THE SEPARATION AND DETERMINATION OF 137_{Cs}

IN BIOLOGICAL MATERIALS

By

ROBERT JERREL EVERETT 41 Bachelor of Arts

University of New Mexico Albuquerque, New Mexico

1956

Submitted to the Faculty of the Graduate College of the Oklahoma State University in partial fulfillment of the requirements for the Degree of MASTER OF SCIENCE May, 1971



THE SEPARATION AND DETERMINATION OF ¹³⁷Cs

IN BIOLOGICAL MATERIALS

Thesis Approved:

Horacio A M. Thesis Ad ole on E. Mo Varga

Dean of the Graduate College

ACKNOWLEDGEMENT

Sincere appreciation is expressed to my adviser, Dr. H. A. Mottola, whose patience and understanding made this work possible. His efforts throughout the study exemplified to me the finest in adviser-student relationships. Thanks are due also to Dr. O. C. Dermer, whose critical review was most helpful. And last but not least, my gratitude is expressed to my wife, Jean, who graciously agreed to suffer the privations of additional graduate work.

e., .

TABLE OF CONTENTS

Chapte	Page	
I.	INTRODUCTION	
	Importance of ¹³⁷ Cs Determination in Biological Material	
II.	STRUCTURE OF AMMONIUM MOLYBDOPHOSPHATE	
III.	ION EXCHANGE COLUMN DESIGN	
IV.	EXPERIMENTAL	New Y
	Materials and Equipment	
	Recommended Procedure	
V.	CONCLUSIONS AND RECOMMENDATIONS	
A SELE	CTED BIBILIOGRAPHY	

LIST OF TABLES

Table								Pa	age
I. Oxygen-Oxygen	Interatomic Dis	stances .	• • •	•		• •	• •	•	8
II. Pauling Ionic	Radii for Some	Ions	• •	• •	• • •	• •	••••	•	9
III. Experimental D	istribution Val	ues	• •	• •		• •	••	•	21
IV. A Comparison o Elution Para	f Measured and meters	Calculat	ed	•		• •	• •		23

LIST OF FIGURES

Figur	re		Page	8
1.	Photographs of Models of the Phosphomolybdate Anion a) showing a flat face b) showing a re-entrant face	••	• (6
2.	Gamma Energy Spectrum for the 86 Rb Standard Solution	••	. 1	3
3.	Gamma Energy Spectrum for the $^{137}\mathrm{Cs}$ Standard Solution	• •	. 1	5
4.	Elution Curve for 86 Rb and 137 Cs Activities Using $\underline{3M}$ NH ₄ NO ₃ and Concentrated NH ₄ NO ₃	•	. 2	4
5.	Elution Curve for 86 Rb Using $4\underline{M}$ NH ₄ NO ₃	• •	. 2	5
6.	Distribution Ratios Vs pH for ¹³⁷ Cs Extraction By 0.5M (BAMP)	· ·	• 3	0
7.	Gamma Energy Spectrum of ¹³⁷ Cs After Analysis	•	• 34	4

v

CHAPTER I

INTRODUCTION

Importance of ¹³⁷Cs Determination in Biological Materials

The importance of ¹³⁷Cs as a major fission product and public health hazard need not be emphasized. Because of its chemistry this isotope has been spread throughout the world and is to be found in air, soil, plant and animal life. ¹³⁷Cs finds its way into man mainly through the ingestion of contaminated food products. The radiation dose suffered by a 70-kilogram adult is 0.06 rad per microcurie of intake¹⁸. ¹³⁷Cs becomes distributed throughout the body uniformly and is eliminated mainly via urinary excretion with a 100-day biological half_life¹⁸. ¹³⁷Cs has been determined in almost every conceivable material, 14 mainly by multi-channel gamma spectrometry, if the 137Cs activity is sufficient to permit resolution. The Los Alamos Biomedical Research group has determined the ¹³⁷Cs content of thousands of humans over the past 20 years, mainly by whole body scintillation counting systems. In addition to ¹³⁷Cs distributed by weapon fall-out, ¹³⁷Cs is used in large quantities as a standard gamma source and in chemical tracer work. The likelihood of accidental human exposure becomes greater as more ¹³⁷Cs is being used throughout the world. A rapid analytical procedure for determining ¹³⁷Cs in urine would be valuable in assessing human exposure from excretion data. It is thought that any procedure suitable for use in separating and isolating ¹³⁷Cs from

samples such as urine could be applied equally well to other biological and environmental samples, such as tissue, blood, plants, soils, etc.; however, this work is concerned only with the analysis of urine. The main problem in the determination of 137 Cs at low levels in biological samples is its separation from bulk matter and other emitters, such as naturally occurring 40 K and 87 Rb as well as fission products.

Typical procedures for determining ¹³⁷Cs in biological materials involve acid digestion and destruction of organic matter and the collection of cesium as silicotungstate or chloroplatinate for counting. These procedures are semiquantitative and time consuming, involve tedious precititations necessary for purification, and require corrections for low yields.

The intent of this work was to investigate ion exchange and solvent extraction procedures that would be quantitative, rapid, effective in removing activity, and amenable to final determination of ¹³⁷Cs by low background beta counting systems.

Analytical Approach to the Determination of ¹³⁷Cs in Urine

In reviewing the literature it was found that several derivatives of the 12-heteropolyacids have been used to isolate cesium from many interfering elements by cation exchange²⁰. In fact, it has been known for some time that ammonium molybdophosphate (AMP) shows ion exchange selectivity for the alkali metal ions ¹⁹, ²⁰, ²¹. This material has been used mainly in England and continental Europe and for some reason has not been widely used in this country. Apart from the ammonium ions, only the large monovalent cations, viz. K^+ , Rb^+ , Cs^+ , Tl^+ , Ag^+ , Hg_2^{++} and some organic bases, have been found to form insoluble salts with

AMP²⁰. The ion exchange reaction between alkali metal ions and AMP can be represented by:

$$M^+ + RNH_4 \rightleftharpoons MR + NH_4^+$$

where R represents the molybdophosphate anion and M^+ the metal ion. The equilibrium expression is:

$$K = \frac{\left[MR\right]\left[NH_{4}^{+}\right]}{\left[M^{+}\right]\left[RNH_{4}^{+}\right]}$$

Where $D_x = \begin{bmatrix} MR \\ M^+ \end{bmatrix}$ = distribution ratio = Quantity of x per unit Mass of AMP

If $\text{RNH}_{\mathcal{L}}$ is present in so great excess that it can be regarded as constant, the expression reduces to:

$$\log D = constant - \log NH_4^+$$

Fortunately, the distribution ratio D for AMP and the above cations are in the order $Cs^+ > Rb^+ > K^+ > T1^+ > Ag^+ > Hg_2^{++}$. These relatively large D values are attributed to size and the unique structure of the molybdophosphate anion. Krtil¹³ has investigated the exchange dependence upon the chemical environment, such as the HNO₃ and NH₄⁺ concentrations. A plot of log D for cesium vs log $[NH_4^+]$ indicates a slope of -1 and thus suggests that strong ammonia solutions would decrease the D value for cesium and therefore be an effective eluting agent after exchange. The dependence of D for cesium on nitric acid concentration is nearly constant up to about 4M acid, where it shows a sharp drop.

If the D values for the alkali metals are sufficiently different, adequate separations should be possible by selective elution with NH_{4}^{+} solutions. The really important separation is that of cesium from rubidium since other alkali metals would be less strongly held and eluted long before rubidium. Good separations have been achieved between cesium and rubidium on relatively clean samples (low ionic strength)^{19,21}, by the elution of rubidium with 3<u>M</u> NH₄NO₃ and the elution of cesium with about 15<u>M</u> NH₄NO₃. The separation and determination of cesium in sea water¹⁵, in rain water⁶ and urine¹⁷ use this basic approach. In these cases, the AMP was utilized as a primary collection material and further purification was necessary.

In view of the above it was decided to consider the application of AMP as a collection and purification material in the determination of cesium in biological materials. It was not known whether biological samples would yield distribution ratios sufficiently large in order to permit a good separation from interfering elements or the necessary column parameters. Later work did indicate large D values for cesium, allowing good separation, but presented another problem in the excessive amount of saturated $\mathrm{NH}_{\mu}\mathrm{NO}_{3}$ required to remove cesium after exchange. The possibility of using other eluting agents in less concentrated solution was studied without success. The recourse, then, was to dissolve the AMP containing cesium with strong base and equilibrate this solution with BAMBP, the only known extractant of cesium from strongly basic solutions 4,24. The BAMBP extraction system was investigated and found to be adequate. The resultant organic extractant solution of ¹³⁷Cs could then be counted directly by gamma spectrometry or the ¹³⁷Cs back-extracted into highly acidic solution, concentrated by evaporation and determined by low level beta counting.

CHAPTER II

STRUCTURE OF AMMONIUM MOLYDOPHOSPHATE

This compound is derived from a group of compounds known as the heteropolyacids. The heteropolyacids are compounds in which one atom of an element such as P, Si, B, or Al is combined with a number of atoms of an element such as W or Mo together with a relatively large number of oxygen atoms. Throughout the whole class of heteropolyacids they all have the same cation-cation ratios, are isomorphous and have similar chemical and physical properties. The best known compounds of this group are the silicotungstic, molybdosilicic, and phosphotungstic acids. The crystalline structure of these 12-heteropolyacids, which is the same for all of them, was clarified by Keggin¹² using X_ray powder photographs of 12-phosphotungstic acrd. A molecule of this acid has the formula $H_3(PW_{12}O_{40})$. The anion $[PW_{12}O_{40}]^{-3}$ consists of a central PO_{4} tetrahedron, surrounded by 12 WO₆ octahedra as a shell, linked together by shared oxygen atoms. (In order to see the unique structure of this molecule, a wooden model was constructed. This model is shown in Figure 1.) It can be seen that each WO₆ octahedron besides sharing two oxygen atoms with octahedra in its own group of three also shares two oxygen atoms with octahedra in other groups. In each anion, each WO_6 octahedron consists of one oxygen shared between three $^{\rm WO}_6$ octahedra and one $^{\rm PO}_4$ tetrahedron, four oxygen atoms shared between two WO_6 octahedra, and one oxygen atom unshared with other polyhedra,



Figure 1a Model of the Phosphomolybdate Anion showing one flat face.



Figure 1b Model of the Phosphomolybdate Anion showing re-entrant face.

these oxygen atoms are symmetrically distributed about the central tungsten atom. It is also noted that two edges of each WO₆ octahedron are shared with edges of two other octahedra. The oxygen atoms in each octahedron can be classified according to their position:



The 01 oxygen atoms are shared between three WO_{6} octahedra and one PO_{μ} tetrahedron. The O2 atoms are shared between two WO_{6} octahedra. The 03 atoms are shared between two WO_6 octahedra. The 04 atoms are unshared. The 01-03 edges are shared between two WO₆ octahedra. The surface presented by this molecule is interesting indeed. The surface consists of 6 cube faces and 8 octahedral faces. The latter can be divided into four flat faces and four re-entrant faces. Since these anions are complete structural units in themselves and when packed together in a crystal are not strongly linked to one another, the acids derived from them would be expected to be highly soluble and contain large amounts of water of crystallization. In the 5-hydrate packing, each of 4 flat faces is directly linked to a flat face of a neighboring anion and each of the 4 re-entrant faces is linked to a water molecule. These water molecules serve to bind together the re-entrant octahedral faces of adjacent anions. Therefore in the 5-hydrate structure¹¹ we have cubic packing, each anion having 14 neighbors. Four are linked directly, flat to flat, four indirectly through water molecules, and 6 are unattached leaving spaces large

enough for water molecules or other ions. The cesium salts of the 12-heteropolyacids form compounds that are isomorphous with the 5-hydrate acids and cesium ions replace water molecules in cube faces¹². In addition to cesium there are a few other monovalent ions that are large enough to be packed stably into this structure¹⁹. These are K^+ , Rb^+ , Tl^+ , Ag^+ , Hg_2^{++} , NH_4^+ and cations derived from some organic bases. The ions evidently lower the crystal energy sufficiently to give salts insoluble in water. Owing to the dependence of crystal packing on size and also to ion exchange it becomes important to determine the sizes of spaces available. The interatomic distances are as follows¹², ¹.

- a. The PO_4 tetrahedra in the center of the anion is undistorted; all edges are 2.80 Å.
- b. The WO₆ octahedra are distorted, the shared edges (01-03) are 2.65 Å.

TABLE I

OXYGEN_OXYGEN INTERATOMIC DISTANCES

 01-02
 2.70 Å

 03-03
 2.90 Å

 02-02
 2.65 Å

 03-02
 2.61 Å

 02-04
 3.10 Å

 04-03
 2.80 Å

In the 5-hydrate heteropolyacid, the packing is cubic, with the edge of the unit cube being 12.140^{+} .005 Å¹². Depending upon the orientation of the anions in the lattice, the maximum separation is believed to be about 3.5 Å¹². The reported ionic radii of the monovalent ions that yield insoluble salts with the 12-heteropolyacids are shown in Table II.

TABLE II

PAULING IONIC RADII FOR SOME IONS



While Cs⁺ has the largest diameter and this undoubtedly accounts for the stability of cesium molybdophosphate, it would seem that other ions may fit more precisely in the AMP lattice; however, after consideration of the obvious next possibilities (Fr, Ba, tetraalkyl ammonium ions) room for exploration in this line seems limited. There seems little doubt that AMP and other heteropolyacid salts owe their cation exchange properties and high selectivity for certain monovalent ions to their unique crystalline structure. Another point worthy of

consideration still related to ionic size, but without direct geometric implications, is that Cs has a low enthalpy of hydration and if these water molecules are lost upon exchange with AMP, this would help explain the large selectivity of AMP for Cs over the other alkali metal ions of smaller size and higher enthalpy of hydration.

 $\Delta H_{hyd}^{(2)} \text{ (kcal mole^{-1})} \qquad \frac{\underline{\text{Li}}^{+}}{-119} \frac{\underline{\text{Na}}^{+}}{-93} \frac{\underline{\text{K}}^{+}}{-73} \frac{\underline{\text{Rb}}^{+}}{-67} \frac{\underline{\text{Cs}}^{+}}{-59} \frac{\underline{\text{NH}}_{\mu}^{+}}{<-59} \text{ (estimate based upon size)}}$

CHAPTER III

ION EXCHANGE COLUMN DESIGN

In order to have an idea as to the characteristics of the desired column, a theoretical calculation was made based upon data that was available.

Ion exchange capacity of AMP = 1.57 meq/g

Particle size (r) = 74 microns = 0.0074 cm

Column cross section = 1.0 cm^2

Fraction of volume containing AMP (B) = 0.5 estimated¹⁰ Diffusion rate (\overline{D}) = 3.7x 10⁻⁷ cm²/sec estimated¹⁰ Diffusion rate in liquid phase (D_m) = 2x10⁻⁵ cm²/sec estimated¹⁰

Density of AMP = 5.5 g/cm^3

Optimum volume distribution ratio (D_v) = amount of ion in one ml of AMP/interstitial volume

Optimum flow rate (V) = ml/min

Of those estimated values D_m appears as the more reliable with B and \overline{D} probably less accurate.

1. Theoretical plate height (H). Using the derivation of Helfferich¹⁰ the plate height is given by:

$$H = 1.64r + \frac{Dv}{(Dv+B)2} \underbrace{\begin{bmatrix} 0.142r^2 V \\ \overline{D} \end{bmatrix}}_{p} + \underbrace{\begin{bmatrix} Dv \\ (Dv+B) \end{bmatrix}}_{m} \underbrace{\begin{bmatrix} 0.266r^2 V \\ D_m(1+70rV) \end{bmatrix}}_{m} + \frac{D_m B}{V} \underbrace{\begin{bmatrix} 0.266r^2 V \\ V \end{bmatrix}}_{r}$$

These four terms reflect the parameters that affect the column. The first term is due to particle size dependence, the second to

particle diffusion, the third to film diffusion and the fourth to longitudinal diffusion. The usual procedure for evaluation is to double the first term, set all other terms equal to this double value, and then sum. Therefore:

$$H = 6(1.64r) = 0.073$$
 cm

2. Optimum volume distribution ratio (D_V) The second term is set equal to the third term, the equation solved for D_V , and this evaluated.

$$D_v = \frac{0.142 \ D_m}{0.266 \ \overline{D}} = 29$$

3. Mean molar distribution ration (D_r)

$$D_{x} = \frac{Dv}{B} + \frac{29}{0.5} = 58$$

4. Optimum flow rate (V)

$$V = \frac{3.28 \text{ Dm} (D_{V}+B)^2}{0.266 \text{ D}_{V}^2 \text{T}} = 2.1 \text{ ml/min}$$

5. Relative peak retention (~)

$$= \frac{D_{v,Cs} + 1}{D_{v,Rb} + 1}$$

Unfortunately, the individual distribution ratios (D_v) for cesium and rubidium are not known and must be derived by other means. The best approach relies on previous elution data,¹⁹ since $V_{max} = V_{T,b}(D_v+B)$, where $V_{t,b}$ is the total bed volume and V_{max} is the volume of eluant required to elute the ion at the highest concentration (ie, maximum peak height). Using values from Figure 2 of Smit's work¹⁹: $V_{max}(RB) = 0.15$ ml and $V_{max}(CS) = 0.7$ ml and $D_{v,Rb} = 8.6$ $D_{v,CS} = 44$ The relative peak retention (\searrow) is: 4.7



6. Number of theoretical plates (N)

Glueckauf⁸ has developed the interrelationship of product purity, relative peak retention, and number of theoretical plates. If we set the maximum allowable Rb concentration at 0.01%, representing a large separation factor, reference to Figure 3 of Glueckauf's work indicates 25 plates are required. The length of the column (L) = NxH = 1.8 cm and a l-cm² cross section gives 1.8 cm^3 of AMP. However, this calculation is based upon pure AMP, and it is known that some substrate such as asbestos is required in order to obtain sufficient porosity for adequate flow^{6,15}. On the assumption that one part asbestos and two parts AMP (by weight) would be required, 1.8 cm^3 of AMP corresponds to 10 g. A 5 g amount of asbestos corresponds to 2.5 cm³ in a l-cm² cross section column. The effective plate height H* now may be calculated as:

$$H^* = \frac{L}{N} = \frac{4.3}{25} = 0.17 \text{ cm}$$

7. Elution

$$V_{max(Cs)} = V_{t,b}$$
 (Dv+B) = 96.8 ml
 $V_{max(Rb)} = V_{t,b}$ (Dv+B) = 20.7 ml

8. Band width (W)

$$W(Cs) = \frac{4V}{\sqrt{N}} = 77.5 \text{ ml}$$
$$W(Rb) = \frac{4V}{\sqrt{N}} = 16.5 \text{ ml}$$

9. Time required (T)

$$\Gamma = \frac{(V_{max}(C_s) + V_{max}(R_b))}{V} = 56 \min$$



These theoretical calculations were used to set up the initial experimental conditions for elution. A comparison will be presented later of theoretical and measured values. The two critical parameters, number of plates and relative peak retention, are thought to be reasonable and should give a good separation of Rb and Cs. It is generally accepted that overdesign is necessary and this was taken into account by imposing a high degree of separation (0.01%), consequently the calculated parameters cannot be considered attainable or to be expected but as guides for further refinement of conditions as adjusted to empirical behavior.

CHAPTER IV

EXPERIMENTAL

Materials and Equipment

A. Equipment

Ainsworth Analytical Balance, type 21N.

Beckman Zeromatic pH meter.

Multi-Channel gamma spectrometer, Nuclear Data Inc., series 1100 with 3" X 3" well-type NaI crystal.

B. Materials

Fisher certified A. C. S. grade chemicals:

Ammonium Nitrate

Phosphomolybdic Acid

Sodium Hydroxide

Phosphoric acid (ortho, 85%)

Nitric acid (71%)

Ammonium Hydroxide (29.6%)

⁸⁶Rb isotope - New England Nuclear standard NE-072: 10 microcuries as RbCl in 0.5 N HCl.

137Cs isotope - New England Nuclear standard NE-018: 10 microcuries as CsCl in 1.0 N HCl.

Methylamine Hydrochloride. Eastman Organic Chemical, white label. Asbestos - acid-washed selected fiber. Fisher Scientific Co.

BAMBP _ K & K labs #23937

(4_<u>sec</u>_butyl_2_(**d**_methylbenzylphenol)

Preparation of Ammonium Molybdophosphate (AMP)

The exchange properties of the AMP used, known to depend on the method of preparation²⁰, were shown to be comparable to the material used by previous workers. This was ascertained by equilibration experiments in which reported D values were duplicated.

Preparation

A saturated solution of phosphomolybdic acid $(20 \text{ Mo0}_3 \cdot 2\text{H}_3\text{PO}_4$ • 48H_20) was prepared in about 2 liters of distilled water. The acid was only slightly soluble but the addition of about 5 ml of comed H_3PO_4 and heating improved the solubility. A saturated solution of NH_4NO_3 (ca 15 <u>M</u>) was prepared and added to the phosphomolybdic acid solution in excess. The AMP precipitate was filtered with (Whatman #42) paper under suction. The precipitate was washed repeatedly with 0.1 <u>M</u> NH_4NO_3 until the supernatant liquid showed a pH greater than 3. The AMP was then dried at 100° C for 8 hours. The fine yellow powder produced was passed through a 200-mesh screen. The particle size was less than 74 microns.

Determination of Distribution Ratios (D)

Sample Preparation

<u>Sample type I</u>: 200 ml of fresh urine was evaporated to dryness with 5 ml concentrated HNO₃. Ashing with addition of small portions of concentrated HNO_3 was repeated until a white residue remained. The residue was then dissolved with 200 ml of 0.1M NH_LNO_3 and the pH adjusted to 1 with HNO_3 .

<u>Sample type II</u>: 200 ml of fresh urine was ashed as for type I. The residue was then dissolved with 100 ml of 0.1 \underline{M} NH₄NO₃ and the pH adjusted to 1.

<u>Sample type III</u>: 200 ml of fresh urine was ashed as for type I. The residue was then dissolved in 100 ml of distilled water and the pH adjusted to 1 with HNO₃.

<u>Sample type IV</u>: raw urine was made 0.1 M in NH_4NO_3 and the pH adjusted to 1 with HNO₃.

<u>Sample type V</u>: raw urine was adjusted to pH = 1 with HNO_3 . <u>Sample type VI</u>: 0.1M NH_4NO_3 was adjusted to pH = 1 with HNO_3 .

Equilibration Time

Since it was not known how long the equilibration would take between cesium and the AMP a series of samples of type I were shaken for 60, 30, 15, and 1 minute with 0.5 g AMP. Analysis of the aqueous phase showed the D values to remain essentially constant throughout these times. Thereafter all samples were shaken with AMP for 1 minute for equilibration.

Procedure

A 10-ml sample of each type was pipetted into 250-ml glassstoppered Erlenmeyer flasks; 0.5 g of AMP was then added to each flask. Standard amounts (activities) of ⁸⁶Rb and ¹³⁷Cs were added to the flasks with the aid of a micropipette. (The ⁸⁶Rb and ¹³⁷Cs activities were added separately and evaluated individually.) Each sample was shaken and the phases were allowed to separate for several hours. Since adequate separation was possible, resort to filtration or centrifugation was not necessary. A 5-ml aliquot of the clear aqueous phase was transferred to a soft glass test tube for counting.

Counting Procedure

Energy spectra of the ⁸⁶Rb and ¹³⁷Cs (Figures 2, 3) standard solutions were obtained with a 256-channel gamma spectrometer, equipped with a well-type 3" X 3" NaI crystal. The spectra revealed adequate purity of the isotopes. The ¹³⁷Cs samples were counted for 2000 seconds (live) and the integral counts were obtained by arithmetic summation of the counts in channels 140-180 (4KEV/channel). These counts were compared to a known activity of ¹³⁷Cs placed in 5 ml of sample type I. For the ⁸⁶Rb samples the same procedure was followed except the summation was taken from channels 196-236. Counting statistics as discussed in reference (7) were taken into consideration but the counting error, being smaller than other errors affecting D values, was finally neglected.

Data

 $D_{\mathbf{x}} = \frac{\text{Activity per gram AMP}}{\text{Activity per ml of sample}}$

TABLE III

Sample type	D (Cs)	D (Rb)	<u>D (Cs)</u> D (Rb)	•
II	2100	98	21	
III	4200;4030	358;417	12;10	
IV	1840;1890	82;97	22;20	
V	2966	338	9	
VI	6770	277	24	

EXPERIMENTAL DISTRIBUTION VALUES

Discussion of Experimental Results

It is of interest to compare these values with those in the literature. Only two are available on sample type VI¹⁹, where the reported values of D(Cs) and D(Rb) are 6000 and 230 respectively. The correlation is satisfactory, however, a value of 6000 may be suspected of having undergone rough rounding off. Perhaps the investigators involved experienced such variability in their values that they rounded them off to 2 significant figures, and the result should have been reported as 6.0×10^3 . It is interesting to note the rather large D values for these types of samples. Since D values drop roughly from 6000 to 1500 in going from $0.1M \, \text{NH}_4 \text{NO}_3$ to sea water, one would expect an increasing salt content to further reduce the D value. This is evidently not the case and each sample type must be evaluated individually.

The dependence of D(Cs) upon the ammonium ion concentration has been determined by Krtil^{13} . A plot of log D vs log $\mathrm{C}_{\mathrm{NH}_{4}\mathrm{NO}_{3}}$ yields a straight line with a slope of -1. The ammonium ion dependence is borne out in the above samples and those samples without added NH_{4}^{+} yield significantly higher D values. The analysis of sample type III appears to be the most advantageous. After ashing, the concentration of the sample is doubled, no $NH_{4}NO_{3}$ is required, and the method yields a large D value. The analysis of raw urine is not normally performed for esthetic reasons, but from the D values above there is no reason why this analysis cannot be done in an emergency.

It should also be noted that even though AMP prepared by different techniques may yield variable D values²⁰ and the same effect is observed if different amounts of AMP are taken for batch equilibration, those variations will be within one order of magnitude.

Elution

Prior to elution of rubidium and cesium, it was found that use of a column 1 cm² in cross section could not be packed well and thus led to very low flow rates. A 6.25-cm² column was substituted in which flow rates could be maintained between 3 and 4 ml per minute. Also it was found that the parameter B had been overestimated. The bed height after settling was 6.5 cm, which corresponds to a total bed volume of about 41 cm³. Since 4.3 cm³ of dry AMP and asbestos was added, B becomes 0.105. The material does appear to pack better in large columns.

Elution Procedure

To a 6.5-cm² column 10 grams of AMP and 5 grams of asbestos were added as a slurry in 0.1 <u>M</u> NH₄NO₃. The column was shaken for complete

mixing and allowed to settle overnight. After the column was drained, activities of 86 Rb and 137 Cs were washed onto it with 100 ml of sample type III. The ^{86}Rb was eluted with 3M NH_4NO_3 and the ^{137}Cs eluted with saturated (ca. 15M) NH_4NO_3 . The elution curve is shown in Figure 4. The eluate was collected in tubes 1-140, each tube representing 5 ml. The radioactivity in the tubes were counted in a multichannel analyzer already described and the activities of each were monitored at the gamma peak maximum for each isotope. The results show an excellent separation of Rb and Cs. The curve is also evidence of complete 40 K removal, the main offender in biological samples. The Rb tail persisted when using $3\underline{M}$ NH_UNO₃ and fell to background only after the addition of saturated $\rm NH_4NO_3.$ This suggests that 3M $\rm NH_4NO_3$ is a little too weak for the purpose and another elution was carried out using $4\underline{M}$ $\rm NH_4NO_3$. The results of this elution are shown in Figure 5. This resulted in improvement and 4M was used in subsequent separations. A comparison of these elution parameters with design predictions is now in order.

TABLE IV

Measured Values			Estimated Values	Calculated Values		
V(max)(Rb)	=	70 ml		20.7		
$V(\max)(Cs)$	=	145 ml	cana (anco	96.8		
D _v (Cs)	=	33	and and	444		
D _v (Rb)	=	15	(21)	8.6		
Ø	=	0.105	0.5			

A COMPARISON OF MEASURED AND CALCULATED ELUTION PARAMETERS





Considering the many assumption and overdesign in the theoretical calculation, the correlation is satisfactory, and points out the usefulness of prior calculations, following Helfferich's pattern, before performing an ion exchange elution.

New Eluting Agents

The large volume of saturated ${\rm NH}_4{\rm NO}_3$ required for the elution of Cs is clearly undesirable. This rather viscous solution would present significant difficulties in final counting. Due to the strong size dependence of AMP and exchangeable ions, it was thought that ions derived from substituted amines larger than $\mathrm{NH}_{l_{L}}^{+}$, might displace Cs more effectively. The column was prepared as described above and standard activities were added, followed by 100 ml of sample type III. A saturated solution of methylamine hydrochloride was added to the column and the eluant monitored for 137Cs activity. In the 200 ml of eluant that was collected, no ¹³⁷Cs was detected; thus indicating the unsuitability of this material as an eluting agent. Sizewise, hydroxyl ammonium ion would qualify as a potential eluting agent, but its strong reducing action prevents its use in the presence of AMP. It is possible that these ions are too large to fit into the crystal lattice space between two AMP molecules. Their ionic diameters are not known, but one can estimate the diameters of the nonsolvated free bases from the reported bond lengths.³



HONH2

These diameters would permit incorporation into the AMP lattice, but once protonated and solvated their sizes are apparently larger, since they did not elute Cs after several column volumes. It should be noted that salts of the heteropolyacids have been prepared from organic bases²⁰ such as 8-quinolinol. Although Cs has shown a favorable distribution ratio in batch determination employing long contact times with AMP, the exchange is believed to take place on the surface only and the Cs is readily eluted with a few column volumes of dilute ammonium nitrate solution. So it is likely that other salts could be formed with different organic bases, but the problem is fitting an ion into the AMP lattice once it has formed, and not the formation of a new ion exchange material.

In reviewing all known metal cationic diameters it is noted that only francium has a diameter larger than Cs. Unfortunately Fr does not exist as a stable isotope and the use of active Fr would interfere with the final Cs determination unless counted by gamma spectroscopy, which would defeat the purpose of this work. Another possibility considered was the use of barium as an eluting agent. An exchange between barium and AMP has been observed²¹ and is believed to take place via a monovalent hydroxide complex:

 $Ba^{++} + H_2 0 \Longrightarrow Ba(OH)^+ + H^+$

and the exchange reaction is:

$$\operatorname{RNH}_{4}$$
 + $\operatorname{Ba(OH)}^{+} \rightleftharpoons \operatorname{NH}_{4}^{+}$ + R $\operatorname{Ba(OH)}$

This mechanism explains the observed dependency of barium absorption on both hydrogen and ammonium ion concentrations²¹. Such a mechanism would require both low H⁺ and NH₄⁺ concentrations since the stability constant for the Ba(OH)⁺ complex is quite low (log K = 0.7 @ pH = 3)²¹. Elution curves show that barium can be eluted with dilute NH₄NO₃; therefore barium cannot be used to elute potassium, rubidium, or cesium.

In summary, the use of new eluting agents for cesium on AMP did not appear fruitful and other means had to be used to collect cesium in a form suitable for counting.

Solvent Extraction Using BAMBP

BAMBP extraction systems have been studied by Oak Ridge scientists⁴,²⁴ with a view toward applying them to the removal of fission products from nuclear fuel. It was thought that this reagent could be utilized for the extraction of Cs from strongly alkaline solutions containing as much as 10 grams of AMP. The dissolution of AMP by a strong base such as NaOH would reduce the time and volume required to remove the Cs as already pointed out. BAMBP can be classified as a hindered phenol, with a slight solubility in aqueous alkali, limited association through hydrogen bonding, and predominance of the dimer as the principal species. Alkali metal extraction by BAMBP involves cation exchange with a phenolic proton to form an alkali phenolate, which is solvated with additional phenol molecules. At high hydroxide concentrations, water is also extracted since it competes with the phenol molecule as the solvating species. The degree of extraction of the alkali ions decreases rapidly with decreasing ionic size. Extraction measurements⁴ have shown an inverse first-power dependence upon the H^+ concentration in the pH range 7.6-10.6 and a third-power dependence upon the concentration of BAMBP in the range 0.1-1 M. The extraction depends upon the equilibrium between the monomer and dimer of BAMBP, which, as expected, has been found to depend upon the concentration of BAMBP.

Monomer: $M^+ + 4ROH + OH^- \rightleftharpoons MOR.3ROH + H_2O$ Dimer: $M^+ + 2(ROH)_2 + OH^- \rightleftharpoons MOR.3ROH + H_2O$

At formal concentrations of BAMP greater than 0.6 <u>M</u> the dimer predominates. One also sees here that increasing the hydroxide concentration would shift the equilibrium to the right. The ability of BAMBP to extract Cs preferentially over the other alkali metals is thought to be due to the smaller hydration energy that has to be overcome. The mole ratio Cs:BAMBP has been found to be 1:4 in the organic phase⁴, as shown by the equations.

The distribution of Cs and Rb between 0.5 M BAMBP in kerosene and 1 M NaOH was investigated. In Figure 6, the distribution ratios are shown vs. pH for the above system. A value of 0.5 was found for the slopes of these lines. The use of 6 M NaOH solutions resulted in a dark viscous third phase with reduced distribution ratio. This phase is thought to have contained hydrated sodium ions and BAMBP. The BAMBP was evidently consumed in the extraction of sodium and an insufficient concentration for Cs extraction was left. Whether BAMBP would extract Cs in the presence of a large amount of AMP was then investigated.



It was found that 20-30 ml of 6 <u>N</u> NaOH was sufficient to dissolve 10 grams of AMP and a series of these samples were prepared with the pH adjusted from 7 to 13. It was found that Cs was indeed extracted, the bulk of the AMP being left in the aqueous phase. There was no third phase observed. The distribution ratio was at maximum for both Cs and Rb at pH 12.5, with values of 10 and 1 respectively. Undoubtedly this extraction can be optimized by increasing the BAMBP concentration and using diluents other than kerosene. For instance it has been reported⁴ that cyclohexane provides larger D' values. For the present work, the use of 0.5 M BAMBP in kerosene and the aqueous phase at a pH of 12.5 appeared adequate. A distribution ratio of 10 for Cs requires multiple extraction to achieve adequate recovery, however. From the relationship:

$$\% \text{ extracted} = \begin{bmatrix} D' \\ D' + \frac{Vaq}{Vorg} \end{bmatrix} X 100$$

where D' = Distribution ratio = $\frac{\mathcal{E}[M^+]}{\mathcal{E}[M^+]} \text{ org}$
 $\mathcal{E}[M^+] aq$

and V_{aq} = volume of aqueous phase V_{org} = volume of organic phase

the number of extractions required for a certain recovery can be calculated.

$$\frac{W_n}{W_o} = \left(\frac{V_{aq}}{D V_{org} + V_{aq}} \right)^n$$

where W_n^{\dagger} = weight of solute remaining after n extractions

 W_{o} = initial weight of solute in aqueous phase.

Thus, if the desired recovery is 98%, D is 10 and if V and V are

 $e_{i} \boldsymbol{\psi}^{i} \boldsymbol{x}^{i} = \sum_{j=1}^{n} \left(\mathbf{x}^{i} - \mathbf{x}^{j} \right)^{T} \left(\mathbf{x}^{i} - \mathbf{x}^{i} \right)^{T}$

40 and 10 ml respectively, the number of extractions becomes three.

Recommended Procedure

To a measured volume of urine (up to 2 liters have been handled successfully), add 20 ml of concentrated HNO3. Dissolve the white residue in distilled water (one-half the original volume) and adjust the solution to a pH of 1, using HNO3. Prepare the ion exchange column by using 10 grams of AMP and 5 grams of asbestos. Make this mixture into a slurry with $0.1\underline{M}$ NH₄NO₃ and pour onto a 6.25-cm² by 50-cm column. A wad of glass wool at the bottom and finally one at the top of the column were found to be beneficial in preventing loss of AMP and entrainment in the solution to be added. Allow the AMP and asbestos to settle for several hours, then add the sample to the column in portions, maintaining a flow rate through the column of about 3 ml per minute. Allow the column to drain, then elute interfering elements using two 100-ml portions of 4 M $\rm NH_4NO_3$. Add 6 M NaOH in 10-ml portions until the AMP is completely dissolved. Collect each portion in a 125-ml separatory funnel. It should be noted that NaOH will dissolve the AMP by percolating it through the undisturbed column but smaller volumes can be obtained by breaking up the partly dissolved bed with a glass rod, to allow greater contact. The solution is now adjusted to a pH of 12.5 using HNO_3 or NaOH as required. Extract the sample 3 times with 10-ml portions of 9.5 M BAMBP in kerosene. For a Cs measurement by beta count, the organic phases can be combined and back-extracted into strong acidic solutions. Final transfer is to a steel counting planchet using a small dropper, each portion being evaporated under an infra red lamp. Alternatively,

if gamma activity counting is employed, the total organic phase can be counted directly. The recovery using this procedure was found to be 93% of the activity added initially. The small Cs loss is attributed to solution transfers and possibly adsorption of Cs on glass or asbestos surfaces.

The gamma energy spectra of 137 Cs following analysis is compared with the energy spectra of the 137 Cs standard solution in Figure 7.



CHAPTER V

CONCLUSIONS AND RECOMMENDATIONS

An analytical procedure has been developed for the determination of ¹³⁷Cs in biological materials. Application of the procedure was demonstrated on human urine. In arriving at a final procedure, the literature was reviewed for ion exchange and solvent extraction systems that might be applied with modifications to the problem at hand. The ion exchange material chosen for study was ammonium molybdophosophate since its usefulness in separating the alkali metals had been demonstrated on samples of low ionic strength. The structure of AMP was considered in order to determine a basis for its exchange properties. The ion exchange properties of AMP were found to depend upon ionic size of an exchangeable univalent metal, $\mathrm{NH}_{\mathrm{L}}^+$ solution concentration, and pH. A column design calculation was made in order to determine the necessary conditions to effect a good separation of cesium and rubidium. Even though rubidium concentrations are quite low in biological samples, a good separation would require complete removal of ⁴⁰K and other interfering activities. The theoretical calculations were useful and a trial elution of ¹³⁷Cs and ⁸⁶Rb resulted in good correlation with them. Distribution ratios were obtained using raw and ashed urine samples and were found to be surprisingly large in comparison with published values. Column parameters and procedures were adjusted to give a maximum rubidium

contamination of 0,01%. Several elutions were performed showing excellent separation. Because of the large D values obtained for cesium, unsuitably large quantities of saturated $\text{NH}_{\mu}\text{NO}_{3}$ were required for elution. Other eluting agents were considered and tried in order to improve the efficiency of elution of cesium but were not fruitful. The solvated methylammonium ion is possibly too large to fit into the AMP lattice. In order to collect the cesium from the column in reasonable volume, a BAMBP extraction system was investigated. The AMP containing cesium was dissolved in strong base and the solution equilibrated with 0.5 M BAMBP in kerosene. Fortunately, AMP did not interfere. Rubidium was extracted only one-tenth as much as Cs, which further reduces any possible rubidium contamination. A final procedure was described for urine samples. It is believed that this method can be applied to a host of biological and environmental samples, and these applications can be tested quickly by batch equilibration of AMP with the sample in question.

Recommendations

- AMP is a valuable ion exchange material for the alkali metal ions. Its application to other biological and environmental samples should be investigated.
- 2. Large sample volumes take a long time for percolation through the column so ways to improve flow rates without loss of resolution should be investigated.
- 3. The size and solvation energies of exchangeable ions on AMP should be investigated and a systematic search for new eluting agents be continued.

4. The solvent extraction of Cs from strongly basic solutions containing AMP should be studied in an effort to improve the distribution ratio. It is believed that there are optimum hydroxide ion and BAMBP concentrations as well as a host of of non-polar solvents to try as the BAMBP carrier.

A SELECTED BIBLIOGRAPHY

- 1. A. J. Bradley and J. W. Illingsworth, Proc. Roy. Soc., Ser. A 157.
- 2. J. A. Campbell, "Chemical Systems", W. A. Freeman and Co., 1970, 1011.
- 3. C. M. Day and J. Selbin, "Theoretical Inorganic Chemistry", Reinhold Book Publishers, New York, New York, 1962.
- 4. B. Z. Egan, R. A. Zingaro, and B. M. Benjamin, J. Inorg. Nucl. Chem., <u>4</u>, 1055 (1965).
- 5. W. W. Flynn, Anal. Chim. Acta, <u>50</u>, 365 (1970).
- 6. A. J. R. DaFonseca, M. H. D. deMatos, and M. M. daCruz, Rev. Port. Quim., <u>6</u>, 17 (1954).
- 7. G. Friedlander and J. W. Kennedy, "Nuclear and Radiochemistry", John Wiley and Sons, New York, 1955.
- 8. E. Glueckauf, Trans. Faraday Soc., <u>51</u>, 34 (1955).
- 9. Peter Gray, "Encyclopedia of Biological Sciences", Reinhold, New York, 1961.
- F. Helfferich, "Ion Exchange", McGraw-Hill Book Co., New York, 1962.
- 11. J. W. Illingsworth and J. F. Keggin, J. Chem. Soc., 1935, 575.
- 12. J. F. Keggin, Proc. Roy. Soc., Ser. A 144, 75 (1934).
- 13. J. Krtil, J. Inorg, Nucl. Chem., <u>24</u>, 1139 (1962).
- 14. P. J. Magno, "Analysis of Environmental Samples", NERHL-64--1 (1964).
- 15. A. Morgan and G. M. Arkell, Health Phys., 2, 857 (1963).
- 16. A. Morgan and G. M. Arkell, Nature, <u>191</u>, 1100 (1961).
- 17. A. Morgan and G. M. Arkell, AERE_R_3675 (1961).
- 18. B. Shleien, Health Phys., <u>18</u>, 267 (1970).

- 19. J. Van R. Smit, Nature, <u>181</u>, 1530 (1958).
- 20. J. Van R. Smit, J. J. Jacobs, and W. Robb, J. Inorg. Nucl. Chem., <u>12</u>, 95 (1959).
- 21. J. Van R. Smit, W. Robb, and J. J. Jacobs, J. Inorg, Nucl. Chem., <u>12</u>, 104 (1959).
- 22. J. Van R. Smit, W. Robb, and J. J. Jacobs, Nucleonics, <u>17</u>, 116 (1959).
- 23. J. Van R. Smit, and W. Robb, J. Inorg. Nucl. Chem., <u>26</u>, 509 (1964).
- "Chemical Technology Division Annual Report for period ending May 31, 1963." Oak Ridge National Laboratory, ORNL 3452 (1963).

ATIV S

Robert Jerrel Everett

Candidate for the Degree of

Master of Science

Thesis: THE SEPARATION AND DETERMINATION OF ¹³⁷Cs IN BIOLOGICAL MATERIALS

Major Field: Chemistry

Biographical:

- Personal Data: Son of Mr. and Mrs. E. B. Everett, Artesia, New Mexico. Married. Three children.
- Education: Graduated from high school at Artesia, New Mexico, June, 1948. Graduated from University of New Mexico, Albuquerque, New Mexico, June, 1956, with Bachelor of Arts Degree in Chemistry and Biology. Admitted to the Graduate College of Oklahoma State University, Stillwater, Oklahoma, in January, 1970. Completed requirements for the Master of Science Degree in May, 1971.
- Professional Experience: 1956-1969, Sandia Corporation, Albuquerque, New Mexico. Held positions in Analytical Chemistry, Health Physics, and Industrial Hygiene. Supervisor of Environmental Health Program and Radiological Effects Division, Aerospace Nuclear Safety. Presently teaching in the Department of Radiation & Nuclear Technology, Oklahoma State University.