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(12) **United States Patent**  
**Prade et al.**(10) **Patent No.:** **US 8,847,031 B2**(45) **Date of Patent:** **Sep. 30, 2014**(54) **THERMOCELLULASES FOR  
LIGNOCELLULOSIC DEGRADATION**(75) Inventors: **Rolf A. Prade**, Stillwater, OK (US);  
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State University**, Stillwater, OK (US)(\*) Notice: Subject to any disclaimer, the term of this  
patent is extended or adjusted under 35  
U.S.C. 154(b) by 560 days.(21) Appl. No.: **13/003,183**(22) PCT Filed: **Jul. 9, 2009**(86) PCT No.: **PCT/US2009/050080**

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(2), (4) Date: **Apr. 8, 2011**(87) PCT Pub. No.: **WO2010/006152**PCT Pub. Date: **Jan. 14, 2010**(65) **Prior Publication Data**

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2008.(51) **Int. Cl.****A01H 5/00** (2006.01)**C12N 9/42** (2006.01)**C07H 21/04** (2006.01)(52) **U.S. Cl.**USPC ..... **800/320.1**; 435/209; 536/23.2(58) **Field of Classification Search**USPC ..... 800/320.1; 435/209; 536/23.2  
See application file for complete search history.(56) **References Cited**

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database, alignment with SEQ ID No. 56 of WO 03/018766 A2,  
performed on Jul. 5, 2013.\*Result 15, search of instant SEQ ID No. 2 in the DNA GenEmbl  
database, alignment w/ *P. furiosus* eg1A gene used in WO  
03/018766A2, performed on Jul. 5, 2013.\*

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(57)

**ABSTRACT**Thermostable cellulase enzyme systems comprising at least  
one each of a thermostable endoglucanase, an exo-proces-  
sive-endoglucanase, and a  $\beta$ -glucosidase carry out the com-  
plete, coordinated hydrolysis of crystalline cellulose to  
monomeric glucose.**3 Claims, 28 Drawing Sheets**

High-temperature operating and thermo-stable cellulases

PROTEIN	Source		Physical Properties						Functional Properties				
	Locus	Codon Usage	CaZy	MW d	pI	Charge pH7	T <sup>o</sup> °C	pH range	Mode of Operation	Specific Activity			
										<sub>sw</sub> Avicel	<sub>av</sub> Cellulose	Paper	pNPG
Tcel1	<i>o-eglA</i>	corn optimized	GH12	34,005	4.80	-13.10	102	7 - 8	exocellulase	63.4	13.6	10.5	-
Tcel2	<i>petroB</i>	bacteria	GH12	31,816	4.77	-13.30	98	6 - 7	exocellulase	8.1	2.2	2.9	-
Tcel3	<i>ph1171</i>	archaea	GH5	51,930	6.47	-3.60	94	6	exocellulase	48.5	8.4	8.6	-
Tcel4	<i>o-E1</i>	rice optimized	GH5	59,980	7.05	0.30	95	5 - 6	endocellulase	6.8	2.2	4.1	-
Tcel5	<i>petroA</i>	bacteria	GH12	38,226	5.58	-6.60	96	5 - 6	endocellulase	34.1	6.8	8.0	-
Tcel6	<i>zp#4</i>	corn optimized	GH12	31,818	5.66	-5.00	85	5 - 6	endocellulase	20.6	5.1	5.1	-
Tcel7	<i>Tpet0898</i>	bacteria	GH3	81,243	5.38	-16.90	98	5	beta-glucosidase	0.8	0.8	-	69.4
Tcel8	<i>Tpet0952</i>	bacteria	GH1	51,509	5.84	-9.10	92	6 - 7	beta-glucosidase	1.7	1.5	-	60.9
Tcel9	<i>g12#3</i>	corn optimized	GH12	45,059	6.16	-2.20	85	5 - 6	endocellulase	5.2	1.4	7.5	-
Tcel10	<i>ph0746</i>	archaea	GH65	85,598	7.80	4.30	94	6	endocellulase	4.9	1.5	3.9	-

v

<sub>sw</sub>AVICEL Phosphoric acid swollen Avicel, <sub>av</sub>Cellulose, Avicel (SigmaCell): Cellulase specific activity, mM reducing sugar/mg protein/day at 85 °C, pH 6  
Beta-glucosidase specific activity, nM pNug protein/min

(56)

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A.

ATGATCTATTTTGTGAGAAATACCACACCTCAGAAGACAAATCCACAAGCAATACCTC  
CTCAACCCCCCTCAAACGACACTTAGCACAACAAAGGTTCTCAAAATTCGGTATCCTG  
ACGACGGCGAATGGCCTGGCGCTCCCATAGACAAAGACGGCGACGGAAATCCTGAGTTC  
TATATCGAAATCAACCTCTGGAACATACTCAACGCGACTGGATTTCGAGAGATGACCTA  
TAACTTGACATCTGGCGTTCTCCATTACGTTCAACAACCTCGATAATATCGTTCTCCGCG  
ATCGCTCAAACCTGGGTACATGGCTATCCTGAAATTTTTTACGGCAATAAACCCCTGGAAC  
CGGAATTATGCCACCGACGGCCCGATCCCTCTCCCCAGTAAAGTTTCCAATCTCACAGA  
CTTTTACTTGACTATCTCCTACAAGCTTGAACCAAAGAACGGACTCCCTATAAATTTTG  
CAATCGAATCTTGGCTTACTAGAGAAGCATGGCGCACTACTGGAATCAACTCCGATGAA  
CAGGAAGTAATGATCTGGATTTACTATGACGGACTCCAACCAGCCGGTTCCAAGGTGAA  
AGAAATCGTTGTACCTATAATCGTTAA'GGCACCCAGTTAATGCTACCTTCGAAGTGT  
GGAAAGCTAATATCGGATGGGAATACGTTGCCTTTAGAATCAAGACACCAATTAAGAA  
GGAACCGTGACAATCCCCTACGGTGCATTCATTAGCGTAGCTGCTAACATTTCTTCCCT  
CCCAAATTACACAGAACTTTACCTGGAAGACGTTGAGATAGGCACAGAGTTTGGAACAC  
CTTCAACTACTAGCGCACATCTCGAATGGTGGATTACTAACATTACCCTCACCCCACTT  
GATCGTCCCCTGATCTCC

B.

MIYFVEKYHTSEDKSTSNTSSTPPQTTLSTTKVLKIRYPDDGEWPGAPIDKDGDGNPEF  
YIEINLWNILNATGFAEMTYNLTSGVLHYVQQLDNIVLRDRSNVHGYPEIFYGNKPWN  
ANYATDGPIPLPSKVSNLTDYFLTISYKLEPKNGLPINFAIESWLTREAWRTTGINSDE  
QEVMIWIYYDGLQFAGSKVKEIVVPIIVNGTPVNATFEVWKANIGWEYVAFRIKTPIKE  
GTVTI PYGAFI SVAANI SSLPNYTELYLEDVEIGTEFGTPSTTSAHLEWITNITLTP  
DRPLIS

Figure 1A-B

A.

ATGAGGTGGGTAGTTCTTCTGATGGTGGCGTTTTCTGCTCTGCTCTTTTCCTCCGAGGT  
GGTTCTCACGAGCGTTGGCGCAGCGGATATCTCCTTCAACGGATTTCCCGTCACCATGG  
AGCTCAACTTCTGGAACATAAAGTCGTATGAGGGAGAAACGTGGCTCAAATTCGATGGA  
GAAAAGGTTGAGTTCTACGCGGATTTGTACAACATCGTTCTTCAGAATCCAGACAGCTG  
GGTGATGGATATCCGGAGATCTACTACGGTTACAAGCCCTGGGCGAGTCACAACAGCG  
GTGTTGAATTTCTTCTGTGAAGGTGAAAGATCTTCCGGATTTCTACGTGACTCTTGAT  
TACTCGATCTGGTACGAAAACAATCTGCCTATCAACCTTGCAATGGAAACATGGATCAC  
GAAAAGCCCCGACCAGACTTCTGTTTTCTTCGGGTGATGCGGAGATCATGGTTTTGGTTTT  
ACAACAACGTTCTGATGCCCGGCGGTGAGAAAGTGGATGAGTTCACCACAACAGTTGAG  
ATAAACGGAGTGAAGCAGGAAGCAAAATGGGATGTTTACTTCGCACCGTGGAGCTGGGA  
TTACCTTGCCTTCAGACTGACAACACCGATGAAAGAAGGAAAGGTGAAGTTCAACGTGA  
AGGACTTCGTTTCAGAAAGCCGCGGAAGTTGTCAAAAAGCACTCAACGAGAATAGACAA  
TTCGAAGAGCTGTATTTCTGCGTCTGGGAGATCGGGACGGAATTTGGAGATCCAAACAC  
ACAACGGCAAAATTCGGCTGGACCTTCAAAGACTTCTTCGTCGAAGTTGTAAAATAA

B.

MRVVLLMVAFSALLEFSSEVVLTSVGAADISFNGFPVTMELNFWNIKSYEGETWLKFDG  
EKVEFYADLYNIVLQNPDSWVHGYPEIYYGYKPWASHNSGVEFLPVKVKDLPDFYVTL  
YSIWYENLPI NLAMETWITKSPDQTSVSSGDAEIMVWFYNNVLMPPGQKVDEF TTTVE  
INGVKQEA KWDVYFAPWSWDYLA FRLTTPMKEGKVKFNVKDFVQKAAEVVKKHSTRIDN  
FEELYFCVWEIGTEFGDPNTTAKFGWTFKDFVEVVK

Figure 2A-B

A.

ATGGAGGGGAATACTATTCTTAAAATCGTACTAATTTGCACTATTTTAGCAGGCCTATT  
CGGGCAAGTCGTGCCAGTATATGCAGAAAATACAACATATCAAACACCGACTGGAATTT  
ACTACGAAGTGAGAGGAGATACGATATACATGATTAATGTCAACCAGTGGAGAGGAACT  
CCCATTTCATCTCTTTGGTGTAAACTGGTTTGGCTTTGAAACACCTAATCATGTAGTGCA  
CGGACTTTGGAAGAGAAAAGTGGGAAGACATGCTTCTTCAGATCAAAGCTTAGGCTTCA  
ATGCAATAAGACTTCCTTTCTGTACTGAGTCTGTAAAACCAGGAACACAACCAATTGGA  
ATAGATTACAGTAAAAATCCAGATCTTCGTGGACTAGATAGCCTACAGATTATGGAAAA  
GATCATAAAGAAGGCCGGAGATCTTGGTATCTTTGTCTTACTCGACTATCATAGGATAG  
GATGCACTCACATAGAACCCCTCTGGTACACGGAAGACTTCTCAGAGGAAGACTTTATT  
AACACATGGATAGAGGTTGCCAAAAGGTTTCGGTAAGTACTGGAACGTAATAGGGGCTGA  
TCTAAAGAATGAGCCTCATAGTGTACCTCACCCCAGCTGCTTATACAGATGGTACCG  
GGGCTACATGGGGTATGGGAAACCCTGCAACCGATTGGAACCTGGCGGCTGAGAGGATA  
GGAAAAGCGATTCTGAAGGTTGCCCTCATTTGGTTGATATTCGTGGAGGGGACACAATT  
TACTAATCCGAAGACTGACAGTAGTTACAAATGGGGCTACAACGCTTGGTGGGGAGGAA  
ATCTAATGGCCGTAAAGGATTATCCAGTTAACTTACCTAGGAATAAGCTAGTATACAGC  
CCTCACGTATATGGGCCAGATGTCTATAATCAACCGTACTTTGGTCCCCTAAGGGTTT  
TCCGGATAATCTTCCAGATATCTGGTATCACCCTTGGATACGTAAAATTAGAAGTAG  
GATATTCAGTTGTAATAGGAGAGTTTGGAGGAAAATATGGGCATGGAGGCGATCCAAGG  
GATGTTATATGGCAAAAATAAGCTAGTTGATTGGATGATAGAGAATAAATTTTGTGATTT  
CTTTTACTGGAGCTGGAATCCAGATAGTGGAGATACCGGAGGGATTCTACAGGATGATT  
GGACAACAATATGGGAAGATAAGTATAATAACCTGAAGAGATTGATGGATAGTTGTTCC  
AAAAGTTCTTCAAGTACTCAATCCGTTATTCCGAGTACCACCCCTACAAAGTCAAATAC  
AAGTAAGAAGATTTGTGGACCAGCAATTCTTATCATCCTAGCAGTATTCTCTCTCTCT  
TAAGAAGGGCTCCCAGGTAG

B.

MEGNTILKIVLICITILAGLFGQVVPVYAENTTYQPTGIYYEVRGDTIYMINVTSGEET  
PIHLFGVNWFGFETPNHVHGLWKRNWEDMLLQIKSLGFNAIRLPFCTESVKPGTQPIG  
IDYSKNPDLRGLDSLQIMEKIKKAGDLGIFVLLDYHRIGCTHIEPLWYTEDFSEEDFI  
NTWIEVAKRFGKYWNVIGADLKNEPHSVTSPPAAAYTDGTGATWGMGNPATDWNLAERI  
GKAILKVAPHWLI FVEGTQFTNPKTSSYKWGYNAWGGNLMVVDY PVNLPRNKLVS  
PHVYGPVYVYQPYFGPAKGFDPDNLPIWYHHFGYVKLELGYSVVIGFEGGKYGHGGDPR  
DVIWQNKLVDMWMIENKFCDFYWSWNPDSGDTGGILQDDWTTIWEDKYNNLKRMLDSCS  
KSSSSTQSVIRSTTPTKSNTSKKICGPAILIILAVFSLLLRRAPR

Figure 3A-B

A.

ATGGAAATCAAGCTCTTCTGCGTGTTTATCGTGTTTCATCATCCTCTTCTCCCCTTTCGT  
GATTGCACTCTCGTATCCAGATGTTAACTATACTGCCGAGAATGGTATTATCTTCGTGC  
AGAACGTCACCTACGGGTGAGAAGAAGCCACTTTATCTTCACGGAGTGTTCATGGTTTGA  
TTCGAGCTGAAGGACCACGTCGTCTATGGCTTGGATAAACCGAACTGGAAAGATATACT  
CAAGGATGTTAAGCGCTTGGGTTTTAATGCTATCAGGCTTCCCTTCTGCTCTGAAAGCA  
TCCGCCCTGATACGCGCCCTTTCGCCTGAGCGGATAAACTACGAGTTGAACCCCGACTTG  
AAGAATCTGACTTCCCTCGAAATAATGGAGAAGATTATTGAATACGCCAACTCAATCGG  
GCTCTACATACTCTTGGATTATCACCCGATCGGTTGTGAGGAGATCGAACCTCTTTGGT  
ATACCGAGAATTACTCAGAGGAGCAGTATATAAAGGATTGGATCTTCCCTCGCAAAGCGG  
TTCGGGAAGTACCCTAACGTGATAGGAGCTGATATCAAGAACGAGCCGCATGGTGAAGC  
CGGGTGGGGTACGGGAGATGAGCGGGATTTCCGCCTCTTTGCCGAGAAGGTGGGGCGG  
AGATACTCAAGGTGGCCCCACACTGGTTGATATTCGTCGAGGGAACGCAATATACCCAT  
GTCCCGAATATTGATGAGATCATCGAGAAGAAGGGCTGGTGGACATTTTGGGGAGAGAA  
TCTTATGGGAGTTAAGGACTATCCAGTCAGGCTTCCGCGCGGCAAGGTCGTGTAACAC  
CGCATGTCATGGACCATCTGTCTACATGATGGACTACTTCAAGTCGCCAGACTTTCCG  
AACAATATGCCGATAATCTGGGAAACACACTTCGGATACTTGACCGACCTGAATTATAC  
CTTGGTCATAGGCGAGTGGGGTGGCAACTATGAGGGCCTTGACAAGGTGTGGCAAGACG  
CTTTTCGTAAGTGGCTGATTAAGAAGAAGATCTATAACTTCTTCTACTGGTGCCTGAAC  
CCGGAGTCGGGTGACACCGGTGGCATCTTCTCGACGACTGGAAAACCGTTAACTGGGA  
AAAGATGAGGGTTATTTACAGGCTCATCAAGGCGGCGAACCCCGAGTTTGAGGAACCCC  
TTTACATCATTTTGAATACTAACGCGACGACATCTATCCTGGGCGTGGGTGAGAGGATC  
CGGATTTACTGGTACACAAATGGCAAAGTTATTGACTCTAACTTCGCGCATTCAGCGA  
AGGCGAAAATGAACATTACAGTGACGAAGTCCATGACTCTGTACATCATCGTGAAGAAGG  
GCAATCAGACACTGAGGAAGGAACTCAAACGTACGTTATCGGCGGCAATTACGGCTCC  
AATATCTCCACTACCCAGCTGGTTACTCCCAAGAAAGGCGGCGAAAGGATTAGCACCAG  
CCTGAAGCTGGCAATTAGCCTGCTCTTCATTCCTCTTTCGTTTGGTATCTCCTCCGGG  
AGAAGCAT

B.

MEIKLFCVFIVFIILFSPFVIALSYPDVNYTAENGIIFVQNVTTGEEKPLYLHGVSWFG  
FELKDHVVYGLDKRNWKDILKDKVRLGFNAIRLPFCSESIRPDTRPSPERINYELNPD  
KNLTSLEIMEKIEYANSIGLYILLDYHRIGCEEIEPLWYTENYSEEQYIKDWIFLAKR  
FGKYPNVIGADIKNEPHGEAGWGTGDERDFRLFAEKVGREILKVAPHWLI FVEGTQYTH  
VPNIDEIEKKGWTFWGENLMGVKDYVRLPRGKVVSYPHVYGPSVYMMDYFKSPDFP  
NNMPIIWETHFGYLTDLNYTLVIGEWGNYEGLDKVWQDAFVKWLIKKKIYNFFYWCLN  
PESGDTGGIFLDDWKTVNWEKMRVIYRLIKAANPEFEEPLYIILKTNATTSILGVGERI  
RIYWYTNGKVIDSNFAHSSEGEMNITVTKSMTLYIIVKKGNTLRKELKLYVIGGNYGS  
NISTTQLVTPKKGGERISTSLKLAISLLFILLFVWYLLREKH

Figure 4A-B

A.

ATGGAAACGCTCCTCCCTGTAGTCGTGGTCCACGATATTGAGCCAGTTTCAATGCGTCT  
TCAGAGGTACAAGAACAAAAATTCGATAAAAAGAGAAAAGCAGGGATTAATACCCCTGT  
TTTTTTATTTTTGGGTGTATTTAGTTCTATTTGCGAATTTTCAGATTTTGAAATGTAAAC  
ATTTTCATAATAAGATGTTTTCTGGAGGTGATAATGGTGGTACTGATGACAAAACCGGG  
AACATCGGATTTTGTATGGAATGGCATTCCCCTTTCCATGGAGCTGAATCTGTGGAACA  
TAAAGGAATACTCCGGTTCGTAGCTATGAAATTCGACGGTGAAAAGGTAAC TTTCGAC  
GCGGACATTCAGAATCTTTCTCCAAAAGAACCAGAAAGGTACGTTCTCGGTTATCCCGA  
GTTCTATTACGGTTATAAACCCCTGGGAAAAGCACACGGCAGAAGGTTTCGAAACTTCCAG  
TACCTGTTTCCTCTATGAAATCATTTTCCGTGCAAGTTTCTTTTCGATATTCACCACGAA  
CCGTCTCTGCCTTTGAACTTTGCCATGGAAACATGGCTCACAAGAGAAAAGTACCAGAC  
GGAAGCGTCGATCGGCGATGTTGAAATCATGGTCTGGTCTATTTCAACAATCTCACAC  
CAGGGGGCAAAAAGATAGAGGAGTTTACGATTCGGTTCGTGCTGAACGGAGAGAGTGTC  
GAAGGCACCTGGGAACTGTGGCACGCGGAGTGGGGATGGGACTACCTCGCTTTCGGCTT  
GAAGGATCCCGTGAAGAAGGGGAAGGTTGAAAGTTCGACGTGAGGCATTTTCTTGATGCCG  
CCGGGAAAGCTCTTTCGAATTCCACTCGTGTGAAAGATTTTGAAAATCTTTACTTCACC  
GTCTGGGAAATTGGAACCGAGTTTGGAAAGCCCGGAAACAAAGAGCGCGCAATTCGGGTG  
GAAGTTTGAAAAC TTCTCTATTGATCTGGAGGTGAGAGAAATGA

B.

METLLPVVVVHDI EPVSMRLQRYKNKNSIKREKQGLIPLFFYFWVYLVLFANFQILNVN  
IFIIRCFLEVIMVVLMTKPGTSDFFVWNGIPLSMELNLWNIKEYSGSVAMKFDGEKVTFD  
ADIQNLSPKEPERYVLGYPEFYGYKPEKHTAEGSKLPVPVSSMKSFSVEVSFDIHHE  
PSLPLNFAMETWLTREKYQTEASIGDVEIMVWFYFNNLTPGGKKIEEFTIPFVLNGESV  
EGTWELWHAEWGWDYLAFLKDPVKKGRVKFDVRHFLDAAGKALSNSSTRVKDFENLYFT  
VWEIGTEFGSPETKSAQFGWKFFENFSIDLEVRE

Figure 5A-B

A.

ATGTTGAAACTTATTCCACTTGTTAATGGCAATTATAAGTTGATTCAATGGGAGCCACT  
CGGCGGCGTGCACGGAGCAGATATCGAGTGCATACATGTTACCCCAAACGTATGGAACA  
TAGATAAATCATCAGTTGGCACTGTACAGATCGAATATGAGCCCCAAGTTGGCTGTCTT  
CGTTTTTCAATTGATTTCCCGAGGATAAGTATAAGACATAATGTAGGCGTAGCGGCATA  
TTCAGAAGTTATTTACGGACACAAGCCGTGGGGCCCCACCACTTGCATGGACCCCTCAGT  
TCAAGTTCCTTATCAAAGTCAATGAGTCAAAGGACTGTACTCGTATGTAAATTATAAC  
GTTAAATCTAGGTCAACAGATGACTCAATCTTTAATATTGCTTACGATCTCTGGCTTAC  
AACGTCCCAAACCTTACAAACGGACCCCAGCCAGGAGACGTAGAAGTTATGATCTGGT  
TGTACTIONCACGGACAGCGCCCTGCAGGCAGACTCATCGGGGAACTCCGCATGCCGATT  
ACATTTGGGCGATAGTGAGGCGGCACGTGACTTTGAAGTATGGGTGGCTGACACAGGAAT  
AGGAATCGGTGAATGGGCGGTAGTGACCTTCAGAATCAAGGACCCAATAAAGGGCGGTT  
TGATAGGAGTTAACCTCATAAACTACATCGAAAGTGCTTTTAAAACGCTCGAAGAACTC  
AACCCGGTCAAGTGGCGGTACGGCGACCTGCTCAACAAAATCTTAATGGAATTGAATT  
CGGCAGTGAGTTTGGTAATGTCTCCTCAGGAATGATAAACTTAATTGGGAACTCTGCG  
GCCTGAGCCTTGTGAAAGACTCTTCT

B.

MLKLIPLVNGNYKLIQWEPLGGVHGADIECIHVTPNVWNIDKSSVGTQIEYEPQVGCL  
RFSIDFPRISIRHNVGVAAYSEVIYGHKPGWPTTCMDPQFKFPIKVNESKGLYSYVNYN  
VKSRSPDDSI FNIAYDLWLTTSPNL TNGPQPGDVEVMIWLYYHGQRPAGRLIGELRMP  
TLGDSEAARDFEVWVADTGIGIGEWAVVTFRIKDPIKGLIGVNLINYIESAFKLEEL  
NPVKWRYGDLLNKYLNGIEFGSEFGNVSSGMIKLNWELCGLSLVKDSS

Figure 6A-B



A.

ATGATGGGAAAAGATCGATGAAATCCTTTACAGCTGACTATTGAAGAAAAAGTGAACT  
TG TAGTGGGGGTTGGTCTTCCAGGACTTTTTGGAAATCCACATTCCAGAGTGGCAGGTG  
CAGCTGGAGAAACGCATCCTGTTCCGAGGCTTGGAATTCCTTCTTTCGTTCTGGCCGAC  
GGTCCCGCGGGCCTCAGAATAAATCCACAAGAGAGAACGACGAAAACACCTATTACAC  
AACAGCGTTTCTGTTGAAATCATGCTCGCTTCCACCTGGAACAAAAGATCTTCTGGAAG  
AAGTAGGAAAAGCTATGGGAGAAGAAGTCAGGGAATACGGTGTGATGTGCTTCTTGCA  
CCTGCGATGAACATTCACAGGAACCCTCTTTGTGGAAGGAATTCGAGTATTATTTCAGA  
AGATCCTGTCTTTCCGGTGAAATGGCTTCAGCCTTTGTCAAGGGAGTTCAATCTCAAG  
GGGTGGGAGCCTGCATAAAACACTTTGTGCGCAACAACCAGGAAACGAACAGGATGGTA  
GTGGACACGATCGTGTCCGAGCGAGCCCTCAGAGAAATATATCTGAAAGGTTTTGAAAT  
TGCCGTCAAGAAAGCAAGACCCTGGACCGTGATGAGCGCTTACAACAAACTGAATGGAA  
AATACTGTTACAGAACGAATGGCTTTTGAAGAAGGTTCTCAGGGAAGAATGGGGATTT  
GACGGTTTTCGTGATGAGCGACTGGTACGCGGGAGACAACCCTGTAGAACAGCTCAAGGC  
CGGAAACGATATGATCATGCCTGGAAAAGCGTATCAGGTGAACACGGAAAGAAGAGATG  
AAATAGAAGAAATCATGGAGGCGTTGAAGGAGGGAAGACTCAGTGAGGAAGTCTTGAAC  
GAATGTGTGAGAAACATCCTCAAAGTTCTTGTGAACGCGCCTTCTTTAAAGGGTACAG  
GTACTCGAACAAACCGGACCTCGAATCTCACGCGAAAGTTGCCTACGAAGCAGGTGTGG  
AGGGTGTGTCCTTCTTGAGAACAACGGTGTCTTCCATTCGATGAAAGTATCCATGTC  
GCCGTCTTTGGCACCGGTCAAATCGAAAACAATAAAGGGAGGAACGGGAAAGTGAGACAC  
CCATCCGAGATACACGATCTCTATCCTTGAAGGCATAAAAAGAAAGAAACATGAAGTTCG  
ACGAAGAACTCACCTCCATCTATGAGGATTACATCAAAAAGATGAGAGAAACAGAGGAA  
TATAAACCCAGAACTGACTCCTGGGGAACGGTTATAAAAACCGAACTTCCAGAGAACTT  
TCTCTCAGAAAAAGAGATAAAGAAGGCTGCGAAGAAAAACGATGCTGCAGTTGTTGTAA  
TCAGTAGGATCTCCGGTGAGGGATACGACAGAAAGCCGGTGAAAGGTGACTTACCTCT  
CCGATGACGAGCTGGAGCTCATAAAAACAGTCTCAAGGGAATTCACGAACAGGGTAAG  
AAGGTTGTGGTTCCTTCTCAACATCGGAAGTCCCATTGAAGTTGCAAGCTGGAGAGATCT  
TGTGGATGGAATCCTTCTCGTCTGGCAAGCAGGACAGGAGATGGGAAGAATAGTGGCCG  
ATGTTCTTGTGGGAAGGGTAAACCCCTCCGAAAACCTTCCAACGACCTTCCCGAAGGAT  
TACTCGGACGTTCCATCCTGGACGTTCCAGGAGAGCCAAAGGACAATCCGCAAAGAGT  
GGTGTACGAGGAAGACATCTACGTGGGATACAGGTA CTACGACACCTTTGGTGTGGAAC  
CTGCCTACGAGTTCGGCTACGGCCTCTCTTACACAAAGTTTGAATACAAAGATTTAAAG  
ATCGCTATCGACGGAGATATACTCAGAGTGTCTGACACGATCACAACACCGGGGACAG  
AGCTGGAAAGGAAGTCTCACAGGTTTATGTCAAAGCTCCAAAAGGGAAAATAGACAAAC  
CCTTCCAGGAGCTGAAAGCGTTCACAAAACAAAACCTTTTGAACCCGGGTGAATCCGAA  
AAGATCTTTCTGGAATTCCTCTTAGAGATCTTGCAGTTCGATGGGAAAAGATGG  
TTGTGAGT CAGGAGAATACGAGGT CAGGGT CGGTGCATCTT CAGGGGATATAGGTTGA  
GAGATATTTTTCTGGTTGAGGGAGAGAAGAGATTCAAACCATGA

Figure 7A

B.

MMGKIDEIILSQLTIEEKVKLVVGVGLPGLFGNPHSRVAGAAGETHPVPRLGIPSFVLAD  
GPAGLRINPTRENDENTYYTTAFPVEIMLASTWNKDLEEVGKAMGEEVREYGVVDVLLA  
PAMNIHRNPLCGRNFYYSSEDPVLSGEMASAFVKGVSQSGVACIKHFVANNQETNRMV  
VDTIVSERALREIYLKGFIEIAVKKARPWTVMSAYNKLNGKYCSQNEWLLKKVLRREEWGF  
DGFVMSDWYAGDNPVEQLKAGNDMIMPGKAYQVNTERRDEIEEIMBALKEGRLSEEVLN  
ECVRNLIKVLVNAPSFKGYRYSNKPDLSEKAVAYEAGVEGVVLLNNGVLPFDES IHV  
AVFGTGQIETIKGGTGSGDTHPRYTISILEGIKERNMKFDEELTSIYEDYIKKMRETEE  
YKPRTDSWGTVIKPKLPENFLSEKEIKKAAKNDAAVVVISRISGEGYDRKPKVKGDFYL  
SDDELELIKTVSREFHEQGKVVVLLNIGSPIEVASWRDLVDGILLVWQAGQEMGRIVA  
DVLVGRVNPSTGKLPPTTFPKDYSDVPSWTFPGEPKDNQVRYEEDIYVGYRYDFTFGVE  
PAYEFGYGLSYTKFEYKDLKIAIDGDILRVSYTITNTGDRAGKEVSQVYVKAPKIDK  
PFQELKAFHKTLLNPGESEKIFLEIPLRDLASFDGKEWVVESGEYEVVRVGASSRDIRL  
RDI FLVEGEKRFKP

Figure 7B

A.

ATGAACGTGAAAAAGTTCCCTGAAGGATTCCCTCTGGGGTGTGCAACAGCTTCCTACCA  
GATCGAGGGTTCTCCCCTCGCAGACGGAGCTGGTATGTCTATCTGGCACACCTTCTCCC  
ATACTCCTGGAAAATGTAAAGAACGGTGACACGGGAGATGTGGCCTGCGACCACTACAAC  
AGATGGAAAGAGGACATTGAAATCATAGAGAACTCGGAGTAAAGGCTTACAGATTTTC  
AATCAGCTGGCCAAGAATACTTCCGGAAGGAACAGGAAGGGTGAATCAGAAAAGGACTGG  
ATTTTTACAACAGGATCATAGACACCCTGCTGGAAAAAGGTATCACACCCTTTGTGACC  
ATCTATCACTGGGATCTTCCCTTCGCTCTTCAGTTGAAAGGAGGATGGGCGAACAGAGA  
AATAGCGGATTGGTTCGCAGAATACTCAAGGGTTCTCTTTGAAAATTTCCGGCGACCGTG  
TGAAGAACTGGATCACCTTGAACGAACCGTGGGTTGTTGCCATAGTGGGGCATCTGTAC  
GGAGTCCACGCTCCTGGAATGAGAGATATTTACGTGGCTTTCCGAGCTGTTCACAATCT  
CTTGAGGGCACACGCCAAAGCGGTGAAAGTGTTCAGGGAAACTGTGAAAGATGGAAAGA  
TCGGAATAGTTTCAACAATGGATATTTCGAACCTGCGAGTGAAAAAGAGGAGGACATC  
AGAGCGGCGAGATTCATGCATCAGTTCAACAATACTCCTCTCTTTCTCAATCCGATCTA  
CAGAGGAGATTATCCGGAGCTCGTTCGGAATTTGCCAGAGAGTATCTACCGGAGAATT  
ACAAAGATGACATGTCCGAGATACAGGAAAAGATCGACTTTGTTGGATTGAACTATTAC  
TCCGGTCATTTGGTGAAGTTCGATCCAGATGCACCAGCTAAGGTCTCTTTTCGTTGAAAG  
GGATCTTCCAAAAACAGCCATGGGATGGGAGATCGTTCAGAAGGAATCTACTGGATCC  
TGAAGAAGGTGAAAGAAGAATACAACCCACCAGAGGTTTACATCACAGAGAATGGGGCT  
GCTTTTGACGACGTAGTTAGTGAAGATGGAAGAGTTCACGATCAAAACAGAATCGATTA  
TTTGAAGGCCCACATTGGTTCAGGCATGGAAGGCCATACAGGAGGGAGTGCCGCTTAAAG  
GTTACTTCGTCTGGTTCGCTCCTCGACAATTTCGAATGGGCAGAGGGATATTCCAAGAGA  
TTTGGTATTGTGTACGTGGACTACAGTACTCAAAAACGCATCATAAAAAGACAGTGGTTA  
CTGGTACTCGAACGTGGTCAAAAGCAACAGTCTGGAAGATTGA

B.

MNVKKFPEGFLWGVATASYQIEGSPADGAGMSIWHTFSHTPGNVKNGDTGDVACDHYN  
RWKEDIEIEKLGVKAYRFSISWPRILPEGTGRVNQKGLDFYNRIIDTLLEKGITPFVVT  
IYHWDLPFALQLKGGWANREIADWFAEYSRVLFENFGDRVKNWITLNEPWVVAIVGHLY  
GVHAPGMRDIYVAFRAVHNLRAHAKAVKVFRETVDKGI GIVFNNGYFEPASEKBEDI  
RAARFMHQFNYPFLNPIYRGDYPELVLEFAREYLPENYKDDMSEIQEKIDFVGLNYY  
SGHLVKFDPDAPAKVSFVERDLPKTAMGWEIVPEGIYWILKKVKEYNPPEVYITENGA  
AFDDVVSSEDGRVHDQNRIDYLKAHIGQAWKAIQEGVPLKGYFVWSLLDNFEWAEGYSKR  
FGIVYVDYSTQKRIIKDSGYWYSNVVKSNSLED

Figure 8A-B

High-temperature operating and thermo-stable cellulases

PROTEIN	Source		Physical Properties						Functional Properties				
	Locus	Codon Usage	CaZy	MW <sub>d</sub>	pI	Charge <sub>pH7</sub>	T <sup>0</sup> <sub>°C</sub>	pH range	Mode of Operation	Specific Activity			
										<sub>sw</sub> Avicel	<sub>av</sub> Cellulose	Paper	pNPG
Tcel1	<i>o-eglA</i>	corn optimized	GH12	34,005	4.80	-13.10	102	7 - 8	exocellulase	63.4	13.6	10.5	-
Tcel2	<i>petroB</i>	bacteria	GH12	31,816	4.77	-13.30	98	6 - 7	exocellulase	8.1	2.2	2.9	-
Tcel3	<i>ph1171</i>	archaea	GH5	51,930	6.47	-3.60	94	6	exocellulase	48.5	8.4	8.6	-
Tcel4	<i>o-E1</i>	rice optimized	GH5	59,980	7.05	0.30	95	5 - 6	endocellulase	6.8	2.2	4.1	-
Tcel5	<i>petroA</i>	bacteria	GH12	38,226	5.58	-6.60	96	5 - 6	endocellulase	34.1	6.8	8.0	-
Tcel6	<i>zp#4</i>	corn optimized	GH12	31,818	5.66	-5.00	85	5 - 6	endocellulase	20.6	5.1	5.1	-
Tcel7	<i>Tpet0898</i>	bacteria	GH3	81,243	5.38	-16.90	98	5	beta-glucosidase	0.8	0.8	-	69.4
Tcel8	<i>Tpet0952</i>	bacteria	GH1	51,509	5.84	-9.10	92	6 - 7	beta-glucosidase	1.7	1.5	-	60.9
Tcel9	<i>g12#3</i>	corn optimized	GH12	45,059	6.16	-2.20	85	5 - 6	endocellulase	5.2	1.4	7.5	-
Tcel10	<i>ph0746</i>	archaea	GH65	85,598	7.80	4.30	94	6	endocellulase	4.9	1.5	3.9	-

<sub>sw</sub>AVICEL Phosphoric acid swollen Avicel, <sub>av</sub>Cellulose, Avicel (SigmaCell): Cellulase specific activity, mM reducing sugar/mg protein/day at 85 °C, pH 6  
 Beta-glucosidase specific activity, nM pN/ug protein/min

Figure 9

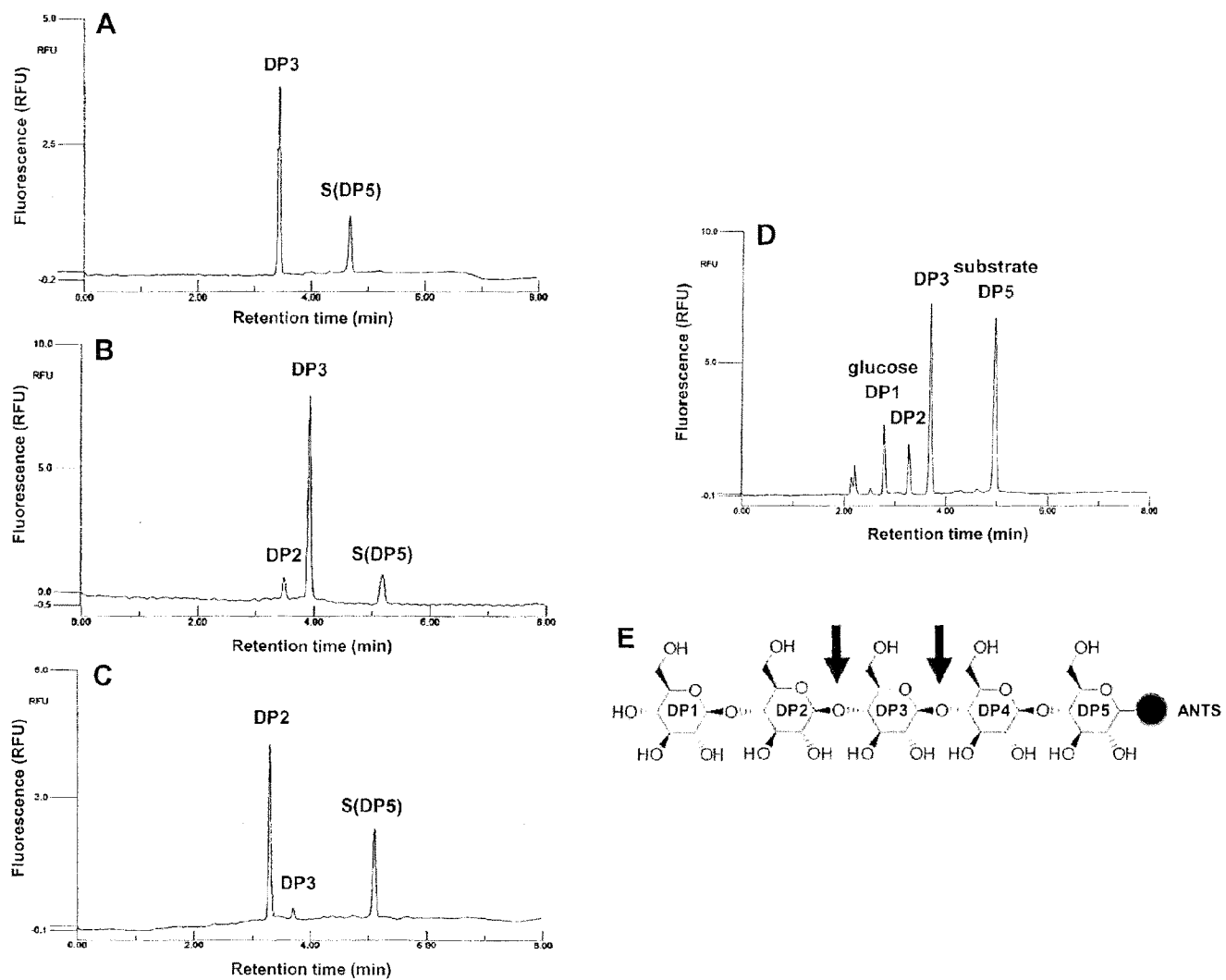


Figure 10A-E

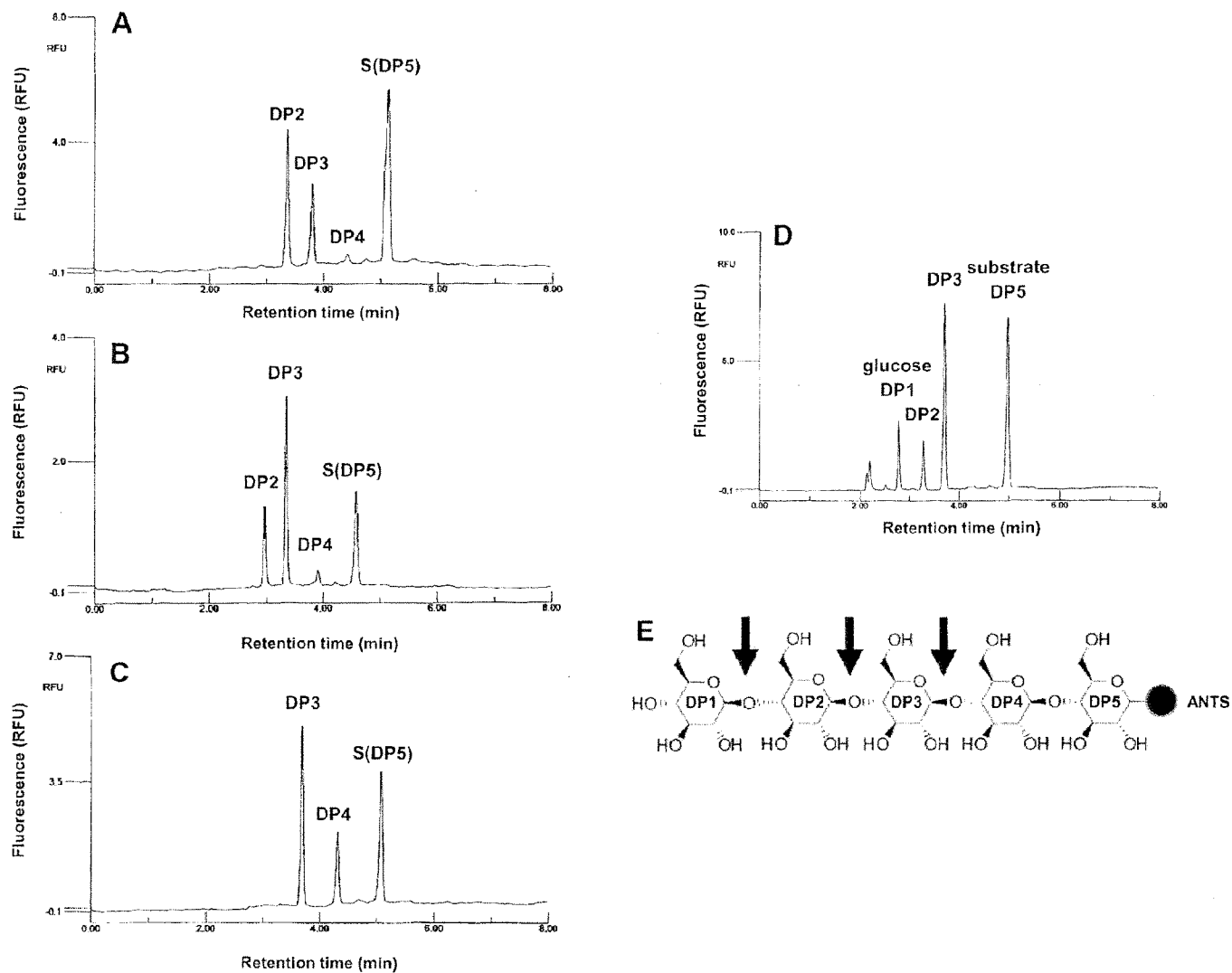


Figure 11A-E

## High-temperature catalytic operating cellulases

PROTEIN	U	T opt °C	% Specific activity at		
			60 °C	45 °C	20 °C
AVICEL <sub>sw</sub>					
Tcel1	63.4	102	56.0	40.0	20.0
Tcel2	8.1	98.0	42.2	25.3	14.1
Tcel3	48.5	94.0	44.7	21.1	10.5
Tcel4	6.8	95.0	26.0	11.6	4.3
Tcel5	34.1	96.0	53.6	34.1	14.6
Tcel6	20.6	85.0	71.4	46.4	14.3
pNPG					
Tcel7	69.4	98.0	4.4	1.1	1.0
Tcel8	60.9	92.0	25.0	12.5	2.5

<sub>sw</sub>AVICEL Phosphoric acid swollen Avicel, <sub>av</sub>Cellulose, Avicel (SigmaCell):  
 Cellulase specific activity,  $\mu\text{M}$  reducing sugar/mg protein/day at 85 °C, pH  
 6, Beta-glucosidase specific activity, nM pN/ug protein/min

Figure 12

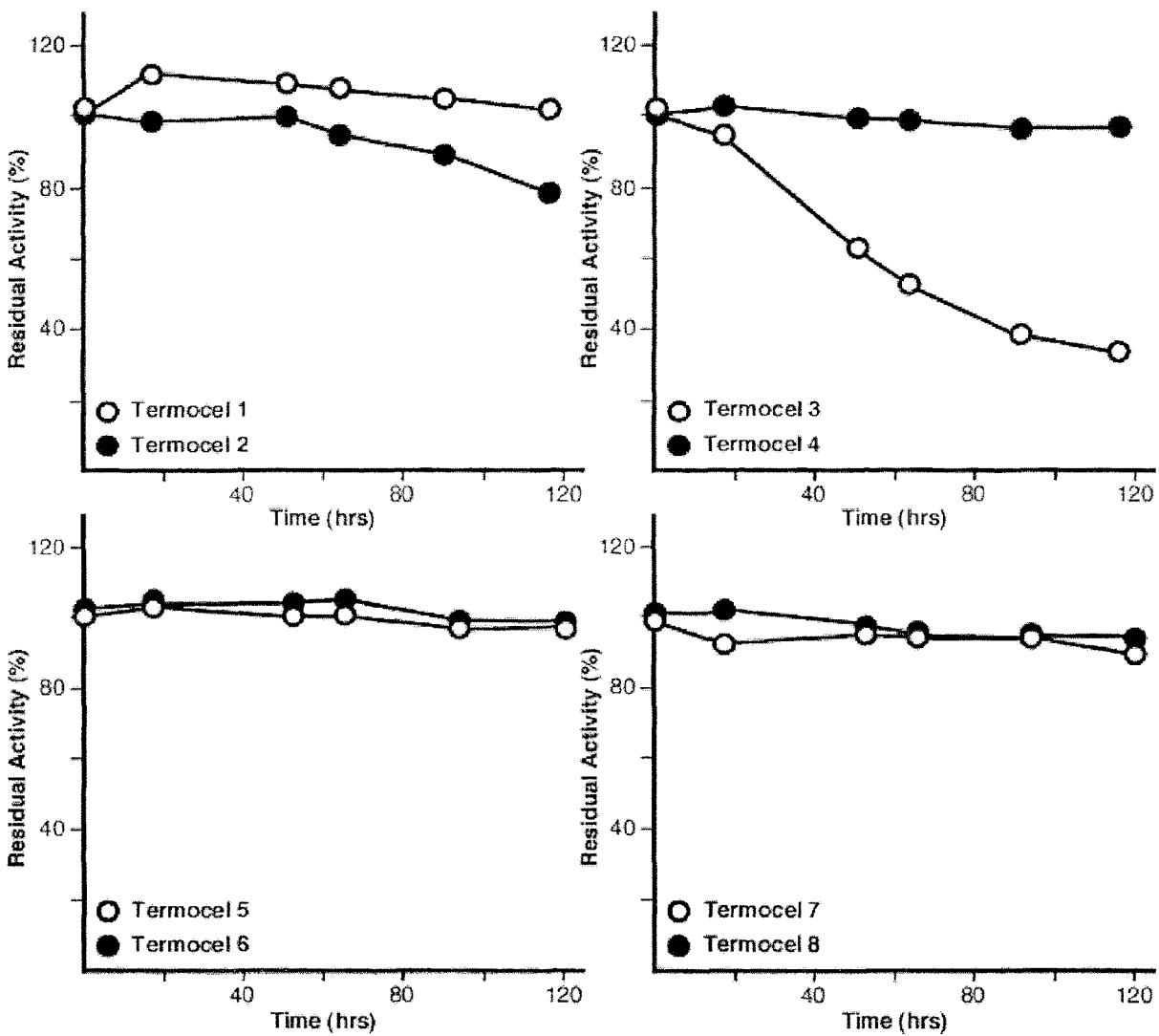


Figure 13A-D



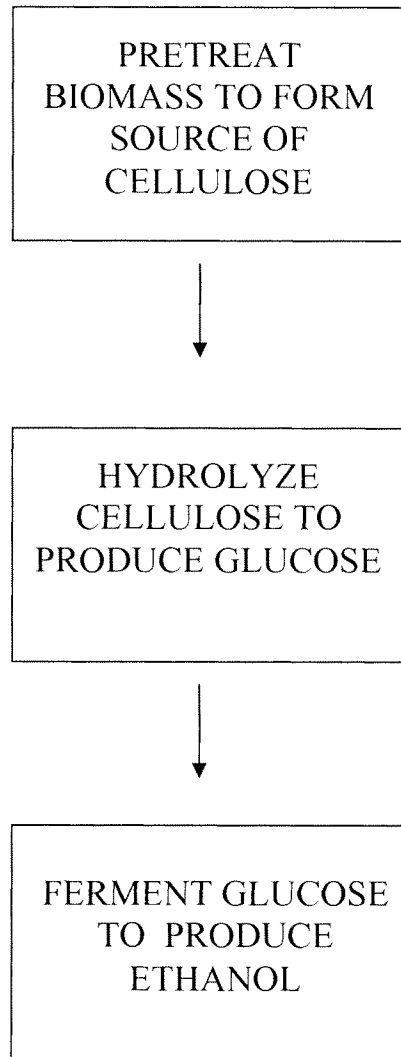


Figure 14

Biomass substrate specificity of Termocels

Substrate	Specific activity (reducing-end $\mu$ mole/mg protein/h)							
	Tcel1	Tcel2	Tcel3	Tcel4	Tcel5	Tcel6	Tcel7	Tcel8
avicel	1.62	-	-	-	5.35	-	0.75	1.73
swollen avicel	29.82	57.48	11.28	21.73	36.67	29.77	0.79	1.57
carboxymethyl cellulose	69.01	234.77	68.66	53.75	131.22	167.28	0.79	1.35
alpha-cellulose	6.12	21.42	1.16	12.52	7.34	5.06	0.71	0.74
barley beta-glucan	67.51	158.26	45.07	32.26	160.36	117.91	2.99	31.34
laminarin	4.22	21.55	3.94	17.45	6.47	10.65	21.08	28.68
lichenan	110.12	269.03	82.17	51.29	238.06	192.05	8.34	44.51
starch	0.45	18.77	3.34	13.94	4.26	6.23	0.48	1.53
birch wood xylan	31.26	18.73	4.68	19.18	nd	nd	0.30	1.23
beechwood xylan	27.55	18.98	3.69	24.31	27.30	136.76	0.89	1.12
oat-spelt xylan	19.07	22.23	3.44	17.51	34.29	70.27	0.70	1.32
Wheat arabinoxylan	23.52	21.85	3.49	15.83	36.24	181.46	0.70	1.18

Figure 15

A.

ATGGACTACTCTATCAACTGCTCTATCAACCCTATAACCCTCATGGTTCGCGCACTCTTC  
TCCCCTGAACCCATCTAACACACTCGAACTTACACTTATTCTCGAAAATGGCATCACCA  
CCACAGTAACTGTCACCGCGACACCACGCAACACTTACCCTATGATCTCCCTTGGCTAC  
ATTAATATTACCCCTAACCTCTGGAACCTTAACACAGCTTCGTTCATCAGGATACGCCTC  
TATGGTCTACGATGCATCACAGGGTGCTCTTTATATTCATGTTAATTTTACAAAGGTTT  
ACCTCAATCAGCAAGTTGGTGTTCGCCCTACTCTGAATTTCATCTATGGCTACAAACCC  
TGGGGCACGCTCACCTCCGAGGCAGGCGGGTTCAATTTTCTGTTAAGCTTACCGAACT  
CGGTTCTCTTCTTTCTGTTTCATCAATTACTCACTCATTTTCATATTCTCCACAAGTCGCTA  
TCTTCGATTGGGCATACGACCTTTGGCTCACAACATCCCCAAATCTCACCAACGGCCCT  
CAACCCGGCGACGTTCGAGGTCATGATCTGGCTCTATTATCACCTGCAACAACCTGCGGG  
TTTTCCCGTCGCTAACGTTACAGTGCCAATATGGGTCAATGGCTCCCTCGTTAACGAAA  
CATTTGAGGTTTGGATTGGTTCTCCACAGATCGAACC CGGCACCCACGCTATAGTCTCC  
TTCAGGCCAACGAATCCAATCCCTAGAGGCCTCGTCGGCGTAAATGTCACGAAGTTCCCT  
TCAACTTGCCGTTAACTATCTCGTGACACTCTACCCCTCATACTGGAACCTACACATATC  
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TACAATATTACACTCAATTGGGTCAATTTATAAAGCTTATCTTATCAAGGTGCCTCTGGA  
GTCACAGGGCACCGTTACCGTCACATATACTACAACCTGTTACATCCACCATGACTGTTA  
CCTCAATCCTTGCTACCACATCCACCGTCACCACTACATCTACACTTACATCTACCGTT  
ACCGCCACTTCAGTTTCTACTTCCACCGTCACGCAGACTCTCACTACCTCCATCGTCAA  
AACCGTCATCCCTGTCTACTATACTGCCACCATAATCGTCCTTCTTATAATCATCGCAG  
TCGTCAATTGCACTTGCGTTCGCCCGCGGCATCCGGGTTTCGTCTCTGT

B.

MDYSINCSINPITLMVAHSSPLNPSNTLELTLILENGITTTVTVTATPRNTYPMISLGY  
INITPNLWNLNTASSSGYASMVYDASQGALYIHVNF TKVYLNQQVGVAAYSEFIYGYKP  
WGLTSEAGGFNFPVKLTELGSLLSFINYSLISYSPQVAIFDWAYDLWLTSPNLTNGP  
QPGDVEVMIWLYYHLQQPAGFPVANVTVP I WVNGLVNETFEVWIGSPQIEPGTHAIVS  
FRPTNPIPRGLVGVNVTKFLQLAVNYLVTLYPSYWNNTY LESKYLNGIEFGSEWGNPST  
YNITLNWVIYKAYLIKVPLESQGTVTVTYTTTTVTSTMTVTSILATTSTVTTTSTLTSTV  
TATSVSTSTVTQTLTTSIVKTVIPVYYTATI I VLLII I AVVIALAFARRGIRVRLC

Figure 16A-B

A.

ATGAGATTTCAATTCGGATTCTCCAAAGAAGATGAACAGGTGCTGGGCACAATACTAAC  
ACTCGGAAATGGACAATTAGGAGTTAGGGGAGAATTTGAACTCGAGAGATCTCCTTATG  
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GGTCCCAGGACTATAGGGATGATAATAATTATAGATGGAGAACTAATAAATCCAAGCTC  
TCAAAAAGTCAAGGAATTCAGAGAGAGCTCGATATAGAAAAAGGCTTATTAAGAACTC  
ACTTAGAGATTGAAACAAAAAATGGAAATAAAATTTTATATAAAAAGTACAAGGATAGTC  
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AGATAATGATCAAGCATTATAGAGTGGACTCGATAAAAAGAGACTGAGGAGGGAGTATAC  
GCTAGGGTGAAAACTTTAGACAATAAGTACACGTTGGAAATTGCAAGTAGCTTGGTTCC  
ATCAGAATATACATCGAGGAGCACCTTTAGAACCGATAATGAAATTGGAGAAATTTACA  
TTGTTAAACTTAAACCAGGAAAAACGTACAAATTTACAAAGTACGTTACAGTATCTAAA  
GGAGCAGCTTTAGAGGAGTTAAAAGATGTTAAGAGATTAGGATTTGAAAAGCTATATGA  
AGAGCATATAAACAGCTGGAAGAGAATATGGGAGAAAGTGAAAGTGAAATCGAAGGAG  
ATAAAGACCTTGAAAATGCCCTAAACTTTAACATTTTTCACTTGATCCAATCCCTTCCA  
CCAACAGATAAAGTCTCGCTACCAGCAAGGGGAATACATGGGTTTGGGTATAGGGGACA  
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AGGCCAGGAGATTGCTCCTCTATAGATGCAACAACCTTAGATGCCGCTAAAGAAAATGCA  
AAGATGAATGGATATCAAGGGTCCAATTTCCCTGGGAGTCGGCAGATGATGGACGCGA  
GGCTACCCCTCTGAGATACCATTGGATATGTTGGGAAGGAAAATCGTTAGAATTTACA  
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CTTGAAGGATCGAGAAAAGTCAAAATATCTAGCTTTGGAAAGGAAGTAGATCTATATCC  
TGGAAAAGAGGTTGTAATAGTAGCTAATTA

Figure 17A

B.

MRFQFGFSKEDEQVLGTILTLGNGQLGVRGEFELERSPYGTIVSGVYDYTPYFYRELVN  
GPRTIGMIIIDGELINPSSQKVKEFQRELDIEKGLLRTHLEIETKNGNKILYKSTRIV  
HMKRKNLILLDFELKASKGGIADVVPNPIEFNTANPGFIDEIMIKHYRVDSIKETEEGVY  
ARVKTLDNKYTLLEIASSLVPSEYTSRSTFRTDNEIGEIIYIVKLPKGKTYKFTKYVTVSK  
GAALEELKDVKRLGFELKLYEEHINSWKRIWEKVKVEIEGDKDLENALNFNIFHLIQSLP  
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KMNGYQGVQFPWESADDGREATPSEIPLDMLGRKIVRIYTGEEHHITADIAYIVDFYY  
QVSGDLEFMNRCGLEIIFETARFASRVFEFEEGKGYVIKKVIGPDEYHEHVNNNFNTL  
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DGYFELMDFEVDPFNIGKTLPEEIRNNIGKTKLVKQADVIMAQYLLKDYFSPEEIKSN  
FNYYIRRTTHASSLSMPPYAIATWIGEVKIAEYEFKRCANIDLKNVYGNTAEGFHLAT  
AGGTWQVLVRGFCGLNVKGNKIELNPNLPEKWKYVKFRIFFKGSWIEFKISRKKVRARM  
LEGSRKVKISSFGKEVDLYPGKEVVIVAN

Figure 17B

```
>termocel1_nt (Tcel1, o-eglA) 903 bp
ATGATCTATTTTGTGAGAAATACCCACCTCAGAGACAAATCCACAAGCAATACCTC
CTCAACCCCCCTCAAACGACACTTAGCACAAACAAGGTTCTCAAATTCGGTATCCTG
ACGACGGCGAATGGCCTGGCGCTCCCATAGACAAAGACGGCGACGGAAATCCTGAGTTC
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GCGAATTATGCCACCGACGGCCCGATCCCTCTCCCCAGTAAAGTTTCCAATCTCACAGA
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GGAAAGCTAATATCGGATCGGAATACGTTGCCCTTAGAATCAAGACACCAATTAAGAA
GGAACCGTGACAATCCCCACGGTGCAATTCATTAGCGTAGCTGCTAACATTTCTTCCCT
CCCAATTAACACAGAACTTTACCTGGAAGACGTTGAGATAGGCACAGAGTTTGGAAACAC
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GATCGTCCCCTGATCTCC (SEQ ID NO:1)
>termocel2_nt (Tcel2, petroB) 825 bp
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AGCTCAACTTCTGGAAATAAAGTCGTATGAGGGAGAAACGTGGCTCAAATTCGATGGA
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(SEQ ID NO:3)
>termocel3_nt (Tcel3, ph1171) 1377 bp
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ACTACGAAGTGAGAGGAGATACGATATACATGATTAATGTCACCAGTGGAGAGGAAACT
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CGGACTTTTGGAAAGAGAAACTGGGAAGACATGCTTCTTCAGATCAAAGCTTAGGCTTCA
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GATGCACTCACATAGAACCCCTCTGGTACACGGAAAGACTTCTCAGAGGAAGACTTTATT
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TCTAAAGAAATGAGCCTCATAGTGTACCTCACCCCAAGCTGCTTATACAGATGGTACCG
```

Figure 18A

GGGCTACATGGGGTATGGGAAACCCCTGCAACCGATTGGAACTTGGCGGCTGAGAGGATA  
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TACTAATCCGAAGACTGACAGTAGTTACAAATGGGGCTACAACGCTTGGTGGGGAGGAA  
ATCTAATGGCCGTAAAGGATTATCCAGTTAACTTACCTAGGAATAAGCTAGTATACAGC  
CCTCACGTATATGGGCCAGATGTCTATAATCAACCGTACTTGGTCCCGCTAAGGGTTT  
TCCGGATAATCTTCCAGATATCTGGTATCACCCTTGGATACGTAAAATTAGAAGTAG  
GATATTAGTGTAAATAGGAGAGTTTGGAGGAAAATATGGCCATGGAGGCGATCCAAGC  
GATGTTATATGGCAAATAAGCTAGTTGATTGGATGATAGAGAATAAAATTTTGTGATTT  
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CGACAACAATATGGGAAGATAAGTATAATAACCTGAAGAGATTGATGGATAGTTGTTCC  
AAAAGTTCTTCAAGTACTCAATCCGTTATTTCGGAGTACCACCCCTACAAAGTCAAATAC  
AAGTAAGAAGATTTGTGGACCAGCAATTCTTATCATCCTAGCAGTATCTCTCTTCTCT  
TAAGAAGGGCTCCCAGGTAG (SEQ ID NO: 5)

>termocel4\_nt (Tcel4, o-E1) 1542 bp

ATGGAAATCAAGCTCTTCTGCGTGTATTCGTGTTTCATCATCCCTCTTCTCCCTTTTCGT  
GATTCGACTCTCGTATCCAGATGTAACTATACTGCCGAGAATGGTATTATCTTCGTGC  
AGAACGTCCTACGGGTGAGAAGAAGCCACTTTATCTTCACGGAGTGTATGGTTTGGGA  
TTCGAGCTGAAGGACCACGTCGTCTATGGCTTGGATAAACGGAACTGGAAAGATATACT  
CAAGGATGTTAAGCGCTTGGGTTTTAATGCTATCAGGCTTCCCTTCTGCTCTGAAAGCA  
TCCGCCCTGATACGCGCCCTTCGCCTGAGCGGATAAACTACGAGTTGAACCCCGACTTG  
AAGAATCTGACTTCCCTCGAAATAATGGAGAAGATTATGAATACGCCAACTCAATCGG  
GCTCTACATACTCTTGGATTATCACCGCATCGGTTGTGAGGAGATCGAACCTCTTTGGT  
ATACCGAGAATTACTCAGAGGAGCAGTATATAAAGGATTGGATCTTCTTCGCAAAGCGG  
TTCGGGAAGTACCCTAACGTGATAGGAGCTGATATCAAGAACGAGCCGCATGGTGAAGC  
CGGGTGGGGTACGGGAGATGAGCGGGATTTCCGCCCTCTTTCGCGAGAAGGTCCGGCGCG  
AGATACTCAAGGTGGCCCCACACTGGTTGATATTCGTGAGGGAACGCAATATACCCAT  
GTCCCGAATATTGATGAGATCATCGAGAAGAAGGGCTGGTGGACATTTTGGGGAGAGAA  
TCTTATGGGAGTTAAGGACTATCCAGTCAGGCTTCCGCGCGGCAAGGTGCTGACTCAC  
CGCATGCTATGGACCATCTGTCTACAATGATGGACTACTTCAAGTCGCCAGACTTTCCG  
AACAAATATGCCGATAATCTGGGAAACACACTTCGGATACTTGACCGACCTGAATTATAC  
CTTGGTTCATAGGCGAGTGGGGTGGCAACTATGAGGGCCTTGACAAGGTGTGGCAAGACG  
CTTTCGTGAAGTGGCTGATTAAGAAGAAGATCTATAAATCTTCTACTGGTGCCTGAAC  
CCGGAGTCCGGTGACACCCGTGGCATCTTCTCGACGACTGGAAAACCGTTAACTGGGA  
AAAGATGAGGGTTATTTACAGGCTCATCAAGGCGGCGAAACCCCGAGTTTGAAGAACCC  
TTTACATCATTTTGAAAAATAACGCGACGACATCTATCCTGGGCGTGGGTGAGAGGATC  
CGGATTTACTGGTACACAAATGGCAAAGTTATTGACTCTAACTTCGCGCATTCCAGCGA  
AGGCGAAATGAACATTACAGTGACGAAGTCCATGACTCTGTACATCATCGTGAAGAAGG  
GCAATCAGACACTGAGGAAGGAACTCAAACGTGACGTTATCGGCGGCAATTACGGCTCC  
AATATCTCCACTACCCAGCTGGTTACTCCCAAGAAAGGCGGCGAAAGGATTAGCACCAG  
CCTGAAGCTGGCAATTAGCCTGCTCTTCATCTCTCTTCTGTTTGGTATCTCCTCCGGG  
AGAAGCAT (SEQ ID NO: 7)

>texmocel5\_nt (Tcel5, petroA) 987 bp

ATGGAAACGCTCCTCCCTGTAGTCGTGGTCCACGATATTGAGCCAGTTTCAATGCGTCT  
TCAGAGGTACAAGAACAATAATTCGATAAAAAGAGAAAAGCAGGGATTAATACCCCTGT

Figure 18B

TTTTTAAATTTTGGGTGTATTTAGTTCATTTGCGAATTTTCAGATTTTGAAATGTAAAC  
ATTTTCAATAAAGATGTTTTCTGGAGGTGATAATGGTGGTACTGATGACAAAACCGGG  
AACATCGSATTTTGTATGGAATGGCATTCCCUUTTCCATGGAGC TGAATCTGTGGAACA  
TAAAGGAATACTCCGGTCTGTAGCTATGAAATTCGACGGTGAAAAGGTAACTTTCGAC  
GCGGACATTCAGAACTTTTCTCCAAAAGAACCAGAAAAGGTACGTTCTCGGTTATCCCGA  
GTTCTATTACGGTTATAAACCCTGGGAAAAGCACACGGCAGAAAGGTTTCGAAACTTCCAG  
TACCTGTTTCCTCTATGAAATCATTTTCCGTGCGAAGTTTCTTTCGATATTCACCACGAA  
CCGTCTCTGCCTTTGAACITTTGCCATGGAAACATGGCTCACAAGAGAAAAGTACCAGAC  
GGAAGCGTCGATCGGCGATGTTGAAATCATGGTCTGGTTCATTTCAACAATCTCACAC  
CAGGGGGCAAAAAGATAGAGGAGTTTACGATTCGGTTCGTGCTGAACGGAGAGAGTGTG  
GAAGGCACCTGGGAACTGTGGCACGGGAGTGGGGATGGGACTACCTCGCTTTCGGCTT  
GAAGGATCCCGTGAAGAAGGGGAAGGTTGAAATTCGACGTTGAGCCATTTTCTTGTATGCC  
CCGGAAAGCTCTTTTGAATTCCTACTCGTGTGAAAGATTTGAAAATCTTTACTTCCACC  
GTCTGGGAAATTTGGAACCGAGTTTGGAAAGCCCGGAAACAAAAGAGCGCCCAATTCGGGTG  
GAAGTTTGAAAACITTCCTCTATTGATCTGGAGGTGAGAGAAATGA (SEQ ID NO: 9)  
>termocel6\_nt (Tcel6) artificial gene with rice codon  
optimization based on *Caldivirga maquilingsis* GH12 gene  
852 bp  
ATGTTGAAACTTATTCACCTTGTAAATGGCAATTATAAGTTGATTCAAATGGGAGCCACT  
CGCCGGCGTGCACGGAGCAGATATCGAGTGCATACATGTTACCCCAAACGTATGGAACA  
TAGATAAATCATCAGTTGGCACTGTACAGATCGAATATGAGCCCCAAGTTGGCTGTCTT  
CGTTTTTCAATTGATTTCCCGAGGATAAGTATAAGACATAATGTAGCCGTAGCCGCATA  
TTCAGAAAGTTATTTACGGACACAAGCCGTGGGGCCCCACCCTTGCATGGACCCTCAGT  
TCAAGTTCCTATCAAAGTCAATGAGTCAAAGGACTGTACTCGTATGTAATTATAAC  
GTTAAATCTAGGTCACCAGATGACTCAATCTTTAAATTTGCTTACGATCTCTGGCTTAC  
AACGTCCCCAAACCTTACAAACGGACCCCGAGCCAGGAGACCTAGAAGTTATGATCTGGT  
TGTACTACCACGGACAGCGCCCTGCAGGCAGACTCATCGGGGAACCTCCGCATGCCGATT  
ACATTTGGGCGATAGTGAGGCGGCACCTGACTTTGAAGTATGGGTGGCTGACACAGGAAT  
AGGAATCGGTGAATGGGCGGTAGTGACCTTCAGAATCAAGGACCCCAATAAAGGGCGGTT  
TGATAGGAGTTAACCTCATAAACTACATCGAAAGTGCTTTTAAAACGCTCGAAGAACTC  
AACCCGGTCAAGTGGCGGTACGGCGACCTGCTCAACAAATATCTTAATGGAATTGAAT  
CGGCAGTGAGTTTGGTAAATGTCCTCAGGAATGATAAAACTTAATTGGGAACTCTGGC  
GCCTGAGCCTTGTGAAAGACTCTTCT (SEQ ID NO: 11)  
>termocel9\_nt (Tcel9) artificial gene with corn codon  
optimization based on *Caldivirga maquilingsis* GH12 1230  
bp  
ATGGACTACTCTATCAACTGCCTATCAACCCATAAACCCCTCATGGTCCGGCACTCTTC  
TCCCCTGAACCCATCTAACACACTCGAACTTACACTTATTTCTCGAAAATGGCATCACCA  
CCACAGTAACITGTCACCGCGACACCACCCAACTTACCCTATGATCTCCCTTGGCTAC  
ATTAATATTTACCCCTAACCTCTGGAACCTTAACACAGCTTCGTCTATCAGGATACGCCTC  
TATGGTCTACGATGCATCACAGGGTGTCTTTATATTCATGTTAATTTACAAAAGGTTT  
ACCTCAATCAGCAAGTTGGTGTTCGCCCTACTCTGAATTCATCTATGGCTACAAAACCC  
TGGGGCACGCTCACCTCCGAGGCAGGCGGCTTCAATTTTCCCTGTTAAGCTTACCGAACT  
CGGTTCTCTTCTTTCGTTTCAATTAATTAATTAATTAATTAATTAATTAATTAATTAATTA  
TCTTCGATTTGGGCATACGACCTTTGGCTCACAACATCCCCAAATCTCACCAACGGCCCT  
CAACCCGGCGACGTCGAGGTGATGATCTGGCTCTATTAATCACCTGCAACAACTTGGGG  
TTTTCCCGTCGCTAACGTTACAGTGCCAATATGGGTCAATGGCTCCCTCGTTAACGAAA  
CATTTGAGGTTTGGATTGGTCTCCACAGATCGAAACCCGGCACCCACCGCTATAGTCTCC

Figure 18C



TTCCAGGCCAACGAATCCAATCCCCTAGAGGCCCTCGTCGGCGTAAA'TGTCACGAAGTTCCCT  
TCAACTTGCCG'FTAAC'FATCTCGTGACACTCTACCCCTCATACTGGAACTACACATATC  
TGGAGAGCAAGTACTTGAATGGCATCGAATTCGGATCAGAATGGGGCAATCCGTCTACA  
TACAA'TATTACACTCAAT'TGGGTCAT'TTATAAAGCTTATCTTATCAAGGTGCCTCTGGA  
GTCACAGGGCACCGTTACCGT'CACATATACTACAAC'TGTTACATCCACCATGACTGTTA  
CCTCAATCCCTTGTACCACATCCACCGT'CAACACTACATCTACACTTACATCT'ACCGTT  
ACCGGCACCT'ACAG'TTCT'ACTTCCACCGT'CACGCAGACTCTCACTACCTCCATCGTCAA  
AACCGTCATCCCTGTCTACTATACTGCCACCATAATCGTCC'TTCTTATAATCATCGCAG  
TCGTCAAT'PGACTTGGCTTCGCCCGCCGGCATCCGGGTTCCGTCT'CGT(SEQ ID NO: 29)  
>termocell10\_nt (Tcell10) Based on *Pyrococcus horikoshii* OT3  
RKU GH65 gene 2214 bp  
ATGAGATTTCAATTCGGATTCTCCAAAGAAGATGAACAGGTGCTGGGCACAATACTAAC  
ACTCGGAAA'TGGACAA'TTAGGAGTTAGGGGAGAATTTGAACTCGAGAGATCTCCTTATG  
GAACGATCGTTAGCGGGGTCTATGAT'TACACTCCCTACTTCTACAGGGAATTCGTAAAT  
GGTCCCAGGACTATAGGGATGATAATAATTAATGATGGAGAACTAATAAATCCAAGCTC  
TCAAAAAGTCAAGGAATTCAGAGAGAGCTCGATATAGAAAAGGCTTATTAAGAACTC  
ACTTAGAGATTGAAACAAAAATGGAAATAAAATTTTATATAAAAAGTACAAGGATAGTC  
CACATGAAAAGAAAAACCTAATCCTTCTAGATTTTGGAGCTAAAAGCTAGCAAGGGAGG  
AATCGCAGTTGTAGTTAATCCCATAGAAATCAATACTGCAAAATCCAGGCTTATAGACG  
AGATAATGATCAAGCATTATAGAGTGGACTCGATAAAAGAGACTGAGGAGGGAGTATAC  
CCTAGGCTGAAAAC'TTAGACAATAAGTACACCTTGGAAATTCGAAGTACCTTGGTTCC  
ATCAGAATATACATCGAGGAGCACCTTTAGAACCAGTAATGAAATTCGAGAAA'TTACA  
TTGTTAAACT'FAAACAGGAAAACGTACAAATTTACAAAGTACGTTACAGTATCTFAAA  
GGAGCACCTTTAGAGGAGTTAAAAGATGTTAAGAGATTAGGATTTGAAAAGCTATATGA  
AGAGCATATAAACAGCTGGAAGAGAATATGGGAGAAAGTGAAGTGGAAATCGAAGGAG  
ATAAAAGACCTTGAAAATGCCCTAAACTTTAACATTTTCACTTGATCCCATCCCTTCCA  
CCAACAGATAAAGTCTCGCTACCGCAAGGGGAATACATGGGTTTGGGTATAGGGGACA  
TATA'TT'CGGATACAGAGATAATATGCATTACCTTTCTTCAATATCACGATGCCAAAAG  
AGGCCAGCAGATTGCTCCTCTATAGATGCAACAACCTTAGATGCCCGCTAAAGAAAA'TGCA  
AAGATGAATGGATATCAAGGGTCCAA'TTCCCTGGGAGTCGCCAGATGATGGACCGCA  
GGCTACCCCTCT'GAGATACCATTGGATATGTTGGGAAGGAAAATCGTTAGAA'TTACA  
CCGGAGAGGAGGAACATCACATAACTGCCGATATAGCATATATAGT'PGATTTTAT'ITAC  
CAAGTCTCTGGAGATCTCGAATTTATGAACAGCTGTGGCC'TGAGATAATCTTTGAGAC  
GGCCCGATTTTGGGCTAGTAGGGTTGAGTTCGAGGAAGGAAAACGGTACGTCATTA  
AAGTAATAGGACCTGATGAATACCATGAGCACGT'FAACAACAAC'TCTTTACAACTTA  
ATGGCCAAGCATAATCTCGAACTTGCAATAAGATACTTTAGAGAGTCAAAGAATAGGGA  
ACCGTGGAAAAGATTTGTCGAAAAATTAACATAAGAGAGGAGGAGGTTGAAAAATGGG  
AAGAGATAGCTAAAAACATGTACATTTCCAGGAAGA'TAGACGGAGTTTTTGAAGAGTTT  
GATGGTTACTTTGAATTGATGGATTTTGAAGTTGATCCCTTCAATATGGAGAAAAAC  
ACTCCCCGAGGAAATCAGGAATAACATAGGGAAAACGAAACTCGTTAAGCAGGCCCGATG  
TCATCATGGCCCAATATCTCCTTAAGGACTACTTCTCTCCAGAGGAAA'TAAAGAGTAAC  
TTAAC'ATATATAAAGGAGAACTACCCATGCTTCACTACTCTCCATGCCCCCATACGC  
GATCAATGCAACCTGGATAGGGGAGGTAAAAGATAGCATATGAGTACTTCAAGAGATGTG  
CAAA'TATAGATCTCAAAAACGTGTACGGAAACACTCCAGAGGGATTTCACTTAGCAACG  
CGGGAGGAACCTGGCAAGTACTCGT'AGAGGATTTTGTGGCCTCAATGTAAGGAAA

Figure 18D

CAAANTAGAGCTTAATCCTAATCTTCCTGAAAAATGGAAGTACGTAAAGTTCAGGATAT  
TCTTCAAAGGTTTCATGGATAGAATTTAAAATTTCTAGGAAGAAAGTTAGGGCTAGAATG  
CTTGAAGGATCGAGAAAAGTCAAAATATCTAGCTTTGGAAAGGAAGTGGATCTATATCC  
TGGAAAAGAGGTTGTAATAGTAGCTAATTAA (SEQ ID NO: 31)  
>termocel7\_nt (Tce17) Based on *Thermotoga petrophila* RKU  
GH3 gene 2169 bp  
ATGATGGGAAAGATCGATGAAATCCTTTTACAGCTGACTATTGAAGAAAAAGTGAAACT  
TGTAGTGGGGGTTGGTCTTCCAGGACTTTTGGAAATCCACATTCAGAGTGGCAGGTG  
CAGCTGGAGAAAACGCATCCTGTTCCGAGGCTTGGAAATCCTTCTTTTCGTTCTGGCCGAC  
GGTCCCGCGGGCCTCAGAATAAAATCCACAAAGAGAGAACGACGAAAACACCTATTACAC  
AACAGCGTTTTCTGTGAAATCATGCTCGCTTCCACCTGGAACAAAAGATCTTCTGGAAG  
AAGTAGGAAAAGCTATGGGAGAAGAAGTACAGGAATACGGTGTGATGTGCTTCTTGCA  
CCTGCGATGAAACATTCACAGGAACCCCTCTTTGTTGGAAGGAATTTGAGTATTATTGAGA  
AGATCCTGTCTTTCCGGTGAATGGCTTCAGCCTTTGTCAAGGGAGTCAATCTCAAG  
GGTGGGAGCCTGCATAAAAACACTTTGTGCGCAACAACCAGGAAACGAACAGGATGGTA  
GTGGACACGATCGTGTCCGAGCGAGCCCTCAGAGAAATATATCTGAAAGGTTTTGAAAT  
TGCCGTCAAGAAAGCAAGACCCTGGACCGTGTGATGAGCGTTACAACAAACTGAATGGAA  
AATACTGTTTACAGAACGAATGGCTTTTGAAGAAGGTTCTCAGGGAAGAATGGGGATTT  
GACGGTTTCGTGATGAGCGACTGGTACGCGGGAGACAACCCTGTAGAACAGCTCAAGGC  
CGGAAACGATATGATCATGCCITGGAAAAGCGTATCAGGTGAACACGGAAAGAAGAGATG  
AAATAGAAGAAATCATGGAGGCCCTTGAAGGAGGGAAGACTCAGTGAGGAAGTCCGAAAC  
GAATGTGTGAGAAACATCCTCAAAGTTCTTGTGAACGCGCCTTCTTTAAAGGGTACAG  
GTACTCGAACAAACCCGACCTCGAATCTCACGCGAAAGTTGCCTACGAAGCAGGTGTGG  
AGGGTGTGTCTTCTTGAAGCAACCGGTGTTCTTCCATTCGATGAAAGTATCCATGTC  
GCCGTCTTTGGCACCCGTCAAATCGAAACAATAAAGGGAGGAACGGGAAGTGGAGACAC  
CCATCCGAGATACACGATCTCTATCCTTGAAGGCATAAAAGAAAGAAACATGAAGTTCCG  
ACGAAGAACTCACCTCCATCTATGAGGATACATCAAAAAGATGAGAGAAAACAGAGGAA  
TATAAACCAGAACTGACTCCTGGGGAACCGTTATAAAAACCGAACTTCCAGAGAACTT  
TCTCTCAGAAAAAGAGATAAAGAAGGCTGCGAAGAAAAACGATGCTGCAGTTGTTGTAA  
TCAGTAGGATCTCCGGTGAAGGATACGACAGAAAGCCGGTGAAGGTGACTTCTACCTC  
TCCGATGACGAGCTGGAGCTCATAAAAAACAGTCTCAAGGGAAATTCACGAACAGGGTAA  
GAAGGTTGTGGTTCTTCTCAACATCGGAAGTCCCATTGAAGTTGCAAGCTGGAGAGATC  
TTGTGGATGGAATCCTTCTCGTCTGGCAAGCAGGACAGGAGATGGGAAGAATAGTGGCC  
GATGTTCTTGTGGGAAGGGTAAACCCCTCCGGAAAACCTCCAACGACCTTCCCGAAGGA  
TTACTCGGACGTTCCATCCTCGACGTTCCGAGGAGAGCCAAAGGACAATCCGCAAAGAG  
TGGTGTACGAGGAAGACATCTACGTGGGATACAGGTAATACGACACCTTTGGTGTGGAA  
CCTGCCTACGAGTTCCGGCTACGGCCCTCTTACACAAAGTTTGAATACAAAGATTTAAA  
GATCGCTATCGACGGAGATATACTCAGAGTGTGCTACACGATCAAAAACCCGGGGACA  
GAGCTGGAAGGAAGTCTCACAGGTTTATGTCAAAGCTCAAAGGGGAAAATAGACAAA  
CCCTTCCAGGAGCTGAAAGCGTTCCACAAAACAAAACCTTTTGAACCCGGGTGAATCCGA  
AAAGATCTTTCTGGAAATTCCTCTAGAGATCTTGGGAGTTTCGATGGGAAAGAATGGG  
TTGTGAGTCAAGGAGAAATACGAGGTCAGGTCGGTGCATCTTCCAGGGATATAAGGTTG  
AGAGATATTTTTCTGGTTGAGGGAGAGAAGAGATTCAAACCATGA (SEQ ID NO: 13)  
>termocel8\_nt (Tce18) Based on *Thermotoga petrophila* RKU  
GH1 gene 1341 bp

Figure 18E

ATGAACGTGAAAAAGTTCCCTGAAGGATTCCTCTGGGGTGTGCAACAGCTTCCTACCA  
GATCGAGGGTTCTCCCTCGCAGACGGAGCTGGTATGTCTATCTGGCACACCTTCTCCC  
ATACTCCTGGAAATGTAAAGAACGGTGACACGGGAGATGTGGCCTGCGACCACTACAAC  
AGATGGAAAGAGGACATTGAAATCATAGAGAAACTCGGAGTAAAGGCTTACAGATTTTC  
AATCAGCTGGCCAAGAATACTTCCGGAAGGAACAGGAAGGGTGAATCAGAAAGGACTGG  
ATTTTTACAACAGGATCATAGACACCCCTGCTGGAAAAAGGTATCACACCCCTTTGTGACC  
ATCTATCACTGGGATCTTCCCTTCGCTCTTCAGTTGAAAGGAGGATGGGCGAACAGAGA  
AATAGCGGATTGGTTTCGAGAAATACTCAAGGGTCTCTTTGAAAATTTCCGGCGACCGTG  
TGAAGAACTGGATCACCTTGAACGAACCGTGGGTGTTGCCATAGTGGGGCATCTGTAC  
GGAGTCCACGCTCCTGGAATGAGAGATATTTACGTGGCTTTCGAGCTGTTCACAATCT  
CTTGAGGGCACACGCCAAAAGCGGTGAAAGTGTTCAGGGAAACTGTGAAAGATGGAAAGA  
TCGGAAATAGTTTTCAACATGGATATTTGAAACCTGCGAGTGAAAAAGAGGAGGACATC  
AGAGCGGCGAGATTCATGCATCAGTTCAACAACATATCCTCTCTTTCTCAATCCGATCTA  
CAGAGGAGATTATCCGGAGCTCGTTCGGAAATTTGCCAGAGAGTATCTACCGGAGAATT  
ACAAAGATGACATGTCCGAGATACAGGAAAAGATCGACTTTGTTGGATTGAACTATTAC  
TCCGGTCATTTGGTGAAGTTCGATCCAGATGCACCAGCTAAGGTCTCTTTCGTTGAAAG  
GGATCTTCCAAAAACAGCCAATGGGATGGGAGATCGTTCAGAAAGGAATCTACTGGATCC  
TGAAGAAGGTGAAAGAAGAAATACAACCCACCAGAGTTTTACATCACAGAGAATGGGGCT  
GCTTTTGACGACGTAGTTAGTGAAGATGGAAGAGTTCACGATCAAAACAGAATCGATTA  
TTTGAAGGCCCCACATTTGGTCAGGCATGGAAGGCCATACAGGAGGGACTGCCGCTTAAAG  
GTTACTTCGTCTGGTCGCTCCTCGACAATTTGAAATGGGCAGAGGGATATTTCAAAGAGA  
TTTGGTATTGTGTACGTGGACTACAGTACTCAAAAACGCATCATAAAAGACAGTGGTTA  
CTGGTACTCGAACGTGGTCAAAGCAACAGTCTGGAAGATTGA (SEQ ID NO: 15)  
>pBAD MYC-HIS TAG 66 bp  
GAACAAAACTCATCTCAGAAGAGGATCTGAATAGCGCCGTCGACCATCATCATCA  
TCATCAT (SEQ ID NO: 17)

>termocell1\_aa (Tcell, o-eglA) endoglucanase 301 aa (WITHOUT N-TERMINAL SIGNAL PEPTIDE, CODONS OPTIMIZED FOR CORN), GH12, MW 34,005, pI 4.80, charge -13.10

MIYFVEKYHTSDEKSTSNSTSTPPQTTLSTTKVLKIRYPDDGEWEGAPIDKDGDCGNPEF  
YIEINLWNILNATGFAEMTYNLTSGVLHYVQQLDNIVLRDRSNWVHGYPEIFYGNKFPWN  
ANYATDGPPIPLPSKVSNLTDYFLTISYKLEPKNGLPINFAIESWLTREAWRTTGINSDE  
QEVMIWIYYDGLQPAQSKVKEIVVPIIVNGTTPVNATFEVWKANIGWEYVAFRIKTPIKE  
GTVTIPYGAFISVAANISLFPNYTELYLEDVEIGTEFGTPTSTSAHLEWITNITLPL  
DRPLIS (SEQ ID NO: 2)

>termocell2\_aa (Tcel2, petroB), endoglucanase, 274 aa, GH12, MW 38,226, pI 5.58, charge -6.60

MRWVVLMLVAPALLFSSEVVLTSSVGAADISFNGFPVTMELNFWNIKSYEGETWLKFDG  
EKVEFYADLYNIVLQNPDSWVHGYPEIYYGYKPKWASHNSGVEFLPVKVKDLDPFYVTLT  
YSIWEYNNLPIINLAMETWITKSPDQTSVSSGDAEIMVWFYNNVLMPPGGQKXVDEFITVE  
INGVKQEAQWDVYFAPWSWDYLAFLRLTTPMKEGKVKFNVKDFVQKAAEVVKKHSTRIDN  
FEELYFCVWEIGTEFGDPNTTTAKFGWTFKDFVVEVVK (SEQ ID NO: 4)

>termocell3\_aa (Tcel3, ph1171) exocellulase 458 aa, GH5, MW 51,930, pI 6.47, charge -3.60

MEGNTILKIVLICTILAGLFGQVVPVYAENTTYQTPTGIYYEVRGDTIYMINVTSGEET  
PIHLFGVNWFGFETPNHVHGLWKRWNEDMLLQIKSLGFNAIRLPFCTESVKPGTQPIG  
IDYSKNPDLRGLDSLQIMEKIIKKAGDLGIFVLLDYHRIGCTHIEPLWYTEDFSEEDFI  
NTWIEVAKRFGKYWNVIGADLKNPHSVTSPPAAAYTDGTGATWGMGNPATDWNLAERI  
GKAILKVAPHWLI FVEGTQFTNPKTDSSYKGYNAWGGNLMVAVKDYVNLPRNKLVS  
PHVYGPDVYNQPYFGPAKGFDPDNLPIWYHHFGYVVKLELGYSVVIGFEGGKYGHGGDPR  
DVIWQNKLVDMWIENKFCDFYWSWNPDSGDTGGILQDDWTTIWEKYNLXRLMDSCS  
KSSSTQSVIRSTTPTKSNSTSKKICGPAILIILAVFSLLLRRAPR (SEQ ID NO: 6)

>termocell4\_aa (Tcel4, o-E1) exocellulase 514 aa (CODONS OPTIMIZED FOR RICE, WITHOUT STOP CODON), GH5, MW 59,980, pI 7.05, charge 0.30

MEIKLFCVFVIFILFSPFVIALSYPDVNYTAENGIIFVQNVTTGEEKPLYLHGVSWFG  
FELKDHVVYGLDKRNWKDILKDVKRLGFNAIRLPFCSESIRPDTRPSPERINYELNPDL  
KNLTSLEIMEKIIEYANSIGLYILLDYHRIGCEIEPLWYTENYSEEQYIKDWIFLAKR  
FGKYPNVIGADIKNEPHGEAGWGTGDERDFRLFAEKVGREILKVAPHWLI FVEGTQYTH  
VPNIDEIIEKKGWTFWGENLMGVKDYVRLPRGKVVYSPHVYGPSVYMMDYFKSPDFP  
NNMPIIWETHFGYLTDLNYTLVIGEWGGNYEGLDKVWQDAFVKWLIKKKIYNFFYWCIN  
PESGDTGGIFLDDWKTVNWEKMRVIYRLIKAANPEFEEPLYIILKTNATTSILGVGERI  
RIYWYTNKGVIDSNFAHSSEGEMNITVTKSMTLYIIVKKGNOQLRKLKLYVIGGNYGS  
NISTQLVTPKKGGERISTSLKLAISLLFILLFVWYLLREKH (SEQ ID NO: 8)

>termocell5\_aa (Tcel5, petroA) endoglucanase 328 aa, GH12, MW 38,226, pI 5.58, charge -6.60

METLLPVVVVDIEPVSMLRQRYKNKNSIKREKQGLIPLFFYFWVYLVLVLFANFQILNVN  
IFIIRCFLVIMVVLMTKPGTSDFVWNGIPLSMELNLWNIKEYSGSVAMKFDGEKVTFD  
ADIQNLSPKEPERYVLGYPEFYGYKPKWEKHTAEGSKLPVPVSSMKSFSVEVSFDIHHE  
PSLPLNFAMETWLTREKYQTEASIGDVEIMVWFYFNNLTPGGKKIEEFTIPVNLNGESV  
EGTWELWHAEWGWDYLAFLRKDPVKKGRVKFDVRFHFLDAAGKALSNSSTRVVKDFENLYFT  
VWEIGTEFGSPETKSAQFGWKFNFSIDLEVRE (SEQ ID NO: 10)

Figure 19A

GH12 31,818 5.66 -5.00

>termoel6\_aa (Tcel6) endoglucanase 284 aa (CODONS OPTIMIZED FOR CORN), GH12, MW 31,818, pI 5.66, charge -5.00
MLKLIPLVNGWYKLTQWEPLGGVHGADIECIHVTPNWNIDKSSVGTVQJEYEPQVQCL
RFSIDFPRIIRHNVGVAAYSEVIYGHKWPWGPTTCMDPQFKFPIKVNESKGLYSYVNYN
VKRSRPPDDSIENIAYDLWLTTSPLNTNGPQPGDVEVMIWLYYHGQRPAGRLIGELRMPI
TLGDSEAARDFEVWVADTGIGIGEWAVVTFRIKDPIKGGIIGVNLINYLIESAFKTLLEEL
NPVKWRYGDLNLYKNGIEFGSEFGNVSSGMIKLNWELCGLSLVKDSS (SEQ ID NO: 12)

>termocel9\_aa (Tcel9) endoglucanase 410 aa (CODONS OPTIMIZED FOR CORN), GH12, MW 45,059, pI 6.16, charge -2.20
MDYSINCSINPITLMVAHSSPLNPSNTLELTLILENGITTTVTVTATPRNTYPMISLGY
INITPNLWNLNTASSSGYASMYVDASQGALYIHVNFTKVYLNQQVGVAAAYSEFIYGYKP
WGLTSEAGGFNFPVKLTLELGSLLSPINYSLSISYSPQVAIFDWAYDLWLTTSPLNTNGP
QPGDVEVMIWLYYHLQQQPAQFPVANVTVP IWNVNGSLVNETFEVWIGSPQIEPGTHAIVS
FRPTNPIFRGLVGVNVTKFLQLAVNYLVTLVPSYWNVYTYLESKYLNGLIEFGSEWGNPST
YNITLNVVIYKAYLIKVPLESQGTVTVTYTTTSTMTVTSILATTSTVTTTSTLTSTV
TATSVSTSTVTQTLTTSIVKTVIPVYYTATIIVLLIIIAVVIALAFARRGIRVRLC (SEQ ID NO: 30)

>termocel10\_aa (Tcel10) endoglucanase 737 aa, GH65, MW 85,598, pI 7.80, charge 4.30
MRPQFGPSKEDEQVLGTLTLGNGQLGVRGFELEERSPYGTIVSGVYDTPYFYRELVN
GPRYIGMIIIDGELINPSSQKVKEFQRELDIEKGLLRTHLEIETKNGNKLKLYKSTRIV
HMKRKNLILLDFELKASKGGIAVVVNPIEFNTANPGFIDEIMI KHYRVDISKETEELGVY
ARVKTLDNKYTLIEASSLVPSEYTSRSTFRDNEIGBIYIVKLPKPKTYKPTKYVTVSK
GAALEELKDVKRLGFEEKLYEBEHINSWKRIWEKVKVEIEGDKDLENALNFNIFHLIQSLP
PTDKVSLPARGIHGFGYRGHIFWDFEYIYALPFFIFTMPKEARRLLLYRCNNLDAAKENA
KMNGYQGVQFPWESADDDGREATPSEIPLDMLGRKIVRIYTGEEHHITADIAYIVDFYY
QVSGDLEFPMNRCGLEIFETARFVASRVEFEEGKGYVIKKVIGPDEYHEHVNNFFTNL
MAKHNLLELAIRYPRESKNREPWKKIVEKLNIREEEVEKWBEEIAKNMYIPRKIDGVFEEF
DGYFELMDFEVDPPFNIGEKTLPEEIRNNGIKTKLVKQADVIMAQYLLKDYFSPBEEIKSN
FNYYIRRTTHASSLSMPPYAIATWIGEVKIAEYEFKRCANIDLQNVYGNATAEGFHLAT
AGGTWQVLRVGRGCLNVKGNKIELNPNLPEKWKYVKFRIFFKGSWIEFKISRKKVRARM
LEGSRKVKISSPGKEVDLYPGKEVVIVAN (SEQ ID NO: 32)

>termocel7\_aa (Tcel7) beta-glucosidase 722 aa, GH3, MW 81,243, pI 5.38, charge -16.9
MMGKIDEILSQTIEEKVKLVVGVGLPGLFGNPHSRVAGAAGETHPVPRLGIPSPVLAD
GPAGLRINPTRENDENTYYTTAFPVEIMLASTWNKDLEEVGKAMGEEVREYGVVDVLLA
PAMNIHRNPLCGRNPEYYSSEDPVLSGEMASAFVKGVQSQCAGACIKHFVANNQETNRMV
VDTIVSERALREIYLKGFELAVKARPWTVMSAYNKLNGKYCSQNEWLLKVLREEWGF
DGFVMSDWYAGDNPEVQLKAGNDMIMP GKAYQVNTERRDEIEEIMEALKEGRLSEEVLN
ECVRNLIKVLVNAFSPFKGYRYSNKPDLSEHAKVAYEAGVEGVVLENNGVLPFDES IHV
AVFGTGQIETIKGGTSGSDTHPRYTI SILEGIKERNMKPFDEELTSIYEDYIKKMETBEE
YKPRTDSWGTVIKPKLPENFLSEKEIKKAAKNDAAVVVISRISGEGYDRKPKVKGDFYL
SDDELELIKTVSRFHEQGKVVVLLNIGSPIEVASWRDLVDGILLVWQAGQEMGRIVA
DVLVGRVNPSPGKLPPTFPKDYSDVPSWTFPGSPKDNFORVVEEDYVGYRYDYDFGVE
PAYEFGYGLSYTKFEYKDLKIAIDGDILRVSYTINTGDRAGKEVSQVYVVKAPKGIKDK
PFOELKAFHKTKLLNPGESEKIFLEIPLRDLASFDGKEWVVESEGEYEVVVGASSRDIRL
RDIFLVEGEKRFK (SEQ ID NO: 14)

Figure 19B

>termocel6\_aa (Tcel6) beta-glucosidase 446 aa, GH1, MW  
51509, pI 5.84, charge -9.1  
MNVKKEPPEGFLWGVATASYQIEGSPADGAGMSIWHTFSHTPGNVKNNGDTGDVACDHYN  
RWKEDIIEIEKLGVKAYRFSISWPRILPEGTGRVNQKGLDPYNRIIDTLLEKGITPFVT  
IYHWDLPFALQLKGGWANREIADWFAEYSRVLPENFGDRVKNWITLNEPWWVAIVGHLY  
GVHAPGMRDIYVAFRAVHNLLRAHAKAVKVFRET'VKDGKIGIVPNNNGYFEPASEKEEDI  
RAARFMHQFNNYPLFLNPIYRGDYPELVLEFAREYLPENYKDDMSEIQEKIDFVGLNYY  
SGHLVKFDPDAPAKVSFVERDLPKTAMGWEIVPEGIYWILKKVKEEYNPPEVYITENGA  
AFDDVVSSEDGRVHDQNRIDYDKAHIQOAWKAIQEGVPLKGYFVWSLLDNFEWAEGYSKR  
FGIVYVDYSTQKRIIKDSGYWYSNVVKSNSLED (SEQ ID NO: 16)  
>pBAD MYC-HIS TAG 21 aa, MW 2,513, pI 6.20 charge -2.60  
EQKLISEEDLNSAVDHHHHHH (SEQ ID NO: 18)

Figure 19C

1

## THERMOCELLULASES FOR LIGNOCELLULOSIC DEGRADATION

### BACKGROUND OF THE INVENTION

#### 1. Field of the Invention

This invention generally relates to thermostable enzymes capable of degrading (hydrolyzing) cellulose at high temperatures, and the incorporation of nucleic acids coding for one or more of such enzymes into a host, and, more particularly, a host that produces or is composed of cellulosic material.

#### 2. Background of the Invention

Cellulose is a polysaccharide consisting of a linear chain of several hundred to over nine thousand  $\beta$  (1 $\rightarrow$ 4) linked D-glucose units [formula (C<sub>6</sub>H<sub>10</sub>O<sub>5</sub>)<sub>n</sub>]. Cellulose is the most abundant organic compound on earth, making up about 33 percent of all plant matter, about 50 percent of wood, and about 90 percent of products such as cotton. In nature, cellulose is present as part of the lignocellulosic biomass of plants, which is composed of cellulose, hemicellulose, and lignin. The carbohydrate polymers (cellulose and hemicelluloses) are tightly bound to the lignin, by hydrogen and covalent bonds.

Many highly desirable products are derived from lignocellulosic biomass. In particular, much interest has recently been focused on recapturing the saccharide building blocks locked in plant biomass for biofuel production. For example, fermentation of plant biomass to ethanol is an attractive carbon neutral energy option since the combustion of ethanol from biomass produces no net carbon dioxide in the earth's atmosphere. Further, biomass is readily available, and its fermentation provides an attractive way to dispose of many industrial and agricultural waste products. Finally, plant biomass is a highly renewable resource. Many dedicated energy crops can provide high energy biomass, which may be harvested multiple times each year.

One barrier to the production of products from biomass is that the cellulosic polymer has evolved to resist degradation and to confer hydrolytic stability and structural robustness to the cell walls of plants. This robustness or "recalcitrance" is due largely to extensive intermolecular hydrogen bonding between cellulose polymer chains. Some organisms, notably fungi, bacteria, and protozoans, but also some plants and animals, have evolved the ability to digest cellulose. In vivo cellulose breakdown typically entails the cooperative interaction of several cellulases, enzymes that catalyze the cellulolysis (hydrolysis) of cellulose. Several different kinds of cellulases, which differ structurally and mechanistically, are known, and some of these have been isolated, characterized and used to break down cellulose in vitro. General categories of cellulases include: endo-cellulases (endoglucanases), which randomly hydrolyze internal bonds to disrupt the crystalline structure of cellulose, thereby exposing individual cellulose polysaccharide chains; and exo-cellulases (exo-processive-endoglucanases), which cleave 2-4 units from the ends of the exposed chains produced by endocellulases to produce tetrasaccharides or disaccharides such as cellobiose. Two major types of exo-cellulases are known, one of which works processively from the reducing end, and one of which works processively from the non-reducing end of cellulose. A third major type of cellulase is cellobiase or beta-glucosidase, which hydrolyses exo-cellulase products such as cellobiose into individual glucose monosaccharides.

Typically, the digestion of cellulose is carried out at temperatures approaching 100° C. because, at high temperatures, intermolecular hydrogen bonds are disrupted and recalcitrant cellulose polymers become accessible to the cellulase

2

enzymes. Therefore, cellulases used commercially in such processes must be able to withstand very high temperatures, preferably for extended periods of time.

There is an ongoing need to identify, isolate and characterize cellulases, especially thermally stable cellulases, for use in the enzymatic hydrolysis of cellulose. Of particular interest is the development of groups or systems of cellulases that include enzymes with endo-cellulase, exo-cellulase and beta-glucosidase activity, the enzymes in the system acting in concert to carry out the complete hydrolysis of cellulose to glucose at high temperatures.

### SUMMARY OF THE INVENTION

Protein sequences which heretofore were not recognized as having enzymatic activity have been isolated and characterized as thermostable enzymes capable of degrading (hydrolyzing) cellulose at high temperatures. The activity is referred to herein as cellulase or cellulase-like. The enzymes, originating from Archaea and various thermophilic bacteria, include: endoglucanases that randomly hydrolyze internal glycosidic bonds; exo-processive-endoglucanases that split off cellobiose dimers; and  $\beta$ -glucosidases that reduce cellobiose into monomeric glucose molecules. While the  $\beta$ -glucosidase enzymes are technically not "cellulases" because cellobiose (not cellulose) is the substrate they cleave, the three groups of enzymes may be sometimes collectively referred to as "cellulases" herein. The enzymes are optimally catalytically active at temperatures at or above about 85° C. and retain >85% of their enzymatic activity even after a 5 day incubation at elevated temperature, e.g. 90° C. In some embodiments, the enzymes, or enzyme systems or groupings comprising multiple thermostable catalytic activities may advantageously be used to degrade cellulose. Preferably, in the case of systems which have multiple thermostable catalytic activities, such a system comprises at least one endoglucanase, at least one exo processive-endoglucanase, and at least one beta-glucosidase enzyme, and thus can carry out the complete hydrolysis of cellulose to glucose at high temperatures in a sequential, cooperative manner. Catalytic consolidation at high-temperatures using the enzyme systems described herein is not additive but synergistic, accessing recalcitrant cellulose and hydrolyzing beta linkages at temperatures above 85° C. Thus, one aspect of the invention is to employ the enzymes, alone or in a group, in processes to break down cellulosic material by contacting the cellulosic material with the enzymes and elevating the temperature to activate the enzymes to break down the cellulosic material. These processes might be performed, for example, in tanks where the cellulosic material is distributed in a liquid carrier; however, the enzymatic breakdown may be achieved simply through elevating the temperature of the cellulosic material with the enzymes being in contact with the cellulosic material.

The invention also contemplates the incorporation of nucleic acids coding for one or more of the enzymes into a host (e.g., a plant, fungi, bacterium or animal). In the case where the host produces or is composed of cellulosic material (e.g., plants such as corn, switch grass, sugar cane, sorghum, pinus and eucalyptus), the host can be subjected to breakdown of the cellulosic material, for example, after harvest. That is, in a particular example, corn or switchgrass transformed to include nucleic acids coding for the enzymes will express the enzymes internally, and after collection or harvest of the corn or switchgrass, the enzymes can be activated to

begin and preferably ultimately to completely degrade the cellulose simply by elevating the temperature of the corn or switchgrass.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1A-B. *Pyrococcus furiosus* Termocel 1 endoglucanase. A, nucleotide sequence (903 bp, SEQ ID NO: 1); and B, amino acid sequence (301 aa, SEQ ID NO: 2). Nucleotide sequence is optimized for expression in corn and shown without N-terminal signal peptide encoding sequence.

FIG. 2A-B. *Thermotoga petrophila* Termocel 2 endoglucanase. A, nucleotide sequence (825 bp, SEQ ID NO: 3); and B, amino acid sequence (274 aa, SEQ ID NO: 4).

FIG. 3A-B. *Pyrococcus horikoshii* Termocel 3 exocellulase. A, nucleotide sequence (1377 bp, SEQ ID NO: 5); and B, amino acid sequence (458 aa, SEQ ID NO: 6).

FIG. 4A-B. *Pyrococcus abyssi* Termocel 4 exocellulase. A, nucleotide sequence (1542 bp, SEQ ID NO: 7); and B, amino acid sequence (514 aa, SEQ ID NO: 8). Nucleotide sequence is optimized for expression in rice and shown without stop codon.

FIG. 5A-B. *Thermotoga petrophila* Termocel 5 endoglucanase. A, nucleotide sequence (987 bp, SEQ ID NO: 9); and B, amino acid sequence (328 aa, SEQ ID NO: 10).

FIG. 6A-B. *Caldivirga maquiligenesis* Termocel 6 endoglucanase. A, nucleotide sequence (852 bp, SEQ ID NO: 11); and B, amino acid sequence (284 aa, SEQ ID NO: 12). Nucleotide sequence is optimized for expression in corn.

FIG. 7A-B. *Thermotoga petrophila* Termocel 7 beta-glucosidase. A, nucleotide sequence (2169 bp, SEQ ID NO: 13); and B, amino acid sequence (722 aa, SEQ ID NO: 14).

FIG. 8A-B. *Thermotoga petrophila* Termocel 8 beta-glucosidase. A, nucleotide sequence (1341 bp, SEQ ID NO: 15); and B, amino acid sequence (446 aa, SEQ ID NO: 16).

FIG. 9. Characteristics of high-temperature operating thermo-stable cellulases. swAvicel=phosphoric acid swollen Avicel; avCellulose=Avicel; cellulase specific activity is expressed as of reducing sugar/mg protein/day at 85° C., pH 6; beta-glucosidase specific activity is expressed as nM p-nitrophenol (pN)/μg protein/minute.

FIG. 10A-E. Processive exocellulases (cellobiohydrolase). Capillary zone electrophoresis (CZE) of 8-aminonaphthalene-1,3,6 trisulfonic acid (ANTS)-labeled cellopento breakdown products incubated with Termocel 1 (A), Termocel 2 (B) and Termocel 3 (C). CZE retention times (D) of purified monomer, (DPI) dimer (DP2), trimer (DP3) and the substrate cellopento (DP5). Sequential predicted cleavage pattern (E) between DP5 and DP4, DP4 and DP3, DP3 and DP2. Assay conditions, Substrate, ANTS-cellopento (FIG. 9C), buffer sodium phosphate/citrate 50 mM, incubated at 95° C., pH 6.

FIG. 11A-E. Processive endocellulases (endoglucanase). Capillary zone electrophoresis (CZE) of ANTS-labeled cellopento breakdown products incubated with Termocel 4 (A), Termocel 5 (B) and Termocel 6 (C). CZE retention times (D) of purified monomer, (DPI) dimer (DP2), trimer (DP3) and the substrate cellopento (DP5). Predicted cleavage pattern (E) between DP2 ad DP3 and DP3 and DP4. Assay conditions, Substrate, ANTS-cellopento (FIG. 9C), buffer sodium phosphate 50 mM, incubated at 95° C., pH 6.

FIG. 12. Temperature optima for high-temperature catalytic cellulases, and activities at 60, 45 and 20° C. swAvicel=phosphoric acid swollen Avicel; avCellulose=Avicel; cellulase specific activity is expressed as μM of reducing sugar/mg protein/day at 85° C.,

pH 6; beta-glucosidase specific activity is expressed as nM p-nitrophenol (pN)/μg protein/minute.

FIG. 13A-D. Termocel thermostability. Termocels were incubated at 90° C. in phosphate/citrate buffer for the indicated number of hours and CMC or PNP activity determined. The amount of residual activity is shown. A, Termocels 1 and 2; B, Termocels 3 and 4; C, Termocels 5 and 6; D, Termocels 7 and 8. With the exception of Termocel 3, all cellulases retained >80% of activity after 120 hrs at 90° C. Thus, these enzymes are stable at high temperatures.

FIG. 14. Flow chart illustrating cellulose treatment steps.

FIG. 15. Table depicting biomass substrate specificity of particular Termocels.

FIG. 16A-B. *Caldivirga maquiligenesis* Termocel 9 endoglucanase. A, nucleotide sequence (1230 bp, SEQ ID NO: 29); and B, amino acid sequence (410 aa, SEQ ID NO: 30). Nucleotide sequence is optimized for expression in corn.

FIG. 17A-B. *Pyrococcus horikoshii* Termocel 10 endoglucanase. A, nucleotide sequence (2214 bp, SEQ ID NO: 31); and B, amino acid sequence (737 aa, SEQ ID NO: 32).

FIG. 18A-F. Nucleotide sequences as set forth in SEQ ID NO: 1 (from *Pyrococcus furiosus*), SEQ ID NO: 3 (from *Thermotoga petrophila*), SEQ ID NO: 5 (from *Pyrococcus horikoshii*), SEQ ID NO: 7 (from *Pyrococcus abyssi*), SEQ ID NO: 9 (from *Thermotoga petrophila*), SEQ ID NO: 11 (from *Caldivirga maquiligenesis*), SEQ ID NO: 29 (from *Caldivirga maquiligenesis*), SEQ ID NO: 31 (from *Pyrococcus horikoshii*), SEQ ID NO: 13 (from *Thermotoga petrophila*), and SEQ ID NO: 15 (from *Thermotoga petrophila*).

FIG. 19A-C. Amino acid sequences as set forth in SEQ ID NO: 2 (from *Pyrococcus furiosus*), SEQ ID NO: 4 (from *Thermotoga petrophila*), SEQ ID NO: 6 (from *Pyrococcus horikoshii*), SEQ ID NO: 8 (from *Pyrococcus abyssi*), SEQ ID NO: 10 (from *Thermotoga petrophila*), SEQ ID NO: 12 (from *Caldivirga maquiligenesis*), SEQ ID NO: 30 (from *Caldivirga maquiligenesis*), SEQ ID NO: 32 (from *Pyrococcus horikoshii*), SEQ ID NO: 14 (from *Thermotoga petrophila*), and SEQ ID NO: 16 (from *Thermotoga petrophila*).

#### DETAILED DESCRIPTION

The present invention is based on the identification and characterization of a comprehensive set of thermostable cellulases that work in concert to catalyze the hydrolysis of cellulose to glucose at very high temperatures. The cellulases, originally identified in archeal and bacterial genomes, include endoglucanases that randomly cleave internal glycosidic bonds; exo-processive-endoglucanases that further hydrolyze cellulose fragments into cellobiose dimers; and β-glucosidases that further hydrolyze cellobiose dimers to glucose monomers. While the enzymes may be used individually, in some embodiments of the invention they are grouped to form a cooperative enzyme system. By combining into a group at least one endoglucanase, at least one exo-processive-endoglucanase and at least one β-glucosidase, an enzyme system is formed that is capable of the complete breakdown of cellulose to glucose. Importantly, the enzyme system's various catalytic activities are optimal at temperatures that are high enough to destabilize the hydrogen bonds between crystalline cellulose strands (e.g. at temperatures greater than 80° C.). Destabilization of hydrogen bonds at high temperatures causes disruption of the crystalline structure of cellulose, thereby facilitating access by the first enzyme in the series (endoglucanase) to internal glycosidic bonds of individual cellulose polymer strands, and allowing the step-wise process of cellulose breakdown to begin.



Exemplary amino acid sequences of the recombinant enzymes of the invention and exemplary nucleotide sequences that encode them are depicted in FIGS. 1-8. However, those of skill in the art will recognize that the invention also encompasses variant proteins comprising amino acid sequences that are based on or derived from the sequences disclosed herein. By an amino acid sequence that is "derived from" or "based on" the sequence disclosed herein, we mean that a derived sequence (or variant sequence) displays at least about 50 to 100% identity to an amino acid sequence disclosed herein, or about 60 to 100% identify, or about 70 to 100% identity, or even from about 80 to 100% identify. In preferred embodiments, a variant sequence displays from about 90 to 100% or about 95 to 100% amino acid identity. In further preferred embodiments, a variant sequence is 95, 96, 97, 98 or 99% identical to at least one sequence disclosed herein. Variations in the sequences may be due to a number of factors and may include, for example: conservative or non-conservative amino acid substitutions; natural variations among different populations as isolated from natural sources; various deletions or insertions (which may be amino terminal, carboxyl terminal, or internal); addition of leader sequences to promote secretion from the cell; addition of targeting sequences to direct the intracellular destination of a polypeptide; etc. Such alterations may be naturally occurring or may be intentionally introduced (e.g. via genetic engineering) for any of a wide variety of reasons, e.g. in order to eliminate or introduce protease cleavage sites, to eliminate or introduce glycosylation sites, in order to improve solubility of the polypeptide, to facilitate polypeptide isolation (e.g. introduction of a histidine or other tag), as a result of a purposeful change in the nucleic acid sequence (see discussion of the nucleic acid sequence below) which results in a non-silent change in one or more codons and thus the translated amino acid, in order to improve thermal stability of the protein, etc. All such variant sequences are encompassed by the present invention, so long as the resulting polypeptide is capable of catalyzing the enzyme activity of the original protein as disclosed herein. For example, the invention includes shorter portions of the sequences that also retain the catalytic activity of the enzyme. The full-length protein sequences and/or active portions thereof are both referred to as polypeptides herein. In addition, the invention also includes chimeric or fusion proteins that include, for example: more than one of the enzymes disclosed herein (or active portions thereof); or one or more of the enzymes disclosed herein (or portions thereof) plus some other useful protein or peptide sequence(s), e.g. signal sequences, spacer or linker sequences, etc.

The invention also comprehends nucleic acid sequences that encode the proteins and polypeptides of the invention. Several exemplary nucleic acid sequences are provided herein. However, as is well known, due to the degeneracy of the nucleic acid triplet code, many other nucleic acid sequences that would encode an identical polypeptide could also be designed, and the invention also encompasses such nucleic acid sequences. Further, as described above, many useful variant forms of the proteins and peptides of the invention also exist, and nucleic acid sequences encoding such variants are intended to be encompassed by the present invention. In addition, such nucleic acid sequences may be varied for any of a variety of reasons, for example, to facilitate cloning, to facilitate transfer of a clone from one construct to another, to increase transcription or translation in a particular host cell (e.g. the sequences may be optimized for expression in, for example, corn, rice, yeast or other hosts), to add or replace promoter sequences, to add or eliminate a restriction cleavage site, etc. In addition, all genera of nucleic acids (e.g.

DNA, RNA, various composite and hybrid nucleic acids, etc.) encoding proteins of the invention (or active portions thereof) are intended to be encompassed by the invention.

The invention further comprehends vectors, which contain nucleic acid sequences encoding the polypeptides of the invention. Those of skill in the art are familiar with the many types of vectors, which can be useful for such a purpose, for example: plasmids, cosmids, various expression vectors, viral vectors, etc.

Production of the nucleic acids and proteins of the invention can be accomplished in any of many ways that are known to those of skill in the art. The sequences may be synthesized chemically using methods that are well-known to those of skill in the art. Alternatively, nucleotide sequences may be cloned using, for example, polymerase chain reaction (PCR) and/or other known molecular biology and genetic engineering techniques. Recombinant proteins may be made from a plasmid contained within a bacterial host such as *Escherichia coli*, in insect expression systems, yeast expression systems, plant cell expression systems, etc. Further, the nucleic acid sequences may be optimized for expression in a particular organism or system. To that end, the present invention also encompasses a host cell that has been transformed or otherwise manipulated to contain nucleic acids encoding the proteins and polypeptides of the invention, either as extra-chromosomal elements, or incorporated into the chromosome of the host. In particular, in the practice of the present invention, nucleic acid sequences encoding one or more of the cellulases (e.g. an entire "system" as described herein) may be introduced into plant cells, seeds, etc., to generate recombinant plants that contain the nucleic acids.

Plant transformation to incorporate one or more nucleic acids coding for one or more cellulase enzymes described herein can be accomplished by a variety of techniques known to those of skill in the art. Plant transformation is the introduction of a foreign piece of DNA, conferring a specific trait, into host plant tissue. Plant transformation can be carried out in a number of different ways; *Agrobacterium* mediated transformation, particle bombardment, electroporation and viral transformation.

Suitable examples of plants that may be transformed to include one or more cellulase enzymes or sets of enzymes include but are not limited to rice, corn, various grasses such as switchgrass, sugar cane, sorghum, pinus and eucalyptus, etc. Advantages of genetically engineering plants to contain and express the cellulase genes include but are not limited to the availability of the enzymes within the cell wall tissues (cellulosic fibers) and ready to be activated by high temperatures (e.g., heating to 70 or 80 C or more). Deposition of these enzymes produced by the plant cells and targeted to the apoplast, should largely overcome the recalcitrant nature of biomass.

The cellulases and/or cellulase enzyme systems of the invention may be used for the breakdown (catalysis) of cellulose in biomass from a wide variety of sources. Biomass comes in many different types, which may be grouped into four main categories: (1) wood residues (including sawmill and paper mill discards); (2) municipal paper waste; (3) agricultural residues (including corn stover and sugarcane bagasse); and (4) dedicated energy crops, which are mostly composed of fast growing tall, woody grasses. Cellulose-containing biomass from any of these or other sources may be acted upon by the enzymes and consolidated enzyme systems of the invention.

Generally, the breakdown of cellulose will be complete, i.e. the endproduct is glucose. This is especially true when a consolidated enzyme system that includes at least three dif-

ferent types of enzymes (for example, an endoglucanase, an exo-processive-endoglucanases, and a  $\beta$ -glucosidase) are employed. However, this need not always be the case. Depending on the goal of the reaction, only one enzyme may be utilized (e.g. an endoglucanase to generate randomly cleaved cellulose polymers); or only two enzymes may be utilized (e.g. an endoglucanase and an exo-processive-endoglucanases to generate dimeric disaccharides such as cellobiose), etc. Any desired grouping of the enzymes of the invention may be utilized to generate any desired endproduct that the enzymes are capable of producing from a suitable substrate. Further, one or more of the enzymes of the invention may be used in combination with other cellulases, or with enzymes having other types of activities. In one embodiment of the invention, a "system" could further include a yeast or other organism capable of fermenting glucose to e.g. ethanol.

The cellulases of the invention have very high temperature optima, an optimal temperature being the temperature at which an enzyme is maximally active (e.g. as an endoglucanase, an exo-processive-endoglucanases, or a  $\beta$ -glucosidase), as determined by a standard assay recognized by those of skill in the art. As described in the Examples section below, the lowest temperature optimum for an enzyme of the invention is about 85° C., and the highest temperature optimum is about 102° C. Further, the enzymes of the invention are thermally stable, i.e. they are capable of retaining catalytic activity at high temperatures (e.g. at their temperature maximum, or at temperatures that deviate somewhat from the maximum) for extended periods of time, for example, for at least for several hours (e.g. 1-24 hours), and in many cases, for several days (e.g. from 1-7 days or even longer). By "retain catalytic activity" we mean that the enzyme retains at least about 10, 20, 30, 40 or 50% or more of the activity displayed at the beginning of the extended time period, when measured under standard conditions; and preferably the enzyme retains 60, 65, 70, 75, 80, 85, 90, 95, or even 100% of the activity displayed at the beginning of the extended time period.

The enzymes of the invention are generally employed in reactions that are carried out at temperatures at or near those which are optimal for their activity. Some enzymes may be used over a wide temperature range (e.g. at a temperature that is about 50, 40, 30, 20, 10, 5 or fewer degrees lower than (below) the temperature optimum, and up to about 5, 10, 15, or more degrees greater than (above) the temperature optimum. For other enzymes, the range may be more restricted, i.e. they may display catalytic activity within a narrow temperature range of only less than about 10, or less than about 5, or fewer degrees of their optimal catalytic temperature. When carrying out a cellulose digestion reaction, the enzymes may be used one at a time sequentially (i.e. one enzyme is added, reaction occurs, and then another enzyme is added, with or without removal of the previous enzyme, and so on), or the reaction mixture may contain two or even all three of the enzymes (an enzyme system) may be added at the same time. When designing groups of enzymes to be included in an enzyme system, those of skill in the art will recognize that a suitable temperature at which all enzymes in the group are active will be selected as the temperature for reaction. Or, conversely, if it is desired to carry out a reaction at a particular temperature, enzymes with optimal activity at or near that temperature would be selected for inclusion in the set. For example, for a reaction to be carried out at 97° C., one might choose a set of enzymes that includes Termocel 5 (endocellulase, optimum=96° C.), plus Termocel 2 (exocellulase, optimum=98° C.), plus Termocel 7 ( $\beta$ -glucosidase, optimum=98° C.); whereas for a reaction that is to be carried out at 90° C., one might choose Termocel 6 (endocellulase, opti-

mum=85° C.), plus Termocel 3 (exocellulase, optimum=94° C.), plus Termocel 7 ( $\beta$ -glucosidase, optimum=92° C.). If an enzyme is used individually, the reaction may be carried out at a temperature near its optimum, or at which the enzyme retains sufficient activity to be useful. In addition, the selection of a reaction temperature may be based on other considerations, e.g. safety or other practical considerations of high temperature operations, or concerns about the cost of keeping a reaction mixture at a high temperature, the temperature used for preparing biomass for the reaction, the temperature of procedures that follow the reaction, etc. Generally, the degradation of cellulose will be carried out at a temperature in the range of from about 70 to about 95° C.

The invention also provides methods of use of the enzymes disclosed herein. The methods generally involve the use of at least three enzymes of the invention, at least one from each of the three classes endoglucanase, exo-processive-endoglucanases, and  $\beta$ -glucosidase. The three classes of enzymes act in concert to sequentially breakdown cellulose to glucose. The methods of the invention may be carried out for any purpose for which it is desirable to prepare glucose (or other products produced by the enzymes), and further metabolize into other chemicals, such as ethanol, xylitol, butanol, amino acids, glycol etc.

Generally, such methods are carried out by first pretreating a cellulose-rich feedstock by removing the lignin (usually through ball milling). The production of sugars (saccharification) of the pretreated cellulose is carried out by suspending the pretreated cellulose in a cellulase broth that contains suitable cellulase enzymes such as those disclosed herein. Generally, the reaction will be carried out at a temperature in the range of from about 70 to about 95 C, and the length of time for a reaction will be in the range of from about one hour to about six days. Reactions are carried out in media such as aqueous buffered to a suitable pH, e.g. in the range of from about pH 4 to about pH 9.

Thereafter, the desired products (e.g. glucose and cellobiose) may be harvested from the broth, or the reaction products may be further processed. For example, for the production of ethanol, fermentation of the glucose in the broth may be carried out by known conventional batch or continuous fermentation processes, usually using yeast. Ethanol may be recovered by known stripping or extractive distillation processes. This process is illustrated schematically in FIG. 14, which shows the steps of pretreating biomass to provide a source of cellulose; contacting the cellulose with one or more cellulase enzymes of the invention to hydrolyze cellulose to glucose, and fermenting the glucose to produce ethanol.

## EXAMPLES

### Example 1

#### Isolation and Characterization of Cellulases that Catalyze High-Temperature Thermo-Stable Bio-Consolidated Cellulose Breakdown

##### Abstract

Cellulose breakdown entails cooperative interaction of various cellulases by accessing and cleaving the recalcitrant cellulosic polymer. At high temperatures, most of the recalcitrant biomass polymers become enzymatically accessible because of intermolecular hydrogen bond disruption. Here, we describe a high-temperature operating thermo-stable cellulose enzyme system, consisting of endoglucanases, exoprocessive-endoglucanases and beta-glucosidases. Two catalytic types of cellulose cleaving

enzymes was found: endoglucanases that randomly hydrolyze internal glycosidic bonds and exo-processive-endoglucanase, which split off cellobiose dimers. Finally, a third activity,  $\beta$ -glucosidase, reduces cellobiose into glucose molecules. The consolidated enzyme system operates optimally at temperatures above 85° C. and retains >85% of its enzymatic activity after a 5 day incubation at 90° C. Catalytic consolidation with high-temperatures is not additive but synergistic, accessing recalcitrant cellulose and hydrolyzing beta linkages above 85° C.

#### Introduction

Cellulose is an abundant biopolymer component of plant cell walls. Cellulose is a linear biopolymer of D-glucose, linked by  $\beta$ -1,4-glucosyl linkages. Cellulosic enzyme systems

completely hydrolyze cellulose rendering glucose molecules. A cellulosic enzymatic system consists of multiple cellulases, endo- $\beta$ -glucanase, cellobiohydrolase and  $\beta$ -glucosidase, which interact synergistically in producing glucose. Endoglucanases randomly hydrolyze the internal glycosidic bonds to decrease the length of the cellulose chain. Cellobiohydrolases are exo- or endo-processive enzymes that split off cellobiose of the shortened cellulose chains. Cellobiose is hydrolyzed by  $\beta$ -glucosidase to glucose.

Native cellulose molecules appear predominantly as crystalline cellulose, which shows a high degree of intermolecular hydrogen bonding explaining its remarkable stability and recalcitrance to enzymes. Thus disrupting crystal intermolecular hydrogen bonds through cellulose swelling and dissolution with high-temperature operating cellulases overcomes recalcitrance and result in enzymatic digestion of native cellulose.

#### Results

##### Isolation and Characterization of High-Temperature Operating and Thermostable Cellulases

A series of ten high-temperature operating and thermostable cellulases were identified through bioinformatics driven searches of archeal and bacterial genomes. The corresponding genes were genetically manipulated to adapt expression to a laboratory tractable system (*Escherichia coli*) by codon optimization and usage controlled promoters. Individual proteins were expressed and isolated (purified) from *E. coli* crude extracts and analyzed for activity and other physical and chemical properties. Data presented in tabular form in FIG. 9 describes the eight enzymes isolated in this study.

Termocel 1 and 2, group into a class with similar physical and catalytic properties, they exhibit a molecular weight of 34,005 and 31,930 D, a pI of 4.8 and 4.77 and a net charge at pH 7 of -13.10 and -13.30, respectively. They appear to function through an exo-processive-endoglucanase cleaving pattern with a specific activity on Avicel of 63.4 and 8.1 U and on swollen cellulose of 13.6 and 2.2 U, respectively.

Termocel 3 and 4 differ slightly with a molecular weight of 51,930 and 59,980 D, a pI of 6.47 and 7.05 and a net charge at pH 7 of -3.60 and 0.30, respectively. These enzymes also seem not to overlap with their predicted mode of operation, one exoprocessive type and the other as an endoglucanase with specific activity on Avicel of 48.5 and 6.8 and on swollen cellulose of 8.4 and 2.2 U, respectively.

Termocel 5 and 6 fall in a third class with similar physical and catalytic properties. They exhibit a molecular weight of 38,226 and 31,818 D, a pI of 5.58 and 5.66 and a net

charge at pH 7 of -6.60 and -5.00, respectively. They appear to function through an internal cleaving pattern (endoglucanase) with a specific activity on Avicel of 34.1 and 20.6 U and on swollen cellulose of 6.8 and 5.1 U respectively.

Termocel 7 and 8 are  $\beta$ -glucosidases with distinct physical properties but similar catalytic activity. They exhibit a molecular weight of 81,243 and 51,509 D, a pI of 5.38 and

5.84 and a net charge at pH7 of -16.90 and -9.10, respectively. They cleave cellobiose with a specific activity on pNPG of 69.4 and 60.9 U, respectively.

Termocel 9 and 10 are endocellulases that exhibit a molecular weight of 45,059 and 85,598 D, a pI of 6.16 and 7.80 and a net charge at pH 7 of -2.20 and 4.30, respectively. They have a specific activity on Avicel of 5.2 and 4.9 U and on swollen cellulose of 1.4 and 1.5 U respectively.

#### Mode of Operation

FIGS. 10 and 11 describe the mode of operations of all six cellulases. Termocel 1, 2 and 3 are cellulases that function by sequentially cleaving glucose residues of the non-reducing end of a polymeric substrate. FIG. 10 shows the sequential depolymerization breakdown products through capillary zone electrophoresis.

Termocel 4, 5 and 6 are cellulases that function by internally cleaving a multimeric substrate. FIG. 11 shows trimeric and dimeric breakdown products, indicating internal cleavage of the pentameric substrate.

#### High-Temperature Catalytic Operation

FIG. 12 shows the optimum temperature of operation of eight Termocels in tabular form. The highest optimum was found for Termocel 1 with an optimum of 102° C. and the lowest optimum was found to be Termocel 6 with 85° C. At 60° C., all Termocels lost at least 40% of their activity (except Termocel 4) and at 20° C. the Termocels operated with less than 20% of their optimum activity.

Among the beta-glucosidases, no significant differences between activity and temperature optimum were apparent. However, catalytic inactivation at lower temperatures (45 and 20° C.) to levels below 1% residual activity for Termocel 7 is remarkable.

#### Thermal Stability

Thermostability of the Termocels was evaluated to determine the working time frame with useful enzymatic activity at high-temperatures. Enzymes were incubated at 90° C. for up to 5 days and then assayed for CMC (endo-glucanase and exo-cellulase) or PNPGE (beta-glucosidase) activity and results are reported as % of residual activity in FIG. 13. With the exception of Termocel 3, all enzymes retained over 80% of their initial enzymatic activity after a 5-day incubation period at 90° C.

#### Modes of Use

These high-temperature operating cellulases can be used in all processes in which cellulose degradation at high temperatures is desired. These applications include but are not restricted to food processing, feedstuff preparation, textile finishing and paper pulping. The consolidated enzyme system is useful to hydrolyze fibrous crystalline cellulosic biomass materials, at high temperatures with Termocel 1, 2, 3, 4, 5, 6, 7 and 8 to produce high-sugar containing fermentation broths. In addition the genes of the high-temperature operating enzyme system can be used in producing transgenic organisms capable of expressing one or more high-temperature operating and thermostable plant cell wall degrading enzymes.

## Methods

## Cloning

Genomic DNA of *Pyrococcus horikoshii* OT3 served as the PCR template for the amplification of the PHI 171 gene. Likewise, genomic DNA of *Thermotoga petrophila* RKU-1 served as PCR template for the cloning of the PetroA, PetroB, Tpet\_0898 and Tpet\_0952 genes. Primer sequences are shown in Table 1. Restriction sites were introduced (bold letters). The O-eglA, ZP and E1 genes were synthesized without using a DNA template; the codons of the three genes were also optimized according to the sequences of corn and rice genomes (FIGS. 1A, 4A and 6A). All gene segments generated were cloned into the NcoI and XbaI sites of the pBAD/Myc-His vector (Invitrogen), which carries a fusion sequence (GAACAAAACTCA TCTCAGAAG AGGATCTGAAT-AGCGCCGTCGACCATCATCATCATCATCAT, SEQ ID NO: 17) encoding six histidine residues at the C-terminus of any protein expressed from the vector (EQKLISEEDLN-SAVDHHHHHH, SEQ ID NO: 18). The expression plasmids were used to transform *Escherichia coli* TOP 10F' (Invitrogen). All constructs were verified by DNA sequencing.

TABLE 1

Oligonucleotide sequences used in this study.		
Primer	Sequence (5' → 3') <sup>a</sup>	SEQ ID NO:
Termocel 3	ATAT <b>CCATGG</b> AGGGGAATACTATTCTTAAATC GTACTAAT (Forward)	19
	ATGCT <b>CTAG</b> AAACCTGGGAGCCCTTCTTAAG (Reverse)	20
Termocel 5	GAAACGCTCCTCCCTGTAGT (Forward)	21
	ATGCT <b>CTAG</b> AAATTTCTCTCACCTCCAGATCAAT AGAGA (Reverse)	22
Termocel 2	AGGTGGGTAGTTCTTCTGATGG (Forward)	23
	ATGCT <b>CTAG</b> AAATTTTACAACCTTCGACGAAGAA GTCTTTGA (Reverse)	24
Termocel 7	ATAT <b>CCATGG</b> GAAAGATCGATGAAATCCTTTCA (Forward)	25
	ATGCT <b>CTAG</b> AAATGGTTTGAATCTTCTCTC CC (Reverse)	26
Termocel 8	AACGTGAAAAAGTCCCTGAAG (Forward)	27
	ATGCT <b>CTAG</b> AAATCTTCCAGACTGTTGCTTT TG (Reverse)	28

<sup>a</sup>Boldface indicates sequences complementary to the primers used to amplify the selectable markers.

## Expression and Purification

An overnight growth of transformed *E. coli* strain containing the fusion protein vector was inoculated into fresh Luria-Bertani medium containing ampicillin. When the OD<sup>600</sup> reached 0.5-0.6, L-arabinose was added to a final concentration of 0.2%. The culture was allowed to grow for another 4-5 h at 37° C. and the cells were collected by centrifugation. The pellet was stored at -80° C. prior to further processes. Cells were disrupted by sonication and the cell debris was removed by centrifugation at 10,000×g for 20 min. The protein pool was then heat treated at 95° C. for 5 min, and denatured proteins were removed by centrifugation at 12,000×g for 20

min. The recombinant protein carrying a His6 tag was then purified by immobilized metal-chelate affinity chromatography (Qiagen). Hydrolysis of cellulose, hemicellulose and starch Hydrolysis of Avicel PH101, carboxymethyl cellulose (CMC), xylan from birch wood, α-cellulose, β-glucan barley, laminarin, lichenan, starch, swollen Avicel PH101, wheat arabinoxylan, xylan from beechwood and xylan from oat-spelt was measured spectrophotometrically by the increase of reducing ends at various temperatures and pH. The amount of reducing sugar ends was determined by the dinitrosalicylic acid (DNS) method. The assay mix contained 10 μl of diluted enzymes, 30 μl of 100 mM sodium phosphate buffer, pH 6.0, and 20 μl of 0.5% (wt/vol) soluble substrates or 1% slurries (wt/vol) of insoluble substrates for 30 min or 1 hour. The reaction was terminated by adding 60 μl of DNS Solution. The absorbance of assay mix was read at 575 nm after the incubation at 100° C. for 5 min. The activity of enzymes as a function of temperature and pH was measured with CMC. Temperature gradient was achieved using PCR cycler (MJ Research). Phosphate/citrate buffers were used to generate pH gradient (ie., 2, 3, 4, 5, 6, 7, 8, 9.1).

For the thermostability assay, each enzyme was incubated at 90° C. An aliquot of enzymes was taken each day. Residual activity was measured with CMC.

## Hydrolysis of p-nitrophenol-β-D-glucoside

Activity of β-glucosidase was determined spectrophotometrically by monitoring the release of p-nitrophenol from the substrate p-nitrophenol-β-D-glucoside (Sigma) at various temperatures and pH. The assay mix contained 10 μl of diluted enzymes, 30 μl of 100 mM pH buffer, and 20 μl of 50 mM p-nitrophenol-β-D-glucoside for 10 min. The reaction was terminated by adding 120 μl of 1M Na<sub>2</sub>CO<sub>3</sub>. The absorbance of assay mix was read at 412 nm. Temperature and pH dependent activities and thermostability were measured as described above except that p-nitrophenol-β-D-glucoside was used as substrate.

## Capillary Electrophoresis of Oligosaccharides

Capillary electrophoresis of oligosaccharides was performed on a BioFocus 2000 (Bio-Rad Laboratories,) with laser-induced fluorescence detection. A fused-silica capillary (TSPO50375, Polymicro Technologies) of internal diameter 50 μm and length 31 cm was used as the separation column for oligosaccharides. The samples were injected by application of 4.5 lbin-2 of helium pressure for 0.22 sec. Electrophoresis conditions were 15 kV/70-100 μA with the cathode at the inlet, 0.1 M sodium phosphate, pH 2.5, as running buffer, and a controlled temperature of 20° C. The capillary was rinsed with 1 M NaOH followed by running buffer with adip-cycle to prevent carryover after injection. Oligomers labeled with APTS were excited at 488 nm and emission was collected through a 520-nm band pass filter.

## Biomass Substrate Specificity of Termocels

A table depicting the biomass substrate specificity of Termocels 1-8 is provided as FIG. 15.

While the invention has been described in terms of its preferred embodiments, those skilled in the art will recognize that the invention can be practiced with modification within the spirit and scope of the appended claims. Accordingly, the present invention should not be limited to the embodiments as described above, but should further include all modifications and equivalents thereof within the spirit and scope of the description provided herein.

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<160> NUMBER OF SEQ ID NOS: 32

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<211> LENGTH: 903

<212> TYPE: DNA

<213> ORGANISM: *Pyrococcus furiosus*

<400> SEQUENCE: 1

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<212> TYPE: PRT

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<400> SEQUENCE: 2

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 35             40             45
Ile Asp Lys Asp Gly Asp Gly Asn Pro Glu Phe Tyr Ile Glu Ile Asn
 50             55             60
Leu Trp Asn Ile Leu Asn Ala Thr Gly Phe Ala Glu Met Thr Tyr Asn
 65             70             75             80
Leu Thr Ser Gly Val Leu His Tyr Val Gln Gln Leu Asp Asn Ile Val
 85             90             95
Leu Arg Asp Arg Ser Asn Trp Val His Gly Tyr Pro Glu Ile Phe Tyr
 100            105            110
Gly Asn Lys Pro Trp Asn Ala Asn Tyr Ala Thr Asp Gly Pro Ile Pro
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Leu Pro Ser Lys Val Ser Asn Leu Thr Asp Phe Tyr Leu Thr Ile Ser
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 195 200 205  
 Pro Val Asn Ala Thr Phe Glu Val Trp Lys Ala Asn Ile Gly Trp Glu  
 210 215 220  
 Tyr Val Ala Phe Arg Ile Lys Thr Pro Ile Lys Glu Gly Thr Val Thr  
 225 230 235 240  
 Ile Pro Tyr Gly Ala Phe Ile Ser Val Ala Ala Asn Ile Ser Ser Leu  
 245 250 255  
 Pro Asn Tyr Thr Glu Leu Tyr Leu Glu Asp Val Glu Ile Gly Thr Glu  
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 tatttctgcg tctgggagat cgggacggaa tttggagatc caaacacaac aacggcaaaa 780  
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 <211> LENGTH: 274  
 <212> TYPE: PRT  
 <213> ORGANISM: Thermotoga petrophila

<400> SEQUENCE: 4

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Tyr	Ala	Asp	Leu	Tyr	Asn	Ile	Val	Leu	Gln	Asn	Pro	Asp	Ser	Trp	Val
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His	Gly	Tyr	Pro	Glu	Ile	Tyr	Tyr	Gly	Tyr	Lys	Pro	Trp	Ala	Ser	His
				85					90					95	
Asn	Ser	Gly	Val	Glu	Phe	Leu	Pro	Val	Lys	Val	Lys	Asp	Leu	Pro	Asp
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Ile	Asn	Leu	Ala	Met	Glu	Thr	Trp	Ile	Thr	Lys	Ser	Pro	Asp	Gln	Thr
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&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Pyrococcus horikoshii

&lt;400&gt; SEQUENCE: 5

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&lt;400&gt; SEQUENCE: 6

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Tyr Gln Thr Pro Thr Gly Ile Tyr Tyr Glu Val Arg Gly Asp Thr Ile
35          40          45
Tyr Met Ile Asn Val Thr Ser Gly Glu Glu Thr Pro Ile His Leu Phe
50          55          60
Gly Val Asn Trp Phe Gly Phe Glu Thr Pro Asn His Val Val His Gly
65          70          75          80
Leu Trp Lys Arg Asn Trp Glu Asp Met Leu Leu Gln Ile Lys Ser Leu
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Gly Thr Gln Pro Ile Gly Ile Asp Tyr Ser Lys Asn Pro Asp Leu Arg
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Asp Leu Gly Ile Phe Val Leu Leu Asp Tyr His Arg Ile Gly Cys Thr
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His Ile Glu Pro Leu Trp Tyr Thr Glu Asp Phe Ser Glu Glu Asp Phe
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Ile Asn Thr Trp Ile Glu Val Ala Lys Arg Phe Gly Lys Tyr Trp Asn
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Val Ile Gly Ala Asp Leu Lys Asn Glu Pro His Ser Val Thr Ser Pro
195         200         205
Pro Ala Ala Tyr Thr Asp Gly Thr Gly Ala Thr Trp Gly Met Gly Asn
210         215         220
Pro Ala Thr Asp Trp Asn Leu Ala Ala Glu Arg Ile Gly Lys Ala Ile
225         230         235         240
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 Asn Lys Phe Cys Asp Phe Phe Tyr Trp Ser Trp Asn Pro Asp Ser Gly  
       370                                  375                                  380  
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 Lys Tyr Asn Asn Leu Lys Arg Leu Met Asp Ser Cys Ser Lys Ser Ser  
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&lt;212&gt; TYPE: DNA

<213> ORGANISM: *Pyrococcus abyssi*

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gatgttaagc gcttgggttt taatgctatc aggcttccct tctgctctga aagcatccgc      300
cctgatacgc gcccttcgcc tgagcggata aactacgagt tgaaccccga cttgaagaat      360
ctgacttccc tcgaaataat ggagaagatt attgaatacg ccaactcaat cgggctctac      420
atactcttgg attatcacgc catcggttgt gaggagatcg aacctctttg gtataccgag      480
aattactcag aggagcagta tataaaggat tggatcttcc tcgcaaagcg gttcgggaag      540
taccctaacg tgataggagc tgatatcaag aacgagccgc atggtgaagc cgggtggggg      600
acgggagatg agcgggattt ccgcctcttt gccgagaagg tcgggcgcga gatactcaag      660
gtggcccccac actggttgat attcgtcgag ggaacgcaat ataccatgt cccgaatatt      720
gatgagatca tcgagaagaa gggctggtgg acattttggg gagagaatct tatgggagtt      780
aaggactatc cagtcaggct tccgcggcgc aaggtcgtgt actcaccgca tgtctatgga      840
ccatctgtct acatgatgga ctacttcaag tcgccagact ttccgaacaa tatgccgata      900
atctgggaaa cacacttcgg atacttgacc gacctgaatt ataccttggg cataggcgag      960
tggggtgcca actatgaggg ccttgacaag gtgtggcaag acgcttctgt gaagtggctg     1020

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attaagaaga agatctataa cttcttctac tggtgctga acccggagtc gggtagacacc 1080
ggtggcatct ttctcgacga ctggaaaacc gttaactggg aaaagatgag ggttatttac 1140
aggctcatca aggcggcgaa ccccgagttt gaggaacccc tttacatcat tttgaaaact 1200
aacgcgacga catctatcct gggcgtgggt gagaggatcc ggatttactg gtacacaaat 1260
ggcaaagtta ttgactctaa cttcgcgcat tccagcgaag gcgaaatgaa cattacagt 1320
acgaagtcca tgactctgta catcatcgtg aagaagggca atcagacact gaggaaggaa 1380
ctcaaaactgt acgttatcgg cggcaattac ggctccaata tctccactac ccagctgggt 1440
actccaaga aaggcggcga aaggattagc accagcctga agctggcaat tagcctgctc 1500
ttcattctcc tcttcgtttg gtatctcttc cgggagaagc at 1542

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<210> SEQ ID NO 8
<211> LENGTH: 514
<212> TYPE: PRT
<213> ORGANISM: Pyrococcus abyssi

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<400> SEQUENCE: 8

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Met Glu Ile Lys Leu Phe Cys Val Phe Ile Val Phe Ile Ile Leu Phe
1           5           10          15
Ser Pro Phe Val Ile Ala Leu Ser Tyr Pro Asp Val Asn Tyr Thr Ala
20          25          30
Glu Asn Gly Ile Ile Phe Val Gln Asn Val Thr Thr Gly Glu Lys Lys
35          40          45
Pro Leu Tyr Leu His Gly Val Ser Trp Phe Gly Phe Glu Leu Lys Asp
50          55          60
His Val Val Tyr Gly Leu Asp Lys Arg Asn Trp Lys Asp Ile Leu Lys
65          70          75          80
Asp Val Lys Arg Leu Gly Phe Asn Ala Ile Arg Leu Pro Phe Cys Ser
85          90          95
Glu Ser Ile Arg Pro Asp Thr Arg Pro Ser Pro Glu Arg Ile Asn Tyr
100         105         110
Glu Leu Asn Pro Asp Leu Lys Asn Leu Thr Ser Leu Glu Ile Met Glu
115         120         125
Lys Ile Ile Glu Tyr Ala Asn Ser Ile Gly Leu Tyr Ile Leu Leu Asp
130         135         140
Tyr His Arg Ile Gly Cys Glu Glu Ile Glu Pro Leu Trp Tyr Thr Glu
145         150         155         160
Asn Tyr Ser Glu Glu Gln Tyr Ile Lys Asp Trp Ile Phe Leu Ala Lys
165         170         175
Arg Phe Gly Lys Tyr Pro Asn Val Ile Gly Ala Asp Ile Lys Asn Glu
180         185         190
Pro His Gly Glu Ala Gly Trp Gly Thr Gly Asp Glu Arg Asp Phe Arg
195         200         205
Leu Phe Ala Glu Lys Val Gly Arg Glu Ile Leu Lys Val Ala Pro His
210         215         220
Trp Leu Ile Phe Val Glu Gly Thr Gln Tyr Thr His Val Pro Asn Ile
225         230         235         240
Asp Glu Ile Ile Glu Lys Lys Gly Trp Trp Thr Phe Trp Gly Glu Asn
245         250         255
Leu Met Gly Val Lys Asp Tyr Pro Val Arg Leu Pro Arg Gly Lys Val
260         265         270
Val Tyr Ser Pro His Val Tyr Gly Pro Ser Val Tyr Met Met Asp Tyr
275         280         285

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Phe Lys Ser Pro Asp Phe Pro Asn Asn Met Pro Ile Ile Trp Glu Thr  
 290 295 300  
 His Phe Gly Tyr Leu Thr Asp Leu Asn Tyr Thr Leu Val Ile Gly Glu  
 305 310 315 320  
 Trp Gly Gly Asn Tyr Glu Gly Leu Asp Lys Val Trp Gln Asp Ala Phe  
 325 330 335  
 Val Lys Trp Leu Ile Lys Lys Lys Ile Tyr Asn Phe Phe Tyr Trp Cys  
 340 345 350  
 Leu Asn Pro Glu Ser Gly Asp Thr Gly Gly Ile Phe Leu Asp Asp Trp  
 355 360 365  
 Lys Thr Val Asn Trp Glu Lys Met Arg Val Ile Tyr Arg Leu Ile Lys  
 370 375 380  
 Ala Ala Asn Pro Glu Phe Glu Glu Pro Leu Tyr Ile Ile Leu Lys Thr  
 385 390 395 400  
 Asn Ala Thr Thr Ser Ile Leu Gly Val Gly Glu Arg Ile Arg Ile Tyr  
 405 410 415  
 Trp Tyr Thr Asn Gly Lys Val Ile Asp Ser Asn Phe Ala His Ser Ser  
 420 425 430  
 Glu Gly Glu Met Asn Ile Thr Val Thr Lys Ser Met Thr Leu Tyr Ile  
 435 440 445  
 Ile Val Lys Lys Gly Asn Gln Thr Leu Arg Lys Glu Leu Lys Leu Tyr  
 450 455 460  
 Val Ile Gly Gly Asn Tyr Gly Ser Asn Ile Ser Thr Thr Gln Leu Val  
 465 470 475 480  
 Thr Pro Lys Lys Gly Gly Glu Arg Ile Ser Thr Ser Leu Lys Leu Ala  
 485 490 495  
 Ile Ser Leu Leu Phe Ile Leu Leu Phe Val Trp Tyr Leu Leu Arg Glu  
 500 505 510  
 Lys His

&lt;210&gt; SEQ ID NO 9

&lt;211&gt; LENGTH: 987

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Thermotoga petrophila

&lt;400&gt; SEQUENCE: 9

atgaaaacgc tcctccctgt agtcgtggtc cacgatattg agccagtttc aatgcgtcct 60  
 cagaggtaca agaacaaaaa ttcgataaaa agagaaaagc agggattaat acccctgttt 120  
 ttttatTTTT gggtgatatt agttctatTT gcgaattttc agattttgaa tgtaaacatt 180  
 ttcataataa gatgttttct ggaggtgata atggtgggtac tgatgacaaa accgggaaca 240  
 tcggattttg tatggaatgg cattccccct tccatggagc tgaatctgtg gaacataaag 300  
 gaatactccg gttctgtagc tatgaaattc gacggtgaaa aggtaacttt cgacgcggac 360  
 attcagaatc tttctccaaa agaaccagaa aggtacgttc tcggttatcc cgagttctat 420  
 tacggttata aaccctggga aaagcacacg gcagaaggtt cgaaacttcc agtacctggt 480  
 tcctctatga aatcattttc cgtcgaagtt tctttcgata ttcaccacga accgtctctg 540  
 cctttgaact ttgccatgga aacatgggtc acaagagaaa agtaccagac ggaagcgtcg 600  
 atcggcgatg ttgaaatcat ggtctgggtc tatttcaaca atctcacacc agggggcaaa 660  
 aagatagagg agtttacgat tccgttcgtg ctgaacggag agagtgtcga aggcacctgg 720  
 gaactgtggc acgcggagtg gggatgggac tacctcgttt tccgcttgaa ggatcccggtg 780

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aagaaggaa gggatgaatt cgacgtgagg cattttcttg atgccgccgg gaaagctctt 840
tcgaattcca ctctgttgaa agattttgaa aatctttact tcaccgtctg gaaattgga 900
accgagtttg gaagcccga aacaaagagc gcgcaattcg ggtggaagtt tgaaaacttc 960
tctattgatc tggaggtgag agaatga 987

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&lt;210&gt; SEQ ID NO 10

&lt;211&gt; LENGTH: 328

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Thermotoga petrophila

&lt;400&gt; SEQUENCE: 10

```

Met Glu Thr Leu Leu Pro Val Val Val Val His Asp Ile Glu Pro Val
1          5          10          15
Ser Met Arg Leu Gln Arg Tyr Lys Asn Lys Asn Ser Ile Lys Arg Glu
20          25          30
Lys Gln Gly Leu Ile Pro Leu Phe Phe Tyr Phe Trp Val Tyr Leu Val
35          40          45
Leu Phe Ala Asn Phe Gln Ile Leu Asn Val Asn Ile Phe Ile Ile Arg
50          55          60
Cys Phe Leu Glu Val Ile Met Val Val Leu Met Thr Lys Pro Gly Thr
65          70          75          80
Ser Asp Phe Val Trp Asn Gly Ile Pro Leu Ser Met Glu Leu Asn Leu
85          90          95
Trp Asn Ile Lys Glu Tyr Ser Gly Ser Val Ala Met Lys Phe Asp Gly
100         105         110
Glu Lys Val Thr Phe Asp Ala Asp Ile Gln Asn Leu Ser Pro Lys Glu
115         120         125
Pro Glu Arg Tyr Val Leu Gly Tyr Pro Glu Phe Tyr Tyr Gly Tyr Lys
130         135         140
Pro Trp Glu Lys His Thr Ala Glu Gly Ser Lys Leu Pro Val Pro Val
145         150         155         160
Ser Ser Met Lys Ser Phe Ser Val Glu Val Ser Phe Asp Ile His His
165         170         175
Glu Pro Ser Leu Pro Leu Asn Phe Ala Met Glu Thr Trp Leu Thr Arg
180         185         190
Glu Lys Tyr Gln Thr Glu Ala Ser Ile Gly Asp Val Glu Ile Met Val
195         200         205
Trp Phe Tyr Phe Asn Asn Leu Thr Pro Gly Gly Lys Lys Ile Glu Glu
210         215         220
Phe Thr Ile Pro Phe Val Leu Asn Gly Glu Ser Val Glu Gly Thr Trp
225         230         235         240
Glu Leu Trp His Ala Glu Trp Gly Trp Asp Tyr Leu Ala Phe Arg Leu
245         250         255
Lys Asp Pro Val Lys Lys Gly Arg Val Lys Phe Asp Val Arg His Phe
260         265         270
Leu Asp Ala Ala Gly Lys Ala Leu Ser Asn Ser Thr Arg Val Lys Asp
275         280         285
Phe Glu Asn Leu Tyr Phe Thr Val Trp Glu Ile Gly Thr Glu Phe Gly
290         295         300
Ser Pro Glu Thr Lys Ser Ala Gln Phe Gly Trp Lys Phe Glu Asn Phe
305         310         315         320
Ser Ile Asp Leu Glu Val Arg Glu
325

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<210> SEQ ID NO 11
<211> LENGTH: 852
<212> TYPE: DNA
<213> ORGANISM: Caldivirga maquilingensis

<400> SEQUENCE: 11
atgttgaaac ttattccact tgtaaatggc aattataagt tgattcaatg ggagccactc   60
ggcggcgtgc acggagcaga tatcgagtgc atacatgta ccccaaacgt atggaacata   120
gataaatcat cagttggcac tgtacagatc gaatatgagc cccaagttgg ctgtcttcgt   180
ttttcaattg atttcccag gataagtata agacataatg taggcgtagc ggcatattca   240
gaagtatttt acggacacaa gccgtggggc cccaccactt gcatggacce tcagttcaag   300
ttccctatca aagtcaatga gtcaaaagga ctgtactcgt atgtaaatta taacgttaaa   360
tctaggtcac cagatgactc aatctttaat attgcttacg atctctggct tacaacgtcc   420
ccaaacctta caaacggacc ccagccagga gacgtagaag ttatgatctg gttgtactac   480
cacggacagc gccctgcagg cagactcadc ggggaactcc gcatgccgat tacattgggc   540
gatagtgagg cggcacgtga ctttgaagta tgggtggctg acacaggaat aggaatcggt   600
gaatgggcgg tagtgacctt cagaatcaag gacccaataa agggcggttt gataggagtt   660
aacctcataa actacatoga aagtgctttt aaaacgctcg aagaactcaa cccggtaag   720
tggcggtacg gcgacctgct caacaaatat cttaatggaa ttgaattcgg cagtgagttt   780
ggtaatgtct cctcaggaat gataaaactt aattgggaac tctgcccct gagccttgtg   840
aaagactctt ct   852

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<210> SEQ ID NO 12
<211> LENGTH: 284
<212> TYPE: PRT
<213> ORGANISM: Caldivirga maquilingensis

<400> SEQUENCE: 12
Met Leu Lys Leu Ile Pro Leu Val Asn Gly Asn Tyr Lys Leu Ile Gln
 1           5           10          15
Trp Glu Pro Leu Gly Gly Val His Gly Ala Asp Ile Glu Cys Ile His
 20          25          30
Val Thr Pro Asn Val Trp Asn Ile Asp Lys Ser Ser Val Gly Thr Val
 35          40          45
Gln Ile Glu Tyr Glu Pro Gln Val Gly Cys Leu Arg Phe Ser Ile Asp
 50          55          60
Phe Pro Arg Ile Ser Ile Arg His Asn Val Gly Val Ala Ala Tyr Ser
 65          70          75          80
Glu Val Ile Tyr Gly His Lys Pro Trp Gly Pro Thr Thr Cys Met Asp
 85          90          95
Pro Gln Phe Lys Phe Pro Ile Lys Val Asn Glu Ser Lys Gly Leu Tyr
 100         105         110
Ser Tyr Val Asn Tyr Asn Val Lys Ser Arg Ser Pro Asp Asp Ser Ile
 115        120        125
Phe Asn Ile Ala Tyr Asp Leu Trp Leu Thr Thr Ser Pro Asn Leu Thr
 130        135        140
Asn Gly Pro Gln Pro Gly Asp Val Glu Val Met Ile Trp Leu Tyr Tyr
 145        150        155        160
His Gly Gln Arg Pro Ala Gly Arg Leu Ile Gly Glu Leu Arg Met Pro
 165        170        175

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Ile	Thr	Leu	Gly	Asp	Ser	Glu	Ala	Ala	Arg	Asp	Phe	Glu	Val	Trp	Val
			180					185					190		
Ala	Asp	Thr	Gly	Ile	Gly	Ile	Gly	Glu	Trp	Ala	Val	Val	Thr	Phe	Arg
		195					200					205			
Ile	Lys	Asp	Pro	Ile	Lys	Gly	Gly	Leu	Ile	Gly	Val	Asn	Leu	Ile	Asn
	210					215					220				
Tyr	Ile	Glu	Ser	Ala	Phe	Lys	Thr	Leu	Glu	Glu	Leu	Asn	Pro	Val	Lys
225					230					235					240
Trp	Arg	Tyr	Gly	Asp	Leu	Leu	Asn	Lys	Tyr	Leu	Asn	Gly	Ile	Glu	Phe
				245					250						255
Gly	Ser	Glu	Phe	Gly	Asn	Val	Ser	Ser	Gly	Met	Ile	Lys	Leu	Asn	Trp
			260					265						270	
Glu	Leu	Cys	Gly	Leu	Ser	Leu	Val	Lys	Asp	Ser	Ser				
		275					280								

&lt;210&gt; SEQ ID NO 13

&lt;211&gt; LENGTH: 2166

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Thermotoga petrophila

&lt;400&gt; SEQUENCE: 13

atgatgggaa agatcgatga aatcctttca cagctgacta ttgaagaaaa agtgaactt	60
gtagtggggg ttggtcttcc aggaactttt ggaaatccac attccagagt ggcaggtgca	120
gctggagaaa cgcacacctgt tccgaggctt ggaattcctt ctttcgttct ggccgacggt	180
cccgcgggcc tcagaataaa tcccacaaga gagaacgacg aaaacaccta ttacacaaca	240
gcgtttctct ttgaaatcat gctcgtctcc acctggaaca aagatcttct ggaagaagta	300
ggaaaagcta tgggagaaga agtcaggaa tacggtgtcg atgtgcttct tgcacctgcg	360
atgaacattc acaggaaccc tctttgtgga aggaatttcg agtattattc agaagatcct	420
gtcctttccg gtgaaatggc ttcagccttt gtcaaggagg ttcaatctca aggggtggga	480
gcctgcataa aacactttgt cgcgaacaac caggaaacga acaggatggt agtggacacg	540
atcgtgtccg agcagaccct cagagaaata tatctgaaag gttttgaaat tgcctcaag	600
aaagcaagac cctggaccgt gatgagcgtc tacaacaac tgaatggaaa atactgttca	660
cagaacgaat ggcttttgaa gaaggttctc agggaagaat ggggatttga cggtttctgt	720
atgagcgact ggtacgctgg agacaaccct gtagaacacg tcaaggccgg aaacgatatg	780
atcatgcctg gaaaaagcgtc tcaggtgaac acggaagaa gagatgaaat agaagaaatc	840
atggaggcgt tgaaggaggg aagactcagt gaggaagtcc tgaacgaatg tgtgagaaac	900
atcctcaaag ttcttgtgaa cgcgccttcc tttaaagggt acaggctacc gaacaaaccg	960
gacctcgaat ctcacgcgaa agttgcctac gaagcagggt tggagggtgt tgtccttctt	1020
gagaacaacg gtgttcttcc attcgatgaa agtatccatg tcgccgtctt tggcaccggt	1080
caaatcgaac caataaaggg aggaacggga agtggagaca cccatccgag atacacgatc	1140
tctatccttg aaggcataaa agaaagaaac atgaagttcg acgaagaact cacctccatc	1200
tatgaggatt acatcaaaaa gatgagagaa acagaggaat ataaaccag aactgactcc	1260
tggggaacgg ttataaaacc gaaacttcca gagaacttcc tctcagaaaa agagataaag	1320
aaggctgcga agaaaaacga tgctgcagtt gttgtaatca gtaggatctc cggtagggga	1380
tacgacagaa agccgggtgaa aggtgacttc acctctccga tgacgagctg gagctcataa	1440
aaacagtctc aagggaattc cacgaacagg gtaagaaggt tgtggttctt ctcaacatcg	1500

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gaagtcccat tgaagttgca agctggagag atcttggtgga tggaaatcctt ctcgtctggc 1560
aagcaggaca ggagatggga agaatagtgg ccgatgttct tgtgggaagg gtaaaccctt 1620
cgggaaaact tccaacgacc tteccgaagg attactcgga cgttccatcc tggacgttcc 1680
caggagagcc aaaggacaat ccgcaaagag tgggtgtacga ggaagacatc tacgtgggat 1740
acaggtacta cgacaccttt ggtgtggaac ctgcctacga gttcggctac ggctctctt 1800
acacaaagtt tgaatacaaa gatttaaaga tcgctatcga cggagatata ctcagagtgt 1860
cgtacacgat cacaaacacc ggggacagag ctggaagga agtctcacag gtttatgtca 1920
aagctccaaa agggaaaata gacaaaccct tccaggagct gaaagcgttc cacaaaaaa 1980
aacttttgaa cccgggtgaa tccgaaaaga tctttctgga aattctctt agagatcttg 2040
cgagtttcga tgggaaagaa tggttgtcga gtcaggagaa tacgaggtca gggtcgggtc 2100
atcttcgagg gatataggtt gagagatatt tttctggtg agggagagaa gagattcaaa 2160
ccatga 2166

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&lt;210&gt; SEQ ID NO 14

&lt;211&gt; LENGTH: 722

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Thermotoga petrophila

&lt;400&gt; SEQUENCE: 14

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Met Met Gly Lys Ile Asp Glu Ile Leu Ser Gln Leu Thr Ile Glu Glu
1           5           10          15
Lys Val Lys Leu Val Val Gly Val Gly Leu Pro Gly Leu Phe Gly Asn
20          25          30
Pro His Ser Arg Val Ala Gly Ala Ala Gly Glu Thr His Pro Val Pro
35          40          45
Arg Leu Gly Ile Pro Ser Phe Val Leu Ala Asp Gly Pro Ala Gly Leu
50          55          60
Arg Ile Asn Pro Thr Arg Glu Asn Asp Glu Asn Thr Tyr Tyr Thr Thr
65          70          75          80
Ala Phe Pro Val Glu Ile Met Leu Ala Ser Thr Trp Asn Lys Asp Leu
85          90          95
Leu Glu Glu Val Gly Lys Ala Met Gly Glu Glu Val Arg Glu Tyr Gly
100         105        110
Val Asp Val Leu Leu Ala Pro Ala Met Asn Ile His Arg Asn Pro Leu
115        120        125
Cys Gly Arg Asn Phe Glu Tyr Tyr Ser Glu Asp Pro Val Leu Ser Gly
130        135        140
Glu Met Ala Ser Ala Phe Val Lys Gly Val Gln Ser Gln Gly Val Gly
145        150        155        160
Ala Cys Ile Lys His Phe Val Ala Asn Asn Gln Glu Thr Asn Arg Met
165        170        175
Val Val Asp Thr Ile Val Ser Glu Arg Ala Leu Arg Glu Ile Tyr Leu
180        185        190
Lys Gly Phe Glu Ile Ala Val Lys Lys Ala Arg Pro Trp Thr Val Met
195        200        205
Ser Ala Tyr Asn Lys Leu Asn Gly Lys Tyr Cys Ser Gln Asn Glu Trp
210        215        220
Leu Leu Lys Lys Val Leu Arg Glu Glu Trp Gly Phe Asp Gly Phe Val
225        230        235        240
Met Ser Asp Trp Tyr Ala Gly Asp Asn Pro Val Glu Gln Leu Lys Ala
245        250        255

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Ser Ile Trp His Thr Phe Ser His Thr Pro Gly Asn Val Lys Asn Gly  
           35                                  40                                  45  
 Asp Thr Gly Asp Val Ala Cys Asp His Tyr Asn Arg Trp Lys Glu Asp  
   50                                  55                                  60  
 Ile Glu Ile Ile Glu Lys Leu Gly Val Lys Ala Tyr Arg Phe Ser Ile  
   65                                  70                                  75                                  80  
 Ser Trp Pro Arg Ile Leu Pro Glu Gly Thr Gly Arg Val Asn Gln Lys  
                                   85                                  90                                  95  
 Gly Leu Asp Phe Tyr Asn Arg Ile Ile Asp Thr Leu Leu Glu Lys Gly  
                                   100                                  105                                  110  
 Ile Thr Pro Phe Val Thr Ile Tyr His Trp Asp Leu Pro Phe Ala Leu  
                                   115                                  120                                  125  
 Gln Leu Lys Gly Gly Trp Ala Asn Arg Glu Ile Ala Asp Trp Phe Ala  
   130                                  135                                  140  
 Glu Tyr Ser Arg Val Leu Phe Glu Asn Phe Gly Asp Arg Val Lys Asn  
   145                                  150                                  155                                  160  
 Trp Ile Thr Leu Asn Glu Pro Trp Val Val Ala Ile Val Gly His Leu  
                                   165                                  170                                  175  
 Tyr Gly Val His Ala Pro Gly Met Arg Asp Ile Tyr Val Ala Phe Arg  
                                   180                                  185                                  190  
 Ala Val His Asn Leu Leu Arg Ala His Ala Lys Ala Val Lys Val Phe  
                                   195                                  200                                  205  
 Arg Glu Thr Val Lys Asp Gly Lys Ile Gly Ile Val Phe Asn Asn Gly  
   210                                  215                                  220  
 Tyr Phe Glu Pro Ala Ser Glu Lys Glu Glu Asp Ile Arg Ala Ala Arg  
   225                                  230                                  235                                  240  
 Phe Met His Gln Phe Asn Asn Tyr Pro Leu Phe Leu Asn Pro Ile Tyr  
                                   245                                  250                                  255  
 Arg Gly Asp Tyr Pro Glu Leu Val Leu Glu Phe Ala Arg Glu Tyr Leu  
                                   260                                  265                                  270  
 Pro Glu Asn Tyr Lys Asp Asp Met Ser Glu Ile Gln Glu Lys Ile Asp  
                                   275                                  280                                  285  
 Phe Val Gly Leu Asn Tyr Tyr Ser Gly His Leu Val Lys Phe Asp Pro  
   290                                  295                                  300  
 Asp Ala Pro Ala Lys Val Ser Phe Val Glu Arg Asp Leu Pro Lys Thr  
   305                                  310                                  315                                  320  
 Ala Met Gly Trp Glu Ile Val Pro Glu Gly Ile Tyr Trp Ile Leu Lys  
                                   325                                  330                                  335  
 Lys Val Lys Glu Glu Tyr Asn Pro Pro Glu Val Tyr Ile Thr Glu Asn  
                                   340                                  345                                  350  
 Gly Ala Ala Phe Asp Asp Val Val Ser Glu Asp Gly Arg Val His Asp  
                                   355                                  360                                  365  
 Gln Asn Arg Ile Asp Tyr Leu Lys Ala His Ile Gly Gln Ala Trp Lys  
   370                                  375                                  380  
 Ala Ile Gln Glu Gly Val Pro Leu Lys Gly Tyr Phe Val Trp Ser Leu  
   385                                  390                                  395                                  400  
 Leu Asp Asn Phe Glu Trp Ala Glu Gly Tyr Ser Lys Arg Phe Gly Ile  
                                   405                                  410                                  415  
 Val Tyr Val Asp Tyr Ser Thr Gln Lys Arg Ile Ile Lys Asp Ser Gly  
                                   420                                  425                                  430  
 Tyr Trp Tyr Ser Asn Val Val Lys Ser Asn Ser Leu Glu Asp  
                                   435                                  440                                  445



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<210> SEQ ID NO 23  
<211> LENGTH: 22  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: synthetic oligonucleotide primer

<400> SEQUENCE: 23

aggtgggtag ttcttctgat gg 22

<210> SEQ ID NO 24  
<211> LENGTH: 41  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: synthetic oligonucleotide primer

<400> SEQUENCE: 24

atgctctaga aattttataa cttcgacgaa gaagtctttg a 41

<210> SEQ ID NO 25  
<211> LENGTH: 33  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: synthetic oligonucleotide primer

<400> SEQUENCE: 25

atatccatgg gaaagatcga tgaatcctt tca 33

<210> SEQ ID NO 26  
<211> LENGTH: 34  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: synthetic oligonucleotide primer

<400> SEQUENCE: 26

atgctctaga aatggtttga atctcttctc tccc 34

<210> SEQ ID NO 27  
<211> LENGTH: 22  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: synthetic oligonucleotide primer

<400> SEQUENCE: 27

aacgtgaaaa agttccctga ag 22

<210> SEQ ID NO 28  
<211> LENGTH: 34  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: synthetic oligonucleotide primer

<400> SEQUENCE: 28

atgctctaga aaatcttoca gactgttgct ttg 34

<210> SEQ ID NO 29  
<211> LENGTH: 1230  
<212> TYPE: DNA  
<213> ORGANISM: Caldivirga maquilingensis

<400> SEQUENCE: 29

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atggactact ctatcaactg ctctatcaac cctataaccc tcatggctgc gcactcttct    60
cccctgaacc catctaacac actogaactt acacttattc tcgaaaatgg catcaccacc    120
acagtaactg tcaccgcgac accacgcaac acttaccccta tgatctccct tggtacatt    180
aatattacc ctaacctctg gaaccttaac acagcttcgt catcaggata cgctctatg    240
gtctacgatg catcacaggg tgctctttat attcattgta atttcacaaa ggtttacctc    300
aatcagcaag ttggtgttgc cgctactct gaattcatct atggctacaa accctggggc    360
acgctcacct cggaggcagg cgggttcaat tttcctgtta agcttacga actcggttct    420
cttctttcgt tcatcaatta ctcaactcatt tcatattctc cacaagtgcg tatcttcgat    480
tgggcatacg acctttggct cacaacatcc ccaaattctc ccaacggccc tcaaccgggc    540
gacgtcgagg tcatgatctg gctctattat cacctgcaac aacctgctgg ttttcccgtc    600
gtaacgtta cagtgcctaat atgggtcaat ggctccctcg ttaacgaaac atttgaggtt    660
tggattgggt ctccacagat cgaaccgggc acccacgcta tagtctcctt caggccaacg    720
aatccaatcc ctgaggcct cgtcggcgta aatgtcacga agttccttca acttgccggt    780
aactatctcg tgacactcta cccctcatac tggaaactaca catatctgga gagcaagtac    840
ttgaatggca tcgaattcgg atcagaatgg ggcaatccgt ctacatacaa tattacactc    900
aattgggtca tttataaagc ttatcttctc aaggtgcctc tggagtcaca gggcacccgtt    960
accgtcacat atactacaac tgttacatcc accatgactg ttacctcaat ccttgetacc   1020
acateccacg tcaccactac atctacactt acatctacog ttaccgcccac ttcagtttct   1080
acttccacog tcacgcagac tctcaactacc tccatogtca aaaccgtcat cctgtctac   1140
tatactgcca ccataatcgt ccttcttata atcatcgcag tcgtcattgc acttgcgttc   1200
gcccgcgcg gcacccgggt tcgtctctgt                                     1230

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&lt;210&gt; SEQ ID NO 30

&lt;211&gt; LENGTH: 410

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Caldivirga maquilingensis

&lt;400&gt; SEQUENCE: 30

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Met Asp Tyr Ser Ile Asn Cys Ser Ile Asn Pro Ile Thr Leu Met Val
1           5           10           15
Ala His Ser Ser Pro Leu Asn Pro Ser Asn Thr Leu Glu Leu Thr Leu
20          25          30
Ile Leu Glu Asn Gly Ile Thr Thr Thr Val Thr Val Thr Ala Thr Pro
35          40          45
Arg Asn Thr Tyr Pro Met Ile Ser Leu Gly Tyr Ile Asn Ile Thr Pro
50          55          60
Asn Leu Trp Asn Leu Asn Thr Ala Ser Ser Ser Gly Tyr Ala Ser Met
65          70          75          80
Val Tyr Asp Ala Ser Gln Gly Ala Leu Tyr Ile His Val Asn Phe Thr
85          90          95
Lys Val Tyr Leu Asn Gln Gln Val Gly Val Ala Ala Tyr Ser Glu Phe
100         105         110
Ile Tyr Gly Tyr Lys Pro Trp Gly Thr Leu Thr Ser Glu Ala Gly Gly
115        120        125
Phe Asn Phe Pro Val Lys Leu Thr Glu Leu Gly Ser Leu Leu Ser Phe
130        135        140
Ile Asn Tyr Ser Leu Ile Ser Tyr Ser Pro Gln Val Ala Ile Phe Asp
145        150        155        160

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Trp Ala Tyr Asp Leu Trp Leu Thr Thr Ser Pro Asn Leu Thr Asn Gly  
                   165                                  170                                  175  
 Pro Gln Pro Gly Asp Val Glu Val Met Ile Trp Leu Tyr Tyr His Leu  
                   180                                  185                                  190  
 Gln Gln Pro Ala Gly Phe Pro Val Ala Asn Val Thr Val Pro Ile Trp  
                   195                                  200                                  205  
 Val Asn Gly Ser Leu Val Asn Glu Thr Phe Glu Val Trp Ile Gly Ser  
                   210                                  215                                  220  
 Pro Gln Ile Glu Pro Gly Thr His Ala Ile Val Ser Phe Arg Pro Thr  
                   225                                  230                                  235                                  240  
 Asn Pro Ile Pro Arg Gly Leu Val Gly Val Asn Val Thr Lys Phe Leu  
                                   245                                  250                                  255  
 Gln Leu Ala Val Asn Tyr Leu Val Thr Leu Tyr Pro Ser Tyr Trp Asn  
                                   260                                  265                                  270  
 Tyr Thr Tyr Leu Glu Ser Lys Tyr Leu Asn Gly Ile Glu Phe Gly Ser  
                                   275                                  280                                  285  
 Glu Trp Gly Asn Pro Ser Thr Tyr Asn Ile Thr Leu Asn Trp Val Ile  
                                   290                                  295                                  300  
 Tyr Lys Ala Tyr Leu Ile Lys Val Pro Leu Glu Ser Gln Gly Thr Val  
                                   305                                  310                                  315                                  320  
 Thr Val Thr Tyr Thr Thr Thr Val Thr Ser Thr Met Thr Val Thr Ser  
                                   325                                  330                                  335  
 Ile Leu Ala Thr Thr Ser Thr Val Thr Thr Thr Ser Thr Leu Thr Ser  
                                   340                                  345                                  350  
 Thr Val Thr Ala Thr Ser Val Ser Thr Ser Thr Val Thr Gln Thr Leu  
                                   355                                  360                                  365  
 Thr Thr Ser Ile Val Lys Thr Val Ile Pro Val Tyr Tyr Thr Ala Thr  
                                   370                                  375                                  380  
 Ile Ile Val Leu Leu Ile Ile Ile Ala Val Val Ile Ala Leu Ala Phe  
                                   385                                  390                                  395                                  400  
 Ala Arg Arg Gly Ile Arg Val Arg Leu Cys  
                                   405                                  410

&lt;210&gt; SEQ ID NO 31

&lt;211&gt; LENGTH: 2214

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Pyrococcus horikoshii

&lt;400&gt; SEQUENCE: 31

```

atgagatttc aattcggatt ctccaagaa gatgaacagg tgctgggcac aactactaaca    60
ctcgaaaatg gacaattagg agttagggga gaatttgaac tcgagagatc tccttatgga    120
acgatcgtta gcggggtcta tgattacact cctacttct acaggaatt ggtaaatggt    180
cccaggacta taggatgat aataattata gatggagaac taataaatcc aagctctcaa    240
aaagtcaagg aattccagag agagctcgat atagaaaaag gcttattaag aactcactta    300
gagattgaaa caaaaaatgg aaataaaatt ttatataaaa gtacaaggat agtccacatg    360
aaaagaaaaa acctaactct tctagatttt gagctaaaag ctagcaaggg aggaatcgca    420
gttgtagtta atcccataga attcaatact gcaaatccag ggtttataga cgagataatg    480
atcaagcatt atagagtgga ctcgataaaa gagactgagg agggagtata cgctagggtg    540
aaaactttag acaataagta cacgttgaa attgcaagta gcttggttcc atcagaatat    600
acatcgagga gcacctttag aaccgataat gaaattggag aaatttcat tgtaaactt    660

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aaaccaggaa aaacgtacaa atttacaag tacgttacag tatctaaagg agcagcttta 720
gaggagttaa aagatgtaa gagattagga ttgaaaagc tatatgaaga gcatataaac 780
agctggaaga gaatatggga gaaagtgaaa gtggaaatcg aaggagataa agacctgaa 840
aatgccttaa actttaacat tttcacttg atccaatccc tccaccaac agataaagtc 900
tcgctaccag caaggggaat acatggggtt gggtataggg gacatatatt ctgggataca 960
gagatatatg cattaccttt cttcatattc acgatgcaa aagaggccag gagattgctc 1020
ctctatagat gcaacaactt agatgcccct aaagaaaatg caaagatgaa tggatatcaa 1080
ggggtccaat ttccctggga gtcggcagat gatggacgcg aggctacccc ctctgagata 1140
ccattggata tgttgggaag gaaaatcgtt agaatttaca ccggagagga ggaacatcac 1200
ataactgagg atatagcata tatagttgat ttttattacc aagtctctgg agatctcgaa 1260
tttatgaaca ggtgtggcct tgagataatc tttgagacgg cccgattttg ggctagtagg 1320
gttgagttcg aggaaggaaa agggtagctc attaaaaaag taataggacc tgatgaatac 1380
catgagcaag ttaacaacaa cttctttaca aacttaatgg ccaagcataa tctcgaactt 1440
gcaataagat acttttagaga gtcaaagaat agggaaacct ggaaaaagat tgcgaaaaa 1500
ttaaacataa gagaggagga ggttgaaaaa tgggaagaga tagctaaaaa catgtacatt 1560
cccaggaaga tagacggagt tttgaagag ttgatggtt actttgaatt gatggatttt 1620
gaagttgatc ccttcaatat tggagaaaaa acactccccg aggaaatcag gaataacata 1680
gggaaaaacg aactcgttaa gcaggccgat gtcacatggt cccaatatct ccttaaggac 1740
tacttctctc cagaggaaat aaagagtaac ttaactatt atataaggag aactacccat 1800
gcttcatcac tctccatgcc cccatagcgc atcattgcaa cctggatagg ggaggtaaag 1860
atagcatatg agtacttcaa gagatgtgca aatatagatc tcaaaaacgt gtacggaaac 1920
actgcagagg gatttcactt agcaacggcg ggaggaacct ggcaagtact cgtcagagga 1980
ttttgtggcc tcaatgtaaa aggaaacaaa atagagctta atcctaactt tcctgaaaaa 2040
tggaagtagc ttaagttcag gatattcttc aaaggttcat ggatagaatt taaaatttct 2100
aggaagaaag ttagggctag aatgctttaa ggatcgagaa aagtcaaaat atctagcttt 2160
ggaaaggaag tagatctata tcctggaaaa gaggttgtaa tagtagctaa ttaa 2214

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&lt;210&gt; SEQ ID NO 32

&lt;211&gt; LENGTH: 737

&lt;212&gt; TYPE: PRT

<213> ORGANISM: *Pyrococcus horikoshii*

&lt;400&gt; SEQUENCE: 32

```

Met Arg Phe Gln Phe Gly Phe Ser Lys Glu Asp Glu Gln Val Leu Gly
1           5           10          15
Thr Ile Leu Thr Leu Gly Asn Gly Gln Leu Gly Val Arg Gly Glu Phe
20          25          30
Glu Leu Glu Arg Ser Pro Tyr Gly Thr Ile Val Ser Gly Val Tyr Asp
35          40          45
Tyr Thr Pro Tyr Phe Tyr Arg Glu Leu Val Asn Gly Pro Arg Thr Ile
50          55          60
Gly Met Ile Ile Ile Ile Asp Gly Glu Leu Ile Asn Pro Ser Ser Gln
65          70          75          80
Lys Val Lys Glu Phe Gln Arg Glu Leu Asp Ile Glu Lys Gly Leu Leu
85          90          95
Arg Thr His Leu Glu Ile Glu Thr Lys Asn Gly Asn Lys Ile Leu Tyr

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100				105				110							
Lys	Ser	Thr	Arg	Ile	Val	His	Met	Lys	Arg	Lys	Asn	Leu	Ile	Leu	Leu
	115						120					125			
Asp	Phe	Glu	Leu	Lys	Ala	Ser	Lys	Gly	Gly	Ile	Ala	Val	Val	Val	Asn
	130					135					140				
Pro	Ile	Glu	Phe	Asn	Thr	Ala	Asn	Pro	Gly	Phe	Ile	Asp	Glu	Ile	Met
	145				150					155					160
Ile	Lys	His	Tyr	Arg	Val	Asp	Ser	Ile	Lys	Glu	Thr	Glu	Glu	Gly	Val
				165					170					175	
Tyr	Ala	Arg	Val	Lys	Thr	Leu	Asp	Asn	Lys	Tyr	Thr	Leu	Glu	Ile	Ala
			180					185						190	
Ser	Ser	Leu	Val	Pro	Ser	Glu	Tyr	Thr	Ser	Arg	Ser	Thr	Phe	Arg	Thr
		195					200					205			
Asp	Asn	Glu	Ile	Gly	Glu	Ile	Tyr	Ile	Val	Lys	Leu	Lys	Pro	Gly	Lys
	210					215					220				
Thr	Tyr	Lys	Phe	Thr	Lys	Tyr	Val	Thr	Val	Ser	Lys	Gly	Ala	Ala	Leu
	225				230					235					240
Glu	Glu	Leu	Lys	Asp	Val	Lys	Arg	Leu	Gly	Phe	Glu	Lys	Leu	Tyr	Glu
				245					250					255	
Glu	His	Ile	Asn	Ser	Trp	Lys	Arg	Ile	Trp	Glu	Lys	Val	Lys	Val	Glu
			260					265						270	
Ile	Glu	Gly	Asp	Lys	Asp	Leu	Glu	Asn	Ala	Leu	Asn	Phe	Asn	Ile	Phe
		275					280					285			
His	Leu	Ile	Gln	Ser	Leu	Pro	Pro	Thr	Asp	Lys	Val	Ser	Leu	Pro	Ala
	290					295					300				
Arg	Gly	Ile	His	Gly	Phe	Gly	Tyr	Arg	Gly	His	Ile	Phe	Trp	Asp	Thr
	305				310					315					320
Glu	Ile	Tyr	Ala	Leu	Pro	Phe	Phe	Ile	Phe	Thr	Met	Pro	Lys	Glu	Ala
			325							330				335	
Arg	Arg	Leu	Leu	Leu	Tyr	Arg	Cys	Asn	Asn	Leu	Asp	Ala	Ala	Lys	Glu
		340						345					350		
Asn	Ala	Lys	Met	Asn	Gly	Tyr	Gln	Gly	Val	Gln	Phe	Pro	Trp	Glu	Ser
		355					360					365			
Ala	Asp	Asp	Gly	Arg	Glu	Ala	Thr	Pro	Ser	Glu	Ile	Pro	Leu	Asp	Met
	370					375					380				
Leu	Gly	Arg	Lys	Ile	Val	Arg	Ile	Tyr	Thr	Gly	Glu	Glu	Glu	His	His
	385				390					395				400	
Ile	Thr	Ala	Asp	Ile	Ala	Tyr	Ile	Val	Asp	Phe	Tyr	Tyr	Gln	Val	Ser
			405						410					415	
Gly	Asp	Leu	Glu	Phe	Met	Asn	Arg	Cys	Gly	Leu	Glu	Ile	Ile	Phe	Glu
		420						425						430	
Thr	Ala	Arg	Phe	Trp	Ala	Ser	Arg	Val	Glu	Phe	Glu	Glu	Gly	Lys	Gly
		435					440					445			
Tyr	Val	Ile	Lys	Lys	Val	Ile	Gly	Pro	Asp	Glu	Tyr	His	Glu	His	Val
	450					455					460				
Asn	Asn	Asn	Phe	Phe	Thr	Asn	Leu	Met	Ala	Lys	His	Asn	Leu	Glu	Leu
	465				470					475				480	
Ala	Ile	Arg	Tyr	Phe	Arg	Glu	Ser	Lys	Asn	Arg	Glu	Pro	Trp	Lys	Lys
			485						490					495	
Ile	Val	Glu	Lys	Leu	Asn	Ile	Arg	Glu	Glu	Glu	Val	Glu	Lys	Trp	Glu
			500						505					510	
Glu	Ile	Ala	Lys	Asn	Met	Tyr	Ile	Pro	Arg	Lys	Ile	Asp	Gly	Val	Phe
		515					520					525			



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Glu Glu Phe Asp Gly Tyr Phe Glu Leu Met Asp Phe Glu Val Asp Pro  
 530 535 540  
 Phe Asn Ile Gly Glu Lys Thr Leu Pro Glu Glu Ile Arg Asn Asn Ile  
 545 550 555 560  
 Gly Lys Thr Lys Leu Val Lys Gln Ala Asp Val Ile Met Ala Gln Tyr  
 565 570 575  
 Leu Leu Lys Asp Tyr Phe Ser Pro Glu Glu Ile Lys Ser Asn Phe Asn  
 580 585 590  
 Tyr Tyr Ile Arg Arg Thr Thr His Ala Ser Ser Leu Ser Met Pro Pro  
 595 600 605  
 Tyr Ala Ile Ile Ala Thr Trp Ile Gly Glu Val Lys Ile Ala Tyr Glu  
 610 615 620  
 Tyr Phe Lys Arg Cys Ala Asn Ile Asp Leu Lys Asn Val Tyr Gly Asn  
 625 630 635 640  
 Thr Ala Glu Gly Phe His Leu Ala Thr Ala Gly Gly Thr Trp Gln Val  
 645 650 655  
 Leu Val Arg Gly Phe Cys Gly Leu Asn Val Lys Gly Asn Lys Ile Glu  
 660 665 670  
 Leu Asn Pro Asn Leu Pro Glu Lys Trp Lys Tyr Val Lys Phe Arg Ile  
 675 680 685  
 Phe Phe Lys Gly Ser Trp Ile Glu Phe Lys Ile Ser Arg Lys Lys Val  
 690 695 700  
 Arg Ala Arg Met Leu Glu Gly Ser Arg Lys Val Lys Ile Ser Ser Phe  
 705 710 715 720  
 Gly Lys Glu Val Asp Leu Tyr Pro Gly Lys Glu Val Val Ile Val Ala  
 725 730 735

Asn

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We claim:

1. A transformed host containing one or more recombinant cellulase enzymes comprising the amino acid sequence of SEQ ID NO: 2, wherein said host is corn. <sup>40</sup>

2. A transformed host comprising one or more nucleic acid sequences encoding one or more recombinant cellulase enzymes comprising the amino acid sequence of SEQ ID NO: 2, wherein at least one of said nucleic acid sequences encoding the amino acid sequence of SEQ ID NO:2 is the nucleic acid sequence of SEQ ID NO:1. <sup>45</sup>

3. A transformed host comprising one or more nucleic acid sequences encoding one or more recombinant cellulase enzymes comprising the amino acid sequence of SEQ ID NO: 2, wherein at least one of said nucleic acid sequences encoding the amino acid sequence of SEQ ID NO: 2 is the nucleic acid sequence of SEQ ID NO: 1, wherein said host is corn.

\* \* \* \* \*

UNITED STATES PATENT AND TRADEMARK OFFICE  
**CERTIFICATE OF CORRECTION**

PATENT NO. : 8,847,031 B2  
APPLICATION NO. : 13/003183  
DATED : September 30, 2014  
INVENTOR(S) : Prade et al.

Page 1 of 1

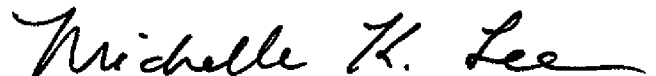
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

In the Specification

At column 1, line 3, please insert the following paragraph:

--This invention was made with U.S. Government support under DOE Grant No. DE-FG36-06GO16107 awarded by the Department of Energy. The Government has certain rights in this invention.--

Signed and Sealed this  
Thirtieth Day of December, 2014



Michelle K. Lee  
*Deputy Director of the United States Patent and Trademark Office*