENTS AND F<sub>1</sub> IN REPLICATE 2\*  
AND REPLICATE 4, IN GRAMS/GREX

Parent	A	G	L	S
A	3.90	4.01	3.54	3.69
	4.10	4.02	3.54	3.59
G		4.12	3.54	3.84
		4.08	3.67	3.70
L			3.22	3.35
			3.24	3.34
S				3.46
				3.47

\*Upper figures are the means in replicate 2.

TABLE IX

MEANS OF PARENTS AND F<sub>2</sub> IN REPLICATE 2\*  
AND REPLICATE 4, IN GRAMS/GREX

Parent	A	G	L	S
A	3.90	3.88	3.60	3.50
	4.10	4.20	3.54	3.49
G		4.12	3.62	3.83
		4.08	3.81	3.76
L			3.22	3.52
			3.24	3.48
S				3.46
				3.47

\*Upper figures are the means in replicate 2.

TABLE X

MEANS OF PARENTS AND  $B_1$  IN REPLICATE 2\*\*  
AND REPLICATE 4, IN GRAMS/GREX

Parent	A	G	L	S
A	3.90	4.09	3.39	3.72
	4.10	4.07	3.42	3.56
G		4.12	3.55	3.89
		4.08	3.78	3.88
L			3.22	3.44
			3.24	3.43
S				3.46
				3.47

\*The average of reciprocal backcrosses

\*\*Upper figures are the means in replicate 2.

since there were no significant differences between replicates, as indicated from the analysis of variance.

Estimates of variances and covariances needed to estimate the diallel cross parameters, calculated from each replicate for each generation, do not differ markedly. The low figures of the estimates are probably due to the scale of measurement used in this study.

Estimates of environmental variances for the parents,  $F_1$ , and  $F_2$  generations, namely,  $E_0$ ,  $E_1$ , and  $E_2$ , were calculated as the within group variance of each generation in each replicate. These estimates and each of their averages can be found in Table XII.

It appears that the estimates in both replicates are homogeneous, but no statistical test is conducted, since the estimates of each replicate will be used separately in the estimation of the diallel cross

TABLE XI

ESTIMATES OF THE SECOND DEGREE STATISTICS, USED FOR THE ESTIMATION OF THE DIALLEL CROSS PARAMETERS, OF  $F_1$ ,  $F_2$ , and  $B_1$  GENERATIONS

Statistics	$F_1$		$F_2$		$B_1$	
	Rep. 2	Rep. 4	Rep. 2	Rep. 4	Rep. 2	Rep. 4
$V_{0L0}$	0.1673	0.1923				
$V_{0L1}$	0.0454	0.0475				
$V_{1L1}$	0.0329	0.0416				
$W_{0L01}$	0.0845	0.0895				
$V_{0L2}$			0.0364	0.0722	0.0588	0.0589
$V_{1L2}$			0.0263	0.0462	0.0473	0.0459
$V_{2L2}$			0.0426	0.1276	0.1241	0.1153
$W_{0L02}$			0.0662	0.0899	0.0962	0.0881

TABLE XII

ESTIMATES OF ENVIRONMENTAL VARIANCES

Statistics	Rep. 2	Rep. 4	Mean
$E_0$	0.0441	0.0291	0.0366
$E_1$	0.0251	0.0536	0.0396
$E_2$	0.0342	0.0422	0.0382

parameters. The averages seem to be more uniform, and were used to determine the values of  $k$  and  $k_2$  for the test of non-allelic interactions. In general, the estimates of environmental variances are rather large, in comparison to some of the estimates of the second degree statistics given in Table XI.

By substituting the value of environmental variances into the equations given in the previous chapter, the estimates of  $D$ ,  $H_1$ ,  $H_2$ , and  $F$  were solved. These estimates, each of their means, and their standard errors are presented in Table XIII. Each of their standard errors were estimated from the variation of each estimate in each replicate in each generation, around their respective means.

Of all the estimates, only those of  $D$  that are consistently positive throughout the replicates and generations, and the mean of these estimates is significantly different from zero at the 5% probability level. The mean of the estimates of  $F$  is also positive, but its standard error is comparatively big. The mean of the estimates of  $F$  is significantly different from zero at approximately 20% probability level. Only by disregarding the estimates from the  $F_1$  generation in the calculation is the mean of  $F$  significantly different from zero at 5% probability level. The negative estimates of  $H_1$  and  $H_2$  are rather puzzling, since their true values should be either zero or positive. Probably the true values of each of these parameters, are very small. It may also be due to the large value of the estimates of environmental variances. As is observed from Table XI and Table XII, some of the estimates of variances that contain these two parameters, are even smaller than the environmental variances.

TABLE XIII  
ESTIMATES OF THE DIALLEL CROSS PARAMETERS,  
THEIR MEANS AND STANDARD ERRORS

Parameter	<sup>F</sup> <sub>1</sub>		<sup>F</sup> <sub>2</sub>		<sup>B</sup> <sub>1</sub>		Mean	Standard Error
	Rep. 2	Rep. 4	Rep. 2	Rep. 4	Rep. 2	Rep. 4		
D	0.1232	0.1632	0.1864	0.3144	0.3212	0.2938	0.2337	0.0787
F	-0.0476	-0.0024	0.3040	0.5968	0.6032	0.5288	0.3305	0.2705
H <sub>1</sub>	-0.1600	-0.1892	-0.3056	0.0544	0.0768	-0.0027	-0.0877	0.1399
H <sub>2</sub>	-0.1468	-0.1796	-0.6368	-0.9216	-0.6214	-0.7136	-0.5371	0.2438



It can be concluded that the estimate of  $D$  is bigger than that of  $H_1$ , or presumably the genetic variance of the parents are primarily additive and/or additive by additive components. The component of variance due to the dominant effects,  $H_1$ , is either zero or very small. If it were zero, most probably the dominant effects of each locus cancel each other.

The positive value of the mean of  $F$  suggests that the frequency of positive alleles, in this case the high tensile strength, in the parents is higher than its negative counterparts. It was rather expected, because from the history of these four varieties, they had been selected for their agronomic characteristics, probably including their tensile strength.

Due to the negative estimates of  $H_1$  and  $H_2$ , (and they are also not significantly different from zero) some of the estimators that contain any or both of these parameters were not estimated. The average degree of dominance, that can be estimated as  $H_1/D$ , was estimated by  $(V_{1111} - E) / (W_{0101} - E/n)$ , where  $E$  is the average of  $E_0$  plus  $E_1$  over all replicates. Since there are only two estimates of  $V_{1111}$  and  $W_{0101}$ , their means were used in the calculation. The estimate calculated thusly is 0.09. Therefore, on the average the dominant effect among the parents is not complete. No statistical test was conducted on this estimates. This value supports the evidence shown from the  $(V_r, W_r)$  graph of partial dominance of the character.

Estimates of heritability were calculated as the additive and/or additive by additive genetic portion of the variance of parents ( $V_{0101}$ ), and as that of the mean variance of arrays ( $V_{1111}$ ). These estimates are 79% and 94%, respectively. In both cases, the heritability estimates

are high.

#### Discussion

Cotton breeder, as many other breeders of self-pollinated crop plants, realize slow progress has been achieved in breeding quantitative characteristics. This slow progress, in part, may be due to a slow development of the breeding principles for the complexly inherited character.

Two important problems are faced by the breeders of self-pollinated plants. First, to choose the best parents for hybridization, and second, the selection of the best lines from hybrid progenies. The efficiency of breeding self-pollinated plants, thus depends on accurate identification of the hybrid combinations that have a good potential of producing maximum improvement, and on identifying in early generations superior lines among the progeny of the promising hybrids.

Diallel cross analysis is probably the most popular design for assessing quantitative variability in self-pollinated crops, because it may be used to characterize crossing relationships among a group of varieties or lines (30). In a way, this study was aimed towards the assessment of the progress that could possibly be obtained from the diallel cross study.

In choosing the best parents for hybridization, naturally, the first thing to see is whether any of the  $F_1$  crosses show some significant heterotic effect in the desirable character. In this study, none of the crosses showed significant heterotic effects, as indicated from the comparison of the  $F_1$  of individual crosses and their mid-parents value. By comparing the array points along the regression line, Acala

4-42 seems to be constantly the variety most removed from the origin. This suggests that the crosses with Acala 4-42 might be the promising hybrids, at least among the crosses in this study. Although the Gregg array in the  $F_1$  generation is almost in the same position as Acala 4-42, it shifts downwards in the  $F_2$  generation, an indication of the instability of this array, which is probably due to non-allelic interactions. Thus, based on this study, although Gregg in itself has tensile strength as high as Acala 4-42, its hybrids are less promising than those of Acala 4-42.

The  $(V_r, W_r)$  graph can also be used to identify which of the parents, if any, contribute to the significant deviation due to non-allelic interactions. It can be done visually from the graph, or probably more accurately by conducting the regression analysis, using all possible combinations of all arrays but one. The results of the regression analysis in this study do not indicate that such a test should be conducted, since in the  $F_1$  and the  $F_2$  generations, the slope of the regression line do not significantly differ from one, and non-allelic interactions are not significant in the materials under study.

In the presence of significant non-allelic interactions, further tests can be conducted to determine in which generation these interactions are manifested, and whether the interactions are complementary or duplicative (14).

The second use of diallel cross is concerning information of the base population, from which the parents is a sample. Due to the small number of parents included in this study, none of the components of variation, either statistical or genetical, can be significant estimates of the population parameters (17). However, based on Eisenhart's (8)

suggestion, Hayman (17) stated that the estimates be interpreted, for the particular set of parents, as if it would have been interpreted for a population when derived from a sufficiently large sample of parents. Thus, as it has been summarized in the conclusion, presumably the additive and/or the additive by additive components composed primarily the genetic variance in the parents.

Large estimates of heritability of the trait suggest that improvement can be efficiently made by mass selection within the promising hybrids. Thus, mass selection applied to Acala 4-42 array, in particular to the best cross among this array, is expected to be effective for improving tensile strength.

One of the important problems in using Hayman's method is to see that the basic assumptions can be fulfilled for the analysis to be valid. It was indicated that certain of these assumptions are not strictly valid for these materials. One of these was that of no non-allelic interactions, as indicated from the unstable position of the array points, and that of no linkage. The latter is presumably present because the materials are selected varieties. Nevertheless, since these partial failures of the assumptions seemed unlikely to produce gross biases in the genetic analysis, it was concluded that the application of the diallel analysis to the data is justified.

Another problem involves practical difficulty. By accepting the assumptions of Crumpacker and Allard (6) that block values be treated as samples, which is also justified by Hayman (17) for small diallel crosses, ample number of blocks should be provided. This means more seeds are needed, and more crosses should be made. Thus, creating a practical problem of tedious hand pollinations, which will increase with

increasing number of parents.

In general, providing that these problems can be overcome, diallel cross analysis is a valuable tool, that can guide the breeders in improving quantitative characters in self-pollinated crop plants. To test the stability of the estimates of the parameters and the interaction of any of the parameters with environment, the experiment should be repeated in time and space. The inclusion of the backcross generations is not quite necessary in this analysis; the  $F_1$  and  $F_2$  generations can provide ample information.

## CHAPTER V

### SUMMARY AND CONCLUSIONS

A diallel cross analysis of fiber strength was conducted among four commercial varieties of upland cotton. The study included the parents,  $F_1$ ,  $F_2$ , and backcross generations.

The objectives of the study were to characterize the crosses among these varieties, and to obtain information on the genetic system of the parents, with respect to fiber strength.

The analysis of variance showed that there were differences among the populations. Comparisons of the means of  $F_1$  and the mid-parents values indicated that heterotic effects were not present in any of the  $F_1$  crosses. Inbreeding depression, as shown from the differences of the  $F_1$  and  $F_2$ , was not apparent.

Regression analysis of  $W_r$  on  $V_r$  indicated that the basic assumptions of the diallel cross analysis were valid, to a certain extent; thus, estimations of the diallel cross parameters were justified. The  $(V_r, W_r)$  graphs revealed, however, that certain of these assumptions are not strictly valid for these materials, as indicated by the instability of the positions of some array points along the regression lines.

Low tensile strength was a partially dominant character as indicated by the positive  $W_r$  intercept of the regression line. This is supported by an estimate of 0.09 for the average degree of dominance.

The positive, recessive alleles among the parents were more frequent than their negative, dominant counterparts. Due to the negative estimates of  $H_1$  and  $H_2$ , the estimator of the average frequencies of the positive versus negative alleles in the parents were not estimated.

Presumably the genetic variance of the parents was primarily additive and/or additive by additive in nature; the dominance component was probably very small comparatively.

The heritability estimates were high, suggesting that mass selection should be effective in improving fiber strength in the materials being studied.

Acala 4-42 seemed to be a promising parent. The position of this array along the regression line in the  $F_1$  and  $F_2$  generations were constantly on the top of the rest of the arrays. This indicates that Acala 4-42 has the least dominant effects. The position of Gregg along the regression line were unstable, suggesting that it was one of the parents that contributed to non-allelic interactions. Thus, based on this study, although Gregg has virtually the same high tensile strength as Acala 4-42, its progeny is not as promising as that of Acala 4-42. Mass selection of the best among Acala 4-42 hybrids should be an effective method in improving fiber strength, at least among the hybrids included in this study.

The conclusions given above cannot be regarded as decisive, not only due to the small number of replicates, but also due to the relatively large estimates of environmental variances. The experiment should be repeated over years and locations to see that the estimates of the parameters are stable, and to be able to test the magnitudes of the genotype-environment interactions (2).

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APPENDIX A

THE REGRESSION ANALYSIS OF  $W_r$  ON  $V_r$  OF THE  $F_1$  FAMILY

Array	$V_r$	$W_r$
A	0.0605	0.1023
G	0.0510	0.0941
L	0.0303	0.0725
S	0.0342	0.0758
Total	0.1760	0.3447
Mean	0.0440	0.0862

$$\sum x^2 = 0.00060498 \quad \sum xy = 0.00061056 \quad \sum y^2 = 0.00061747$$

$$b_1 = 0.00061056 / 0.00060498 = 1.009$$

$$b_0 = 0.0862 - 1.009 \times 0.0440 = 0.0418$$

Analysis of Variance

Source of variation	d.f.	Sums of Squares	Mean Squares
Total	4	0.03032199	
Due to $b_0$	1	0.02970452	0.02970452
Due to $b_1/b_0$	1	0.00061619	0.00061619
Residual	2	0.00000128	0.00000064

$$s_{b_1}^2 = 0.00000064 / 0.00060498 = 0.00105788 \quad s_{b_1} = 0.0325$$

$$s_{b_0}^2 = 0.00000064 (0.25 + 0.00208724 / 0.00060498) = 0.000023$$

$$s_{b_0} = 0.000023$$

$$b_0; t = (0.0418 - 0) / 0.000023 = 18.1 \times 10^2; t(0.05, 2) = 4.303$$

$$b_1; t = (1.009 - 1) / 0.0325 = 0.0028$$

APPENDIX B

THE REGRESSION ANALYSIS OF  $W_r$  ON  $V_r$  OF THE  $F_2$  FAMILY

Array	$V_r$	$W_r$
A	0.0795	0.1110
G	0.0337	0.0765
L	0.0420	0.0785
S	0.0242	0.0423
Total	0.1794	0.3083
Mean	0.0449	0.0771

$$\sum x^2 = 0.00175949 \quad \sum xy = 0.00189596 \quad \sum y^2 = 0.00236257$$

$$b_1 = 0.000189596 / 0.00175949 = 1.077$$

$$b_0 = 0.0771 - 1.077 \times 0.049 = 0.0287$$

Analysis of Variance

Source of variation	d.f.	Sums of Squares	Mean Squares
Total	4	0.02612479	
Due to $b_0$	1	0.02376222	0.02376222
Due to $b_1$	1	0.00204195	0.00204195
Residual	2	0.00032063	0.00016032

$$s_{b_1}^2 = 0.00016032 / 0.00175949 = 0.0911737 \quad s_{b_1} = 0.3018$$

$$s_{b_0}^2 = 0.00016032 (0.25 + 0.00245139 / 0.00175949) = 0.00026344$$

$$s_{b_0} = 0.0162$$

$$b_0; t = (0.0287 - 0) / 0.0162 = 1.770; t(0.05, 2) = 4.303$$

$$b_1; t = (1.077 - 1) / 0.3018 = 0.255$$

VITA

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