

US008734808B2

(10) **Patent No.:**

(45) Date of Patent:

(12) United States Patent

Kapil

(54) ISOLATION OF A VIRUS RELATED TO CANINE PARVOVIRUS-2 FROM A RACCOON

- (75) Inventor: Sanjay Kapil, Stillwater, OK (US)
- (73) Assignee: The Board of Regents for Oklahoma State University, Stillwater, OK (US)
- (*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.
- (21) Appl. No.: 13/501,895
- (22) PCT Filed: Oct. 14, 2010
- (86) PCT No.: PCT/US2010/052677
 § 371 (c)(1),
 (2), (4) Date: Apr. 13, 2012
- (87) PCT Pub. No.: WO2011/047158PCT Pub. Date: Apr. 21, 2011

(65) **Prior Publication Data**

US 2012/0201848 A1 Aug. 9, 2012

Related U.S. Application Data

- (60) Provisional application No. 61/251,432, filed on Oct. 14, 2009.
- (51) Int. Cl.

A61K 39/12	(2006.01)
A61K 39/23	(2006.01)
C07K 14/015	(2006.01)

- (52) U.S. Cl. USPC 424/202.1; 424/233.1; 435/235.1; 435/320.1; 530/350; 536/23.72
- (58) Field of Classification Search CPC A61K 39/12; A61K 39/23; C07K 14/015 See application file for complete search history.

(56) References Cited

U.S. PATENT DOCUMENTS

4,904,468	A *	2/1990	Gill et al.	424/202.1
5,756,103	Α	5/1998	Paoletti et al.	
5,885,585	Α	3/1999	Parrish et al.	
7,736,658	B2 *	6/2010	Dominowski et al	424/201.1
2008/0014260	A1*	1/2008	Seager	424/458
2012/0201848	A1*	8/2012	Kapil	424/202.1

FOREIGN PATENT DOCUMENTS

CN	101035558 A1	9/2007
EP	0 863151 A1	9/1998
WO	WO 2008/157236 A1	12/2008

OTHER PUBLICATIONS

US 8,734,808 B2

May 27, 2014

Nakamura et al. (Archives of Virology. 2004; 149: 2261-2269).* EF599098.2. NCBI-GenBank (online) Feb. 2008 [retrieved on Dec. 29, 2010] Retrieved from the Internet URL:http://www.ncbi.nim.

nih.gov/nucleotide/160250242?report=genbank&log\$= nuclalign&blast_rank=86&RID=H1ZCHUHG01P. See nucleotide sequence and amino acid sequence.

Q98VH1.VP2.NCBI-GenBank (online) Nov. 28, 2006 [retrieved on Dec. 20, 2010] Retrieved from the Internet URL: http://www.ncbi.nim.nih.gov/protein/81965481>. See amino acid sequence.

Ikeda et al., "Predominance of Canine Parvovirus (CPV) in Unvaccinated Cat Populations and Emergence of New Antigenic Types of CPVs in Cats", "Virology", 2000, pp. 13-19, vol. 278, Publisher: Academic Press, Published in: US.

"PCT International Search Report, Application No. PCT/US2010/ 52677, mailed Jan. 13, 2011".

"PCT Written Opinion, Application No. PCT/US2010/52677, mailed Jan. 13, 2011".

Allison, et al., "Role of Multiple Hosts in the Cross-Species Transmission and Emergence of a Pandemic Parvovirus", Nov. 9, 2011, pp. 865-872, vol. 86, No. 2, Publisher: Journal of Virology, Published in: US.

Allison, Andrew et al., "Frequent Cross-Species Transmission of Parvoviruses Among Diverse Carnivore Hosts", Feb. 2013, pp. 2342-2347, vol. 87, No. 4, Publisher: Journal of Virology, Published in: US. Kang, et al., "Prevalance and Genetic Characterization of Canine Parvoviruses in Korea", Jan. 3, 2008, vol. 36, No. 1, Publisher: Virus Genes, Kluwer Academic Publishers, Published in: US.

Harbison, et al., "The Parvovirus Capsid Odyssey: From the Cell Surface to the Nucleus", May 1, 2008, vol. 16, No. 5, Publisher: Trends in Microbiology, Elsevier Science Ltd., Published in: GB.

Parrish et al., "Canine Host Range and a Specific Epitope Map Along With Variant Sequences in the Capsid Protein Gene of Canine Parvoviru", Oct. 1, 1998, pp. 293-307, vol. 166, No. 2, Publisher: Virology, Elsevier, Amsterdam, NL, Published in: NL.

EP10824094, Supplementary European Search Report, Feb. 12, 2013.

Mochizuki, et al., "Isolation of a Canine Parvovirus From a Cat Manifesting Clinical Signs of Feline Panleukopenia", Sep. 1996, pp. 2101-2105, vol. 34, No. 9, Publisher: Journal of Clinical Microbiology, Published in: US.

Tsai, Chich H., et al., "Localization of the VP2 Protein of Canine Parvovirus Type 2 on the Baculovirus Envelop and Its Immunogenicity in a Mouse Model", Dec. 2012, pp. 178-185, No. 2, Published in: US.

* cited by examiner

Primary Examiner — Shannon A Foley

(74) Attorney, Agent, or Firm—Fellers, Snider, Blankenship, Bailey & Tippens, P.C.; Terry L. Watt

(57) ABSTRACT

Vaccines preparations against canine parvovirus are provided. The vaccines include a novel canine parvovirus-2 isolated from a raccoon, and related nucleic acids and proteins.

13 Claims, 12 Drawing Sheets

FIGURE 1



Number of cases

ATGAGTGATGGAGCAGTTCAACCAGACGGTGGTCAACCTGCTGTCAGAAATGAAA TGTGGGGATTTCTACGGGTACTTTCAATAATCAGACGGAATTTAAATTTTTGGAA AACGGATGGGTGGAAATCACAGCAAACTCAAGCAGACTTGTACATTTAAATATGC CAGAAAGTGAAAATTATAGAAGAGTGGTTGTAAATAATTTAGATAAAACTGCAGT TAACGGAAACATGGCTTTAGATGATACTCATGCACAAATTGTAACACCTTGGTCA TTGGTTGATGCAAATGCTTGGGGAGTTTGGTTTAATCCAGGAGATTGGCAACTAA TTGTTAATACTATGAGTGAGTTGCATTTAGTTAGTTTTGAACAAGAAATTTTTAA TGTTGTTTTAAAGACTGTTTCAGAATCTGCTACTCAGCCACCAACTAAAGTTTAT AATAATGATTTAACTGCATCATTGATGGTTGCATTAGATAGTAATAATACTATGC CATTTACTCCAGCAGCTATGAGATCTGAGACATTGGGTTTTTATCCATGGAAACC AACCATACCAACTCCATGGAGATATTATTTTCAATGGGATAGAACATTAATACCA TCTCATACTGGAACTAGTGGCACACCAACAAATACATACCATGGTACAGATCCAG ATGATGTTCAATTTTATACTATTGAAAATTCTGTGCCAGTACACTTACTAAGAAC AGGTGATGAATTTGCTACAGGAACATTTTTTTTTTGATTGTAAACCATGTAGACTA ACACATACATGGCAAACAAATAGAGCATTGGGCTTACCACCATTTCTAAATTCTT TGCCTCAATCTGAAGGAGATACTAACTTTGGTGATATAGGAATTCAACAAGATAA AAGACGTGGTGTAACTCAAATGGGAAATACAAACTATATTACTGAAGCTACTATT ATGAGACCAGCTGAGGTTGGTTATAGTGCACCATATTATTCTTTTGAGGCGTCTA TGAAAATCAAGCAGCAGATGGTGATCCAAGATATGCATTTGGTAGACAACATGGT CAAAAAACTACCACAACAGGAGAAACACCTGAGAGATTTACATATAGCACATC AAGATACAGGAAGATATCCAGAAGGAGATTGGATTCAAAATATTAACTTTAACCT TCCTGTAANAAATGATAATGTATTGCTACCAACAGATCCAATTGGAGGTAAAACA GGAATTAACTATACTAATATATTTAATACTTATGGTCCTTTAACTGCATTAAATA ATGTACCACCAGTTTATCCAAATGGTCAAATTTGGGATAAAGAATTTGATACTGA GGTCAATTATTTGTAAAAGTTGCGCCTAATTTAACAAATGAATATGATCCTGATG CATCTGCTAATATGTCAAGAATTGTAACTTACTCAGATTTTTGGTGGAAAGGTAA ATTAGTATTTAAAGCTAAACTAAGAGCCTCTCATACTTGGAATCCAATTCAACAA ATGAGTATTAATGTAGATAACCAATTTAACTATGTACCAAGTAATATTGGAGGTA TGAAAATTGTATATGAAAAATCTCAACTAGCACCTAGAAAATTATAT**TAA**CATAC TTACTATGTTTTTATGTTTATTACATATCAACTAGCACCA

(SEQ ID NO: 1)

ATGAGTGATGGAGCAGTTCAACCAGACGGTGGTCAACCTGCTGTCAGAAATGAAA TGTGGGGATTTCTACGGGTACTTTCAATAATCAGACGGAATTTAAATTTTTGGAA AACGGATGGGTGGAAATCACAGCAAACTCAAGCAGACTTGTACATTTAAATATGC CAGAAAGTGAAAATTATAGAAGAGTGGTTGTAAATAATTTAGATAAAACTGCAGT TAACGGAAACATGGCTTTAGATGATACTCATGCACAAATTGTAACACCTTGGTCA TTGGTTGATGCAAATGCTTGGGGAGTTTGGTTTAATCCAGGAGATTGGCAACTAA TTGTTAATACTATGAGTGAGTTGCATTTAGTTAGTTTTGAACAAGAAATTTTTAA TGTTGTTTTAAAGACTGTTTCAGAATCTGCTACTCAGCCACCAACTAAAGTTTAT AATAATGATTTAACTGCATCATTGATGGTTGCATTAGATAGTAATAATACTATGC CATTTACTCCAGCAGCTATGAGATCTGAGACATTGGGTTTTTATCCATGGAAACC AACCATACCAACTCCATGGAGATATTATTTTCAATGGGATAGAACATTAATACCA TCTCATACTGGAACTAGTGGCACACCAACAAATACATACCATGGTACAGATCCAG ATGATGTTCAATTTTATACTATTGAAAATTCTGTGCCAGTACACTTACTAAGAAC AGGTGATGAATTTGCTACAGGAACATTTTTTTTTGATTGTAAACCATGTAGACTA ACACATACATGGCAAACAAATAGAGCATTGGGCTTACCACCATTTCTAAATTCTT TGCCTCAATCTGAAGGAGATACTAACTTTGGTGATATAGGAATTCAACAAGATAA AAGACGTGGTGTAACTCAAATGGGAAATACAAACTATATTACTGAAGCTACTATT ATGAGACCAGCTGAGGTTGGTTATAGTGCACCATATTATTCTTTTGAGGCGTCTA TGAAAATCAAGCAGCAGATGGTGATCCAAGATATGCATTTGGTAGACAACATGGT CAAAAAACTACCACAACAGGAGAAACACCTGAGAGATTTACATATAGCACATC AAGATACAGGAAGATATCCAGAAGGAGATTGGATTCAAAATATTAACTTTAACCT TCCTGTAANAAATGATAATGTATTGCTACCAACAGATCCAATTGGAGGTAAAACA **GGAATTAACTATACTAATATATTTAATACTTATGGTCCTTTAACTGCATTAAATA** ATGTACCACCAGTTTATCCAAATGGTCAAATTTGGGATAAAGAATTTGATACTGA GGTCAATTATTTGTAAAAGTTGCGCCTAATTTAACAAATGAATATGATCCTGATG CATCTGCTAATATGTCAAGAATTGTAACTTACTCAGATTTTTGGTGGAAAGGTAA ATTAGTATTTAAAGCTAAACTAAGAGCCTCTCATACTTGGAATCCAATTCAACAA ATGAGTATTAATGTAGATAACCAATTTAACTATGTACCAAGTAATATTGGAGGTA TGAAAATTGTATATGAAAAATCTCAACTAGCACCTAGAAAATTATAT

(SEO ID NO: 2)

Met Ser Asp Gly Ala Val Gln Pro Asp Gly Gly Gln Pro Ala Val Arg 1 5 10 15 Asn Glu Arg Ala Thr Gly Ser Gly Asn Gly Ser Gly Gly Gly Gly Gly 20 25 30 Gly Gly Ser Gly Gly Val Gly Ile Ser Thr Gly Thr Phe Asn Asn Gln 35 40 45 Thr Glu Phe Lys Phe Leu Glu Asn Gly Trp Val Glu Ile Thr Ala Asn 50 55 60 Ser Ser Arg Leu Val His Leu Asn Met Pro Glu Ser Glu Asn Tyr Arg 65 70 75 80 Arg Val Val Asn Asn Leu Asp Lys Thr Ala Val Asn Gly Asn Met 85 90 95 Ala Leu Asp Asp Thr His Ala Gln Ile Val Thr Pro Trp Ser Leu Val 100 105 110 Asp Ala Asn Ala Trp Gly Val Trp Phe Asn Pro Gly Asp Trp Gln Leu 115 120 125 Ile Val Asn Thr Met Ser Glu Leu His Leu Val Ser Phe Glu Gln Glu 130 135 140 Ile Phe Asn Val Val Leu Lys Thr Val Ser Glu Ser Ala Thr Gln Pro 145 150 155 160 Pro Thr Lys Val Tyr Asn Asn Asp Leu Thr Ala Ser Leu Met Val Ala 165 170 175 Leu Asp Ser Asn Asn Thr Met Pro Phe Thr Pro Ala Ala Met Arg Ser 180 185 190 Glu Thr Leu Gly Phe Tyr Pro Trp Lys Pro Thr Ile Pro Thr Pro Trp 195 200 205 Arg Tyr Tyr Phe Gln Trp Asp Arg Thr Leu Ile Pro Ser His Thr Gly 210 215 220 Thr Ser Gly Thr Pro Thr Asn Thr Tyr His Gly Thr Asp Pro Asp Asp 225 230 235 240 val Gln Phe Tyr Thr Ile Glu Asn Ser Val Pro Val His Leu Leu Arg 245 250 255 250 Thr Gly Asp Glu Phe Ala Thr Gly Thr Phe Phe Asp Cys Lys Pro 260 265 270 Cys Arg Leu Thr His Thr Trp Gln Thr Asn Arg Ala Leu Gly Leu Pro 275 280 285 Pro Phe Leu Asn Ser Leu Pro Gln Ser Glu Gly Asp Thr Asn Phe Gly 290 295 300 Asp Ile Gly Ile Gln Gln Asp Lys Arg Arg Gly Val Thr Gln Met Gly 305 310 315 320 Asn Thr Asn Tyr Ile Thr Glu Ala Thr Ile Met Arg Pro Ala Glu Val 325 330 335

Gly Tyr Ser Ala Pro Tyr Tyr Ser Phe Glu Ala Ser Thr Gln Gly Pro 340 345 350 Phe Lys Thr Pro Ile Ala Ala Gly Arg Gly Gly Ala Gln Thr Asp Glu 355 360 365 Asn Gln Ala Ala Asp Gly Asp Pro Arg Tyr Ala Phe Gly Arg Gln His 370 375 380 Gly Gln Lys Thr Thr Thr Gly Glu Thr Pro Glu Arg Phe Thr Tyr 385 390 395 400 Ile Ala His Gln Asp Thr Gly Arg Tyr Pro Glu Gly Asp Trp Ile Gln 405 410 415 Asn Ile Asn Phe Asn Leu Pro Val Xaa Asn Asp Asn Val Leu Leu Pro 420 425 430 Thr Asp Pro Ile Gly Gly Lys Thr Gly Ile Asn Tyr Thr Asn Ile Phe 435 440 445 Asn Thr Tyr Gly Pro Leu Thr Ala Leu Asn Asn Val Pro Pro Val Tyr 450 455 460 Pro Asn Gly Gln Ile Trp Asp Lys Glu Phe Asp Thr Asp Leu Lys Pro 465 470 475 480 Arg Leu His Val Asn Ala Pro Phe Val Cys Gln Asn Asn Cys Pro Gly 485 490 495 Gln Leu Phe Val Lys Val Ala Pro Asn Leu Thr Asn Glu Tyr Asp Pro 500 505 510 Asp Ala Ser Ala Asn Met Ser Arg Ile val Thr Tyr Ser Asp Phe Trp 515 520 525 Trp Lys Gly Lys Leu Val Phe Lys Ala Lys Leu Arg Ala Ser His Thr 530 535 540 Trp Asn Pro Ile Gln Gln Met Ser Ile Asn Val Asp Asn Gln Phe Asn 545 550 555 560 Tyr Val Pro Ser Asn Ile Gly Gly Met Lys Ile Val Tyr Glu Lys Ser 565 570 575 Gln Leu Ala Pro Arg Lys Leu Tyr His Thr Tyr Tyr Val Phe Met Phe 580 585 590 Ile Thr Tyr Gln Pro Ser Thr 595

FIGURE 3A (con't)

Met Ser Asp Gly Ala Val Gln Pro Asp Gly Gly Gln Pro Ala Val Arg 1 5 10 15 Asn Glu Arg Ala Thr Gly Ser Gly Asn Gly Ser Gly Gly Gly Gly Gly 20 25 30 Gly Gly Ser Gly Gly Val Gly Ile Ser Thr Gly Thr Phe Asn Asn Gln 35 40 45 Thr Glu Phe Lys Phe Leu Glu Asn Gly Trp Val Glu Ile Thr Ala Asn 50 55 60 Ser Ser Arg Leu Val His Leu Asn Met Pro Glu Ser Glu Asn Tyr Arg 65 70 75 80 Arg Val Val Asn Asn Leu Asp Lys Thr Ala Val Asn Gly Asn Met 85 90 95 Ala Leu Asp Asp Thr His Ala Gln Ile Val Thr Pro Trp Ser Leu Val 100 105 110 Asp Ala Asn Ala Trp Gly Val Trp Phe Asn Pro Gly Asp Trp Gln Leu 115 120 125 Ile Val Asn Thr Met Ser Glu Leu His Leu Val Ser Phe Glu Gln Glu 130 135 140 Ile Phe Asn Val Val Leu Lys Thr Val Ser Glu Ser Ala Thr Gln Pro 145 150 155 160 Pro Thr Lys Val Tyr Asn Asn Asp Leu Thr Ala Ser Leu Met Val Ala 165 170 175 Leu Asp Ser Asn Asn Thr Met Pro Phe Thr Pro Ala Ala Met Arg Ser 180 185 190 Glu Thr Leu Gly Phe Tyr Pro Trp Lys Pro Thr Ile Pro Thr Pro Trp 195 200 205 Arg Tyr Tyr Phe Gln Trp Asp Arg Thr Leu Ile Pro Ser His Thr Gly 210 215 220 Thr Ser Gly Thr Pro Thr Asn Thr Tyr His Gly Thr Asp Pro Asp Asp 225 230 235 240 Val Gln Phe Tyr Thr Ile Glu Asn Ser Val Pro Val His Leu Leu Arg 245 250 255 Thr Gly Asp Glu Phe Ala Thr Gly Thr Phe Phe Asp Cys Lys Pro 260 265 270 Cys Arg Leu Thr His Thr Trp Gln Thr Asn Arg Ala Leu Gly Leu Pro 275 280 285 Pro Phe Leu Asn Ser Leu Pro Gln Ser Glu Gly Asp Thr Asn Phe Gly 290 295 300 Asp Ile Gly Ile Gln Gln Asp Lys Arg Arg Gly Val Thr Gln Met Gly 305 310 315 320 Asn Thr Asn Tyr Ile Thr Glu Ala Thr Ile Met Arg Pro Ala Glu Val 325 330 330

Gly Tyr Ser Ala Pro Tyr Tyr Ser Phe Glu Ala Ser Thr Gln Gly Pro 340 345 350 Phe Lys Thr Pro Ile Ala Ala Gly Arg Gly Gly Ala Gln Thr Asp Glu 355 360 365 Asn Gln Ala Ala Asp Gly Asp Pro Arg Tyr Ala Phe Gly Arg Gln His 370 375 380 Gly Gln Lys Thr Thr Thr Gly Glu Thr Pro Glu Arg Phe Thr Tyr 385 390 395 400 Ile Ala His Gln Asp Thr Gly Arg Tyr Pro Glu Gly Asp Trp Ile Gln 405 410 415 Asn Ile Asn Phe Asn Leu Pro Val Xaa Asn Asp Asn Val Leu Leu Pro 420 425 430 Thr Asp Pro Ile Gly Gly Lys Thr Gly Ile Asn Tyr Thr Asn Ile Phe 435 440 445 Asn Thr Tyr Gly Pro Leu Thr Ala Leu Asn Asn Val Pro Pro Val Tyr 450 455 460 Pro Asn Gly Gln Ile Trp Asp Lys Glu Phe Asp Thr Asp Leu Lys Pro465470475480 Arg Leu His Val Asn Ala Pro Phe Val Cys Gln Asn Asn Cys Pro Gly 485 490 495 Gln Leu Phe Val Lys Val Ala Pro Asn Leu Thr Asn Glu Tyr Asp Pro 500 505 510 Asp Ala Ser Ala Asn Met Ser Arg Ile Val Thr Tyr Ser Asp Phe Trp 515 520 525 Trp Lys Gly Lys Leu Val Phe Lys Ala Lys Leu Arg Ala Ser His Thr 530 535 540 Trp Asn Pro Ile Gln Gln Met Ser Ile Asn Val Asp Asn Gln Phe Asn 545 550 555 560 Tyr Val Pro Ser Asn Ile Gly Gly Met Lys Ile Val Tyr Glu Lys Ser 565 570 575 Gln Leu Ala Pro Arg Lys Leu Tyr 580

FIGURE 3B (con't)



Figure 4A-C











			· · · · · · · · · · · · · · · · · · ·										
Virus	80	87	93	101	103	232	297	300	305	426	555	564	568
FPV	Lys	Met	Lys	Ile	Val	Val	Ser	Ala	Asp	Asn	Val	Asn	Ala
CPV-2	Arg	Met	Asn	Ile	Val	Ile	Ser	Ala	Asp	Asn	Ile	Ser	Gly
CPV-2a	Arg	Leu	Asn	Thr	Ala	Ile	Ser	Gly	Tyr	Asn	Val	Ser	Gly
CPV-2b	Arg	Leu	Asn	Thr	Ala	Ile	Ser	Gly	Tyr	Asp	Val	Ser	Gly
CPV-2c	Arg	Leu	Asn	Thr		Ile	Ala	Gly	Tyr	Glu	Val	Ser	Gly
MEV	Lys	Leu	Lys	Ile		Val	Ser	Ala	Asp	Asn	Val	Asn	Ala
RPV	Arg	Leu	Asn	Thr	Ala	Thr	Ser	Asp	Asp	Asn	Val	Ser	Gly

Fig. 7



FIGURE 8

ISOLATION OF A VIRUS RELATED TO CANINE PARVOVIRUS-2 FROM A RACCOON

SEQUENCE LISTING

This application includes as the Sequence Listing the complete contents of the accompanying text file "PCT Sequence Listing_ST25.txt", created Oct. 14, 2010, containing 18,268 bytes, hereby incorporated by reference.

DESCRIPTION

BACKGROUND OF THE INVENTION

1. Field of the Invention

The invention generally relates to improved vaccines against canine parvovirus-like viruses. In particular, the invention provides vaccines suitable for puppies which are based on a novel parvovirus isolated from a raccoon.

2. Background of the Invention

Canine parvovirus (CPV) is primarily an enteric pathogen that infects dogs, especially young dogs. Parvovirus infection is characterized by acute diarrhea, fever and leukopenia in dogs and puppies more than 4 to 5 weeks old, and myocardial disease in younger puppies. The mortality rate from the dis- 25 ease in unvaccinated dogs is very high. While vaccines against CPV are available, because CPV is a single-stranded DNA virus and has an extreme ability to mutate, the virus shows a remarkable ability to vary antigenically (Parrish and Kawaoka 2005) and thereby elude the immune protection 30 afforded by vaccines. Thus, constant monitoring of the antigenic type and genotype of circulating viruses, and adjustment of vaccine components accordingly, is necessary.

Newborn puppies acquire immunities against diseases such as CPV infection by nursing from their mother, espe-35 cially during the first two days of life. A puppy that nurses takes in colostrum in the milk that is first produced and antibodies in the colostrum are passed to the puppy. For dogs and many other mammals as well, the immunity given by the colostrum loses its affect sometime around the fifth week of 40 age.

A particular challenge when vaccinating puppies is to administer vaccines according to a time frame that provides protection which overlaps the protection provided by maternal antibodies and begins as maternal antibodies wane. Cur- 45 rently, vaccine regimens for puppies typically begin at about 6 weeks of age and boosters are given about every 3 weeks thereafter, e.g. at 9, 12 and 15 weeks. However, in order for this regimen to provide full protection, the first vaccine dose must immediately elicit a protective immune response. This is 50 completely unrealistic due to the immaturity of the puppy's immune system and the time period required to mount an immune response. Full protection usually does not develop until the entire course of vaccinations is given. The age-based mortality due to CPV is depicted in FIG. 1, which shows that 55 maximum mortality due to CPV occurs before vaccine protocols can be completed.

A simple answer might be to begin the vaccination program even earlier, e.g. at 2-3 weeks. However, this would be futile because, for puppies whose mothers have been vacci- 60 nated with or otherwise exposed to a viral strain with the same antigenic determinants, maternal antibodies passed to the puppy would neutralize viruses in the vaccine, thereby preventing the puppy's own immune system from responding to the virus. 65

Another challenge in veterinary medicine is the treatment of cancer, e.g., in dogs. There are many limitations in the

existing tools for cancer therapy, especially for geriatric dogs. The administration of oncolytic parvoviruses to kill cancer cells shows great promise as an effective cancer treatment (Rommelaere et al, Cvtokine & Growth Factor Reviews 21:185-195, 2010; and U.S. Pat. No. 7,179,456 to Rommelaere et al, the complete contents of which are herein incorporated by reference) and might be applied to canines. However, the existence of pre-existing antibodies to parvoviruses (e.g. as a result of vaccination) would render this 10 method ineffective, since the parvovirus would be neutralized by the existing antibodies. In addition, gene therapy in dogs is rarely undertaken at present but would be a promising method for treating several disorders, if suitable nucleic acid vectors are identified.

The prior art has thus far failed to provide solutions to these problems.

SUMMARY OF THE INVENTION

The present invention is based on the discovery of novel 20 parvoviruses (isolated from raccoons in the USA). Analyses of nucleotide and amino acid sequences showed that the viruses represent a unique variant of canine parvovirus type 2 (CPV-2). This new CPV-2, denominated "AR08071304" (and referred to herein as raccoon parvovirus, "RPV", "08071304", and "Parvovirus-2 (Raccoon)" etc.) is useful as a component of vaccines against canine parvovirus. In particular, the variant is useful as a component of vaccines for very young dogs, e.g. puppies who are still nursing or who are in the process of being weaned, and thus solves the problems of how to provide adequate puppy vaccine protocols. This raccoon parvovirus will not be completely neutralized by existing canine parvovirus-2 antibodies in dogs. In addition, this raccoon parvovirus can be used as an oncolytic agent and as a vector (carrier, vehicle) for gene therapy, due to the lack of neutralization by pre-existing antibodies in a recipient or host organism, e.g. a dog.

The variant virus has close homology to the predominant CPV-2 viruses currently in circulation. Thus, administration of a vaccine containing the new variant is likely to result in an immune response that also affords protection against known circulating CPV-2 viruses. However, there are some antigenic differences between the new isolate and known CPV-2 viruses to which a mother dog is likely to have been exposed (e.g. via a vaccine preparation or prior CPV exposure). In particular, amino acid residue 232 of the variant capsid protein. VP-2 is Thr (encoded by ACA) and amino acid residue 300 is Asp (encoded by GAT). These differences in sequence are sufficient to result in a unique immunoreactive profile for the variant. Therefore, maternal antibodies passed to a puppy (e.g. from a previously vaccinated/exposed mother) are not likely to completely inactivate the new virus when it is administered to a puppy as a vaccine. As a result, an immune response by the puppy's own immune system will be elicited upon exposure to the RPV of the present invention via vaccination, and the immune response is likely to be broadly protective against currently circulating CPV's in general. Moreover, this raccoon parvovirus appears to be a natural chimera of carnivore parvoviruses. Although the nucleotide sequence encoding the RPV VP-2 protein has the highest total homology score (3309) with canine parvovirus-2a (CPV-15), it also has high homology with feline panleukopaenia, previously isolated raccoon parvoviruses, and mink enteritis virus. This type of chimeric sequence has not been observed previously in canine or feline parvovirus sequences, although similar parvoviruses now appear to be circulating in raccoon populations in the USA (see Examples section below). This RPV

will provide broad immune protection in several carnivore species and is thus suitable for use as a broad spectrum parvovirus vaccine. Under rare, unusual circumstances, a dog or cat might succumb to infection with a heterologus live, nonattenuated carnivore parvovirus; however, the benefits of 5 using the RPV of the invention as described herein far outweigh this minimal risk.

Although the present invention is not premised on any evolutionary model or theory, one interesting interpretation of the genetic data disclosed herein is that the RPV of the 10 present invention may represent the ancestor of present-day canine and feline parvoviruses. There is an ongoing debate in virology community concerning the origin of parvovirus-2 of dogs, and the RPV of the invention would, within the confines of this theory, be a "living fossil" of canine parvovirus-2. In any case, the RPV described herein encodes a unique VP-2 protein, which comprises a mosaic of feline and canine parvovirus amino acid sequences. Thus the RPV of the present invention is a candidate for being the "ancestor" of both feline and canine parvoviruses, i.e., these newer carnivore parvovi- 20 ruses having originated from this RPV. Alternatively, the RPV of the present invention may represent a chimeric intermediate at the "intersection" of parvovirus evolution. In any case, this RPV provides an avenue for cross-immunization against heterologous parvoviruses that is otherwise not presently 25 available with any of the more species-specialized canine and feline parvoviruses. Moreover, the RPV of the invention is weakly agglutinating for porcine erythrocytes (transferrin); does not share some critical epitopes (e.g., amino acid residue 300 of the VP-2 protein sequence); and grows poorly in feline 30 kidney cell lines (see Examples below). Taken together, these indications are consistent with a virus that either predates or is an early intermediate in the evolutionary cascade of canine and feline parvoviruses.

The invention provides a parvovirus comprising the char-35 acteristics of the Parvovirus-2 (Raccoon) of ATCC NO. PTA-11400, deposited at the American Type Culture Collection, 10801 University Blvd, Manassas, Va., on Oct. 7, 2010.

In addition, the invention provides vaccines comprising a parvovirus comprising the characteristics of the Parvovirus-2 40 (Raccoon) having the ATCC NO. PTA-11400. In one embodiment, the vaccine further comprises one or more antigenic components selected from the group consisting of canine distemper virus (CDV), canine adenovirus type 2, canine parainfluenza virus, canine corona virus, canine herpes virus, 45 canine rotavirus, one or more Leptospira serovars, and a canine parvovirus-2 in which the amino acid at position 232 of VP-2 protein is not Thr and the amino acid at position 300 of VP-2 protein is not Asp. In some embodiments, the one or more Leptospira serovars is selected from the group consist- 50 ing of Leptospira interrogans serovar canicolar, Leptospira interrogans serovar icterohaemorrhagiae, Leptospira interrogans serovar Pomona, and Leptospira kirschneri serovar grippotyphosa.

The invention also provides isolated nucleic acids comprising the nucleotide sequence SEQ ID NO: 1, or a portion of SEQ ID NO: 1; wherein the portion of SEQ ID NO: 1 encodes an antigenic region of a VP-2 protein comprising amino acid residue 232 of SEQ ID NO: 3, amino acid residue 300 of SEQ ID NO: 3 or both amino acid residue 232 and amino acid residue 300 of SEQ ID NO: 3. In other embodiments, the invention provides immununogenic compositions comprising this nucleic acid, wherein the nucleic acid is present in a killed or attenuated parvovirus virion or low passage raccoon parvovirus (RPV). In some embodiments, the killed parvovi-65 rus virion comprises the characteristics of Parvovirus-2 (Raccoon) having the ATCC NO. PTA-11400. In other embodi4

ments, the attenuated parvovirus virion is present in a solid carrier suitable for supralingual dissolution. In yet other embodiments, the immununogenic composition is suitable for subcutaneous administration.

The invention also provides a method of eliciting an immune response in an animal against parvovirus infection. The method comprises the step of administering to the animal an immunogenic composition comprising a nucleic acid comprising the nucleotide sequence SEQ ID NO: 1, or a portion of SEQ ID NO: 1; wherein said portion of SEQ ID NO: 1 encodes an antigenic region of a VP-2 protein comprising amino acid residue 232 of SEQ ID NO: 3, amino acid residue 300 of SEQ ID NO: 3 or both amino acid residue 232 and amino acid residue 300 of SEQ ID NO: 3. In some embodiments, the nucleic acid is present in a killed or attenuated parvovirus virion or low passage RPV in a killed or attenuated parvovirus virion. In some embodiments, the killed parvovirus virion comprises the characteristics of Parvovirus-2(Raccoon) having the ATCC NO. PTA-11400. In some embodiments, the attenuated parvovirus virion is present in a solid carrier suitable for supralingual dissolution, and in some embodiments, the animal is a puppy. In other embodiments, the immununogenic composition is suitable for subcutaneous administration.

The invention further provides a substantially purified parvovirus VP-2 protein that has a threonine residue at amino acid position 232 and an aspartic acid residue at amino acid position 300; or an antigenic fragment thereof, wherein said antigenic fragment comprises at least a portion of said VP-2 protein that has: a threonine residue at amino acid position 232; or an aspartic acid residue at amino acid position 300; or both a threonine residue at amino acid position 232 and an aspartic acid residue at amino acid position 232 and an aspartic acid residue at amino acid position 300. In some embodiments, the substantially purified parvovirus VP-2 protein comprises amino acid sequence SEQ ID NO: 3.

The invention also provides expression vectors comprising a nucleotide sequence encoding the amino acid sequence of SEQ ID NO: 3, or a portion of SEQ ID NO: 3; wherein said portion of SEQ ID NO: 3 is an antigenic region of a VP-2 protein comprising amino acid residue 232 of SEQ ID NO: 3, amino acid residue 300 of SEQ ID NO: 3 or both amino acid residue 232 and amino acid residue 300 of SEQ ID NO: 3. In some embodiments, the expression vector is a recombinant viral expression vector. In other embodiments, the recombinant viral expression vector is a canarypox viral expression vector.

The invention also provides methods of killing tumor cells in a mammal. The methods comprise the step of administering to the mammal a composition comprising a parvovirus comprising the characteristics of the Parvovirus-2 (Raccoon) of ATCC NO. PTA-11400, in a quantity sufficient to infect and kill said tumor cells in the mammal. In some embodiments, the mammal is a dog that is seropositive for canine parvovirus-2. In other embodiments, the method is carried out by a route of administration selected from the group consisting of intravenous and intratumoral. In yet other embodiments, the parvovirus is a low passage parvovirus.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1. Distribution of CPV-2 genotypes and number of fatal cases of CPV infection with respect to age of dog.

FIGS. **2**A and B. Nucleic acid sequences encoding the VP-2 proteins of the new AR08071304 CPV-2 variant. A, the longer version of the nucleotide sequence (SEQ ID NO: 1) which encodes a 599 amino acid protein. The internal stop codon located at nucleotides 1753-1755 is shown in bold and

underlined. B, the shorter version of the nucleotide sequence (SEQ ID NO: 2) which encodes a 584 amino acid protein. In both A and B, the nucleotides encoding Thr at position 232 (nts 694-696) and the nucleotides encoding Asp at position 300 (nts 898-900) are underlined.

FIGS. 3A and B. Amino acid sequence of the VP-2 protein of the new AR08071304 CPV-2 variant. A. (SEQ ID NO: 3), is the longer version of the protein when all 599 encoded amino acids are translated; B, (SEQ ID NO: 4), is the shorter version of the protein when termination occurs at the stop 10 condon. Amino acid positions 232 and 300, which are Thr and Asp, respectively in both proteins, are boxed.

FIG. 4A-C. Schematics of: A, raccoon parvovirus (RPV) DNA; B, RPV open reading frame arrangement; C, binding of RPV to eukaryotic cells.

FIGS. 5A and B.A, Schematic of raccoon parvovirus binding to cell surface transferrin receptor (TFR); B, schematic of general strategy for preparing autonomous parvovirus vectors

FIG. 6. Relatedness of the newly isolated raccoon parvovi- 20 rus to other similar viruses.

FIG. 7. Amino acids at critical epitopes in VP-2 protein of the newly isolated raccoon parvovirus.

FIG. 8. Phylogenetic tree comparing the VP-2 protein of the newly isolated raccoon parvovirus with those of other 25 parvoviruses. carnivore CPV=canine parvovirus; RPV=raccoon FPV=feline parvovirus; parvovirus; PV=parvovirus; MEV=mink enteritis virus; FPL=feline panleukopenia virus; Aleu=Aleutian mink disease parvovirus; MVC=minute virus of canines. 30

DETAILED DESCRIPTION

The invention provides vaccine and virotherapy preparations comprising attenuated forms of a novel parvovirus vari- 35 ant isolated from an infected raccoon. While the vaccine preparations are suitable for administration to a wide variety of carnivores (e.g. dogs, cats, mink, raccoon, members of canidae, procyonidea, mustelidae, and viveridae etc.), they are particularly useful for administration to very young dogs 40 (puppies). This is because the new variant differs enough from CPV variants that are used in current vaccines so as to make it unlikely that maternal antibodies (developed in response to vaccine administration and passed to the puppies while nursing) would recognize and inactivate the virus. 45 Similarly, for other applications in dogs, including adult dogs (e.g. as a treatment for cancer and as a gene therapy vector) the RPV will bypass the preexisting antibodies from previous vaccinations, and hence evade immediate immune clearance. Notably, in the VP-2 protein of the raccoon isolate amino acid 50 residue 232 is Thr (encoded by ACA) and residue 300 is Asp (encoded by GAT). These two amino acid residues in the RPV are thus distinct from those of the VP-2 proteins of all other known CPV isolates and may be responsible for the unique properties (e.g. antibody reactivity, see Examples below) of 55 this RPV. Due to attenuation, the virus will not cause disease in vaccine recipients. On the other hand, the new isolate is similar enough to viruses which are currently in circulation (e.g. the amino acid sequence identity is ~99% for the VP-2 protein) that is it likely that administration of the virus to a 60 young animal would result in the production of antibodies, at least some of which would cross react with known circulating viruses, and thus afford protection against those viruses to the vaccinated young animal.

A nucleic acid sequence described herein (SEQ ID NO: 1) 65 contains an internal stop codon at nucleotides 1753-1755 (see FIG. 2A). The VP-2 protein translated therefrom thus has two

6

different forms. One form, shown in FIG. 3A (SEQ ID NO 3) results from readthrough ("leakiness") of the stop codon and is thus longer (599 amino acids) than the second form, which is shown in FIG. **3**B, and which terminates at the stop codon. The second form of the protein contains only 584 amino acids (SEQ ID NO: 4). In the first, longer form of VP-2 the stop codon is not translated but is "skipped", resulting in the 599 amino acid protein. The term "VP-2" protein as used herein encompasses both the first longer form of VP-2 and the second, shorter form of VP-2, and both forms of the protein may be used in the practice of the invention. These two forms of the protein can be detected using Western blots. In addition, both nucleic acid sequences (SEQ ID NOS: 1 and 2) may be used in the practice of the present invention, e.g. either or both of the sequences may be used in a vaccine preparation, in a vector, etc. In some embodiments, the amino acid sequence of SEQ ID NO: 3 is encoded by the nucleotide sequence of SEQ ID NO: 1. In other embodiments, the amino acid sequence of SEQ ID NO: 4 is encoded by SEQ ID NO: 2. In yet other embodiments of the invention, SEQ ID NO: 1 may be modified by elimination (deletion) of the stop codon (e.g. by genetic engineering) so that the amino acid sequence of SEQ ID NO: 3 is translated therefrom.

This novel RPV also serves as a model for proper boosting of other animal and human infections. For example in human influenza (H1N1) based on available information, vaccine response varies depending on the existence of pre-existing antibodies. If a person has pre existing antibodies, the titer does not go up but the affinity of the antibodies does increase. If a person does not have a pre-existing titer, then the person does respond with development of neutralizing protective immunity.

The new isolate was deposited at the American Type Culture Collection in Manassas, Va., on Oct. 7, 2010, as Parvovirus-2 (Raccoon), on behalf of Oklahoma State University, (acknowledgement from the ATCC of receipt was received on Oct. 12, 2010), and assigned ATCC Deposit No. PTA-11400. The invention also provides an isolated parvovirus comprising the characteristics of ATCC Deposit No. PTA-11400, and progeny thereof. The invention further encompasses killed or attenuated parvoviruses comprising the characteristics of ATCC Deposit No. PTA-11400, deposited on Oct. 7, 2010 and progeny of the attenuated parvoviruses, as well as vaccines and immunogenic compositions comprising the same.

The invention also provides other RPV's isolated in Illinois and propagated in CRFK. (10071199-A and 10071199-C), the partial sequences of which are given in the Examples section. Thus, the invention also encompasses raccoon parvoviruses that are circulating in the USA (e.g. in Arkansas and Illinois), the common feature of which is that they do not react with the canine parvovirus monoclonal antibody (MoAb) 3B10, the MoAb that is most commonly used for diagnostics. 3B10 is specific for amino acid position 300 of VP-2, which is usually glycine. Thus, these RPV have a mutation at amino acid 300 of VP-2 that is included within the foot-print of this MoAb and prevents it from recognizing the proteins. Otherwise, these viruses have about 98% homology with the VP-2 of other carnivore parvoviruses-2. However, it is well-established that even a small number of mutations at critical locations can have a large impact on the properties of a virus (e.g. see Qu e al, 2005, where mutations in two critical amino acid residues of coronavirus spike protein, which altered its tropism, are described.)

Phylogenetic studies of this virus (e.g. see Examples 1 and 5) have shown that this RPV [Parvovirus-2 (Raccoon)] is closely related to, and in fact may be, a variant of canine

CPV-2. As such, domestic adult dogs may also be susceptible to the virus and would likely benefit from receiving vaccines which include this variant.

Transmission of the newly isolated parvovirus from raccoons to domestic pets such as dogs, and also to other wildlife 5 is likely to occur due to raccoon behavioral characteristics. For example, raccoons are highly intelligent and very quick to adapt to new food sources. They are omnivorous, eating a wide rage of plants and animals, depending upon season and availability. Due to their ready integration into and adaption 10 to urban areas, raccoons commonly interact with and make close contact with domestic dogs and cats. Raccoons have excellent prehensile abilities and readily learn, e.g. to open containers, to locate and open stored animal food, etc. Due to this behavior, they are potentially able to transmit parvovirus 1 to domestic carnivores. Raccoons also interact with other wildlife such as skunks and there is thus a potential for interspecies interaction and transmission of parvoviruses as well. Thus, adult dogs and other domestic pets would thus benefit from receiving vaccines which included the raccoon virus 20 described herein, even if they have previously been vaccinated with another parvovirus vaccine; and other species, whether domesticated or not, would also benefit from receiving such vaccines. The benefits could be two-fold: 1) vaccination would provide immunity to infection with this particu- 25 lar CPV; and 2) this CPV is similar enough to other parvoviruses (e.g. about 99% homologous to other carnivore parvoviruses) that is would likely also elicit and boost at least some immunity against them.

In some embodiments, the vaccines and immunogenic 30 compositions of the invention are monovalent in nature, i.e. that contain a single agent which is the parvovirus isolate described herein (e.g. with the characteristics of Parvovirus-2 (Raccoon), ATCC Deposit No. PTA-11400 deposited on Oct. 7, 2010), or an attenuated or killed form of the isolate 35 described herein, or progeny of any of these. In other embodiments, the vaccines and immunogenic compositions are polyvalent, i.e. they contain a plurality of antigenic agents, one of which is the isolate of the present invention. Exemplary additional components of the multivalent compositions include 40 but are not limited to one or more of a canine distemper viruses (CDV), canine adenovirus type 2, canine parainfluenza virus, canine corona virus, canine herpes virus, canine rotavirus, one or more Leptospira serovars, and a canine parvovirus-2 which differs in sequence of the VP-2 gene, i.e. 45 in which the amino acid at position 232 of the VP-2 protein is not Thr and/or the amino acid at position 300 of the VP-2 protein is not Asp; or in which the codon encoding Thr at amino acid position 232 of the VP-2 protein is not ACA, and/or the codon encoding Asp at amino acid position 300 of 50 the VP-2 is not GAT. Exemplary CDVs include but are not limited to those described in U.S. patent application Ser. No. 12/696,983 (Kapil), published as US2010/0196420, the complete contents of which are hereby incorporated by reference. Exemplary Leptospira serovars include but are not limited to 55 Leptospira interrogans serovar canicolar, Leptospira interrogans serovar icterohaemorrhagiae, Leptospira interrogans serovar pomona, and Leptospira kirschneri serovar grippotyphosa. Further, the isolate of the invention may be combined with a plurality of different antigens e.g. those of known 60 multivalent vaccines such as Galaxy® DA2PPV, or Nobivac DA2PPv+L4, etc.

In a particular embodiment, the recipient of a vaccine of the invention is a juvenile animal (e.g. a puppy) and the mode of administration is via a "puppy lollipop". Puppy lollipops for 65 use in administering vaccine compositions to young dogs (or the young of other species) are described in detail in co-

8

pending PCT patent application no. PCT/US2010/042142 to Kapil et al., filed Jul. 15, 2010, the complete contents of which is hereby incorporated by reference. Briefly, a puppy lollipop or similar device is used to deliver the vaccine to the dorsal side of the tongue, i.e. supralingually. In particular, the vaccine preparation is administered in a manner that delivers the vaccine at or near the basal cells of the tongue. Basal cells are infected by the attenuated parvovirus in the vaccine, and an antigen depot (e.g. a reservoir of virus) is established which gradually, over time and from a very young age, releases attenuated parvovirus within the puppy. This method takes advantage of the fact that the tongue is relatively immunologically privileged (devoid of immune response). The vaccine may, for example, be included in a hand-held "puppy lollipop" that is administered supralingually by a caregiver, e.g. by holding a stick, string or other delivery vehicle attached to a solid vaccine formulation which is placed in the puppy's mouth. The solid formulation includes a solid, inert carrier; with the parvovirus dispersed or distributed throughout the solid carrier, and the innate suckling response by the puppy gradually dissolves the carrier and the parvovirus is released to the tongue.

In one embodiment, the invention comprises nucleic acids with a nucleotide sequence encoding the VP-2 protein of the RPV, such as that which is set forth in SEQ ID NO: 1, or alternatively SEQ ID NO: 2 (see FIGS. 2A and B), or portions of that nucleotide sequence that encode antigenic regions or epitopes sufficient to elicit an immune response to the RPV in a vaccine recipient, and vaccines comprising the same. In particular, such nucleic acid sequences will include one or both of the codons which encode positions 232 and 300 of the VP-2 protein. Those codons are ACA and GAT, respectively, encoding Thr and Asp, respectively. In another embodiment, the invention comprises nucleic acids with a nucleotide sequence encoding a VP-2 protein comprising the amino acid sequence as set forth in SEQ ID NO: 3 (or alternatively, SEQ ID NO: 4, which is encompassed by SEQ ID NO: 3, see FIGS. 3A and B) or portions of the amino acid sequence set forth in SEQ ID NO: 3 which include antigenic regions or epitopes sufficient to elicit an immune response to the antigenic regions or epitopes in a vaccine recipient, and vaccines comprising the same. In particular, such proteins or portions thereof will include one or both of amino acid residues 232 and 300 of the RPV VP-2 protein, which are Thr and Asp, respectively. Variants and/or derivatives of such sequences, as described herein, and vaccine preparations which include the variants and/or derivatives, are also encompassed by the invention. For example, those of skill in the art will recognize that, due to the redundancy of the genetic code, sequences other than SEQ ID NO: 1 may be utilized to encode the amino acid sequence set forth in SEQ ID NO: 3. In addition, while the nucleotide sequence set forth in SEQ ID NO: 1 represents single-strand (ss)DNA, the invention also includes corresponding double-strand (ds) DNA, complementary DNA, and RNA of any form (e.g. mRNA, RNA/DNA hybrids, etc.) that is based on, derived from or that complements these sequences. Such sequences may be either sense or antisense sequences. Further, sequences which display at least about 50% homology, preferably about 60%, more preferably about 70, 80, or 90% homology, or even about 95, 96, 97, 98 or 99% or greater homology to SEQ ID NO: 1 are also contemplated for use in the vaccines, so long as the sequence encodes one or both of amino acid residues 232 and 300 as Thr and Asp, respectively. Such sequences may differ, for example, by containing alternate codons that encode the same amino acid at one or more positions.

2

In addition, portions of these sequences which encode antigenic regions or epitopes of the AR08071304 VP-2 protein are also contemplated. In particular, nucleotide sequences encompassing codons which encode one or both of amino acid residues 232 and 300 as Thr and Asp, respectively, 5 are contemplated. In one embodiment, the codon encoding amino acid residue 232 as Thr is ACA and the codon encoding residue 300 as Asp is GAT. In addition, sequences which encode amino acid sequences displaying 70%, or more preferably about 80, 90, or 95% or even greater identity (e.g. 96, 97, 98 or 99% identity) to SEQ ID NO: 3, so long as the sequence encodes one or both of amino acid residues 232 and 300 as Thr and Asp, respectively, or variants and/or derivatives thereof, including shorter antigenic regions or epitopes, are also included. Such variants and/or derivatives of SEQ ID 15 NO: 3 and/or antigenic regions or epitopes thereof may vary, for example, by containing conservative or non-conservative amino acid substitutions, or deletions (especially amino or carboxy terminal deletions), or various insertions, etc., so long as the resulting protein/peptide is antigenic and elicits an 20 immune response in a vaccine recipient. Such antigenic regions or epitopes are preferably at least about 10 amino acids in length and encompass one or more of amino acid positions 232 and 300, with reference to amino acid residue numbering in the VP-2 protein as depicted in FIG. 3, which 25 are Thr and Asp respectively. An antigenic region may, however, encompass the entire AR08071304 VP-2 gene/protein.

Further, nucleic acid sequences which hybridize to sequences disclosed herein (or to portions of those sequences) under stringent conditions (especially conditions 30 of high stringency) are also contemplated, so long as the sequence encodes one or both of amino acid residues 232 and 300 as Thr and Asp, respectively. Stringent conditions refer to hybridization conditions which allow a nucleic acid sequence to hybridize to a particular sequence. In general, high strin- 35 gent conditions refer to the hybridization conditions which allow a nucleic acid sequence of at least 50 nucleotides and preferably about 200 or more nucleotides to hybridize to a particular sequence at about 65° C. in a solution comprising about 1 M salt, preferably 6×SSC or any other solution having 40 a comparable ionic strength, and washing at 65° C. in a solution comprising about 0.1 M salt, or less, preferably $0.2 \times SSC$ or any other solution having a comparable ionic strength. These conditions allow the detection of sequences having about 90% or more sequence identity. In general, 45 lower stringent conditions refer to the hybridization conditions which allow a nucleic acid sequence of at least 50 nucleotides and preferably about 200 or more nucleotides to hybridize to a particular sequence at about 45° C. in a solution comprising about 1 M salt, preferably 6×SSC or any other 50 solution having a comparable ionic strength, and washing at room temperature in a solution comprising about 1 M salt, preferably 6×SSC or any other solution having a comparable ionic strength. These conditions allow the detection of sequences having up to 50% sequence identity. The person 55 skilled in the art will be able to modify these hybridization conditions in order to identify sequences varying in identity between 50% and 90%.

The invention thus also provides various types of recombinant and/or expression vectors that contain and express the 60 nucleic acid sequences disclosed herein (or portions thereof that encode antigenic peptides and/or polypeptides). Such vectors may be used in vaccine preparations, or may serve other purposes such as for manipulation of the disclosed sequence in a laboratory setting. Examples of such vectors 65 and expression systems include but are not limited to: various bacterial (e.g. *Escherichia coli*) or probiotic-based (e.g. *Lac*-

tobacillus) expression vectors; various recombinant viral vectors such as adenoviral vectors, baculovirus, canarypox vectors, etc.; *Pichia*, and yeast expression systems, etc. Such recombinant vectors and expression systems may be utilized, for example, in vaccine preparations, or, alternatively, for other purposes such as for laboratory manipulation of the sequences, or for research or diagnostic purposes.

The invention also contemplates chimeric proteins and genes encoding the same, such proteins comprising the VP-2 protein of the RPV of the invention, or antigenic regions or epitopes thereof. In certain embodiments, such epitopes are of a size in the range of from at least about 5 to about 20 or more amino acids. Such chimeric proteins (or nucleic acid sequences that encode them) may include other useful amino acid sequences, such as adjuvants, other proteins that are vaccine targets e.g. from other organisms that cause disease in carnivores, especially canids, etc. Nucleic acids encoding such proteins may include linker or spacer sequences e.g. between antigenic regions or epitopes. Also included are recombinant and/or expression vectors that encode and/or expresse such chimeric (or fusion) proteins.

Several methods of making vaccines suitable for vaccination against parvovirus are known in the art. See, for example, U.S. Pat. Nos. 4,193,990 and 4,193,991 to Appel et al., U.S. Pat. No. 4,303,645 to Carmichael et al., U.S. Pat. No. 4,971, 793 to Wood et al.; U.S. Pat. No. 5,882,652 to Valdes et al., and U.S. Pat. No. 5,885,585 to Parrish et al., each of which offers variations of suitable vaccine-formulating strategies. The complete contents of each of these patents are hereby incorporated by reference. Generally, to manufacture a vaccine, a viral vector containing the described nucleic acid sequences (e.g. ssDNA naturally occurring within a virus, or ssDNA or other equivalent form genetically engineered into a non-native viral vector (e.g. dsDNA, ss or dsRNA, RNA-DNA hybrids, etc.) will be employed. Examples include RPV (or other) viruses that are "killed", inactivated or otherwise attenuated so as to not cause severe disease symptoms in the animal to which it is administered, together with a suitable physiological carrier. Preferably, no disease symptoms will occur as a result of administration. However, those of skill in the art will recognize that many effective vaccine compositions cause some discomfort or relatively minor distress upon or after administration. However, the benefits of being protected against full-blown disease far outweigh this possibility. As an alternative, a heterotypic virus that does not naturally infect or that does not normally cause disease in the animal being vaccinated may be utilized. Examples of ways of inactivating the antigens of the present invention (e.g., the RPV of the present invention) include, but are not limited to, heat, formaldehyde, formalin, biethylene amine, radiation, and beta-propiolactone treatment.

Other suitable vaccine components, e.g. pharmacologically acceptable carriers, are well-known to those of skill in the art, as is the preparation of such compositions for use as vaccines. Typically, such compositions are prepared either as liquid solutions or suspensions, however solid forms such as tablets, pills, powders and the like are also contemplated. Solid forms suitable for solution in, or suspension in, liquids prior to administration, or for dissolution in the mouth, may also be prepared. The preparation may also be emulsified. The active ingredients may be mixed with excipients which are pharmaceutically acceptable and compatible with the active ingredients. Suitable excipients are, for example, water, saline, dextrose, glycerol, ethanol and the like, or combinations thereof. In addition, the composition may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, and the like. If it is desired to

administer an oral form of the composition, various thickeners, flavorings, diluents, emulsifiers, dispersing aids or binders and the like may be added. The composition of the present invention may contain any such additional ingredients so as to provide the composition in a form suitable for administration. 5 The final amount of the translatable nucleic acid in the formulations may vary. However, in general, the amount will be from about 1-99%. The compositions may further comprise an adjuvant, suitable examples of which include but are not limited to Seppic, Quil A, Alhydrogel, etc.

Generally, for administration as a vaccine, a nucleic acid will be included in a virus or virion particle and the entire particle will be a vaccine component. As will be understood by those of skill in the art, when the vaccine contains virions (such as the RPV of the present invention) they will be inac- 15 tivated (killed) or attenuated, i.e. obtained after several passages of the virus in cell culture so that particles do not cause symptoms of disease, or cause only mild, non-life threatening symptoms. Those of skill in the art will also recognize that other methods of attenuating viruses exist, and any of these 20 may be used in the practice of the present invention to develop a suitable form of the variant virus disclosed herein. For example, various mutations that interfere with virus toxicity, ability to reproduce, etc. may be introduced.

Alternatively, the nucleotide sequence presented in SEQ 25 ID NO: 1 (or segments thereof that encode antigenic regions or epitopes as described herein, for example SEQ ID NO: 2)) may be delivered in a heterologous vector such as a different virus (e.g. a virus that does not cause disease in the vaccine recipient species), or another type of construct that is 30 designed to result in expression of the encoded protein in the vaccine recipient, without causing symptoms of disease. Examples of such viruses include but are not limited to feline panleukopenia virus (FPV), various herpesviruses, nonpathogenic "orphan viruses", enteric viruses such as enterovi- 35 rus, etc. Other forms of the vaccine are also contemplated. For example, "empty" virion particle vaccines (without nucleic acid) are also contemplated, as are vaccines comprising proteins that are not assembled into a capsid. The RPVs of the invention may also be substantially or partially purified and/ 40 or concentrated by panning e.g. with transferrin coated beads in a column, in dishes, etc.

The RPVs of the invention may be safely administered at very high titers. For example, a usual dose of CPV-2 in a vaccine is about 10,000 virus particles per injection. The 45 amount administered when RPV is used ranges from about 10,000 particles per injection to about 100,000 particles per injection, or even higher, e.g. from about 100,000 up to about 1,000,000 particles per injection.

Alternatively, the vaccine preparation may include a VP-2 50 protein (polypeptide, peptide, etc.) sequence as set forth in SEQ ID NO: 3, or in SEQ ID MO: 4, or antigenic regions or epitopes thereof. Further, sequences which display at least about 50% identity, preferably about 60%, more preferably about 70, 80, or 90% identity or similarity, or even about 95, 55 96, 97, 98 or 99% or greater identity or similarity to SEQ ID NO: 3 or SEQ ID NO: 4 as described above, or antigenic regions or epitopes thereof are also contemplated for use in the vaccines, so long as the sequence includes one or both of amino acid residues 232 and 300 as Thr and Asp, respectively. 60 The terms "identity" and "similarity" are know in the art, where use of the term "identity" generally refers to a sequence comparison based on identical matches between corresponding identical positions in the sequences being compared.

The term "similarity" refers to a comparison between 65 amino acid sequences, and takes into account not only identical amino acids in corresponding positions, but also func12

tionally similar amino acids in corresponding positions (e.g. Asp and Glu, which both possess a side chain that is usually negatively charged at neutral pH, and thus may function similarly). Thus, similarity between polypeptide sequences indicates functional similarity, in addition to sequence similarity. The sequences encompassed by the invention may differ from those which are explicitly disclosed, for example, by having conservative amino acid substitutions, as are understood by those of skill in the art (e.g. positively charged amino acids may be substituted for one another, negatively charged amino acids may be substituted for one another, aliphatic amino acids may be substituted for one another, etc.), so long as the amino acid sequence retains the ability to elicit an effective, suitable immune response upon administration to a vaccine recipient. Further, the invention also encompasses amino acid sequences of antigenic regions or epitopes that are short contiguous sequences within SEQ ID NO: 3 (i.e. truncated or partial sequences which represent a portion of SEQ ID NO: 3) but which retain the ability to elicit an effective, suitable immune response against the development of symptoms of canine parvovirus infection in a vaccine recipient. Such antigenic regions are generally in the range of about 5-10 amino acids in length, but may be about 15, 20, 25, 30, 35, 40, 45 or even about 50 amino acids or more in length, and will generally encompass one or both of amino acid residues 232 and 300.

In one embodiment of the invention, the vaccine recipient is a puppy. By "puppy" we mean a young dog that is less than about 8 weeks of age. Typically, the vaccine is administered to a puppy at an age of e.g. 2-3 weeks, followed by booster doses at 4 and 6 weeks. In some embodiments, the vaccine is not used after 8 weeks, as after that time, vaccines directed to other known circulating viruses may then be employed without interference by maternal antibodies.

While in some embodiments, the vaccine recipient is a puppy, those of skill in the art will recognize that this need not always be the case. The vaccine may be administered to a dog of any age and to other species of animals (especially carnivores) that are susceptible to infection by parvoviruses, both adults and juvenile animals. Examples include but are not limited to canids (e.g. wild canids such as wolves, wild dog species, coyotes, foxes such as gray, kit and red, etc.], etc.), skunks, cats (including domestic cats and kittens, and larger species of cats, whether domesticated or wild such as bobcats, cougars, lion, tigers, etc.), mink, red panda, etc.; various procyonidae, including raccoons, coatis, kinkajous, olingos, ringtails and cacomistles, etc. Such animals may be domesticated (e.g. pets), "working" animals (e.g. service animals), livestock, animals in the wild, animals in captivity or on reserves, in zoos, in shelters, etc., so long as they can benefit by administration of the vaccine.

The immunogenic/vaccine preparations of the present invention may be administered by any of many suitable means which are well known to those of skill in the art, including but not limited to by injection, administered parenterally, for example, intramuscularly, subcutaneously, intraperitoneally, intradermally or the like, orally via a "puppy lollipop" as described above, intranasally, by ingestion of a food product containing the antigen, etc. However, in preferred a embodiment, the mode of administration to an adult recipient is by injection, and, as described above, the mode of injection to a juvenile is via a solid preparation that dissolves in the mouth upon sucking. The compositions of the invention may be administered alone or in combination with other medicaments or immunogenic compositions, e.g. as part of a multi-component vaccine. Further, administration may be a single event, or multiple booster doses may be

25

35

administered at various timed intervals to augment the immune response. In the case of administration to puppies, a typical administration regimen would be (as described above) initial administration at e.g. 2-3 weeks of age, followed by booster doses at 4 and 6 weeks. Preferably, administration is prophylactic, i.e. before exposure to the virus has occurred, or is suspected to have occurred, but may also be after the fact, i.e. after a known or suspected exposure, or therapeutically, e.g. after the occurrence of disease symptoms associated with viral infection.

The invention also provides methods of eliciting an immune response in a patient or individual in need thereof, as well as methods of vaccinating a patient or individual (especially a carnivore such as a juvenile canid, e.g. a puppy) against parvovirus infection. The methods include administering one or more doses of an immune eliciting preparation or a vaccine preparation to the recipient. The amount of the dose that is administered is sufficient to elicit an immune response, preferably a protective immune response, to the antigens encoded by the nucleic acids that are administered. Alternatively, in the case of administering one or more amino acid sequences as described herein, the immune response is to the antigens present in the amino acid sequences. Preferably, the immune response is protective and no disease symptoms will occur as a result of administration. However, those of skill in the art will recognize that many effective vaccine compositions cause some relatively minor symptoms upon administration, or attenuate without eliminating all symptoms of disease once the recipient is infected with a disease causing organism. However, the benefits of being protected against full-blown disease far outweigh these possibilities.

In other embodiments, the raccoon parvovirus of the invention is used as a gene delivery system. The raccoon parvovirus (RPV) described herein is an autonomously replicating parvovirus (ARP). The use of ARPs as gene delivery vehicles and expression vectors has been described (see U.S. Pat. No. 5,585,254 to Maxwell, et al.). Since the RPV of the invention is antigenically non-reactive with sera from dogs vaccinated with CPV or which have been naturally exposed to CPV, this RPV is well-suited for use as a gene vehicle and/or expression vector for canines, and also for other species (see below). Genetically engineered RPVs are used to carry into a cell of interest "foreign" or "heterologous" nucleic acid sequences, i.e. sequences which do not originate from RPV, but which encode and express within the cell of interest, or otherwise promote or facilitate expression of, a polypeptide or peptide of interest. For example, one or more immunogens may be encoded and the RPV may be administered to a subject in order to elicit an immune response to the one or more immunogens (e.g. to vaccinate the subject).

RPV is small, non-enveloped single-stand DNA-containing virus. Like other autonomous parvoviruses of vertebrates, the RPV genome has two open reading frames, ORF1 and ORF2 flanked by inverted terminal repeats (ITRs) which are required for DNA replication (FIG. 4A). Expression of ORF1 and OEF2 is driven by promoters P4 and P38, respectively (FIG. 4B). ORF1 encodes non-structural (NS) proteins NS1 and NS2, and ORF2 encodes capsid proteins VP-1, VP-2 and VP-3 (FIG. 4C). NS1 is a multifunctional protein that is necessary for viral DNA replication and promoter trans-activation, such as that which drives transcription of VP-1 and VP-2. NS2 is essential for replication, virus production and nuclear egress of progeny virions. Parvovirus particles have icosahedral symmetry and are about 20 nm in diameter. The particles contain about 50% protein and about 50% DNA, with neutralizing epitopes being present on the exposed surface of the particles.

It is possible to insert heterologous sequences within canine parvovirus capsid proteins, e.g. the VP-2 protein, and the same or similar strategies are used to genetically engineer the RPV of the present invention. (Austin et al., Tropic determinant for canine parvovirus and feline panleukopenia virus functions through the capsid protein VP-2, Journal of General Virology (1997), 78, 925-928.) Table 1 lists restriction sites located within the gene encoding VP-2. One or more of these restriction sites may be used as sites for insertion of foreign DNA.

TABLE 1

		R	estrictio	on map	of VP-2	2 of Rac	coon pa	arvovin	15.		
Enzyme	Number of Cuts			[В	lase 5' to	o Cleav	age Site	es]	
AatII	0										
AccI	3	825	1048	1149							
AclI	0										
AcyI	1	1045									
AfIII	0										
AfIIII	0										
AgeI	0										
AhaIII	5	159	218	455	1065	1611					
AluI	5	64	571	988	1006	1615					
AlwNI	2	73	472								
ApaBI	0										
ApaI	0										
ApaLI	1	1022									
ApoI	9	154	160	437	747	784	879	926	1409	1423	
AscI	0										
AsuI	2	1057	1360								
AsuII	0										
AvaI	0										
AvaII	1	1360									
AvrII	0										
BalI	0										
BamHI	0										
BclI	0										
BetI	0										
BglI	0										
BglII	1	577									

US 8,734,808 B2

		R	estrictio	on map	of VP-2	of Rac	coon pa	arvoviru	ıs.			
Enzyme	Number of Cuts			[В	ase 5' t	o Cleav	age Site	s]		
BsaAI	0											
BsaBI	2	1109	1127									
BsePI	2	493	1592									
BsiYI	2	34	1315									
Bsp1407I	1	210										
BspHI	0											
BspLUIII	0											
BstEII	0											
BstXI	2	380	626									
Cac8I	0											
Cauli Cfr10I	0											
CfrI	ŏ											
ClaI	0											
CviJI CviBI	10	64 277	295	483	571	863	988 520	1006	1059	1615	1627	1467
CVINI	12	1546	515	540	414	517	552	1024	1078	1141	1572	1402
DdeI	7	479	581	769	1007	1190	1578	1621				
DpnI	6	72	579	716	1130	1307	1538					
Drall Drall	0											
DrdI	1	1157										
DsaI	3	601	625	705								
Eam1105I	0											
EcoNI EcoNI	0											
EcoRI	1	926										
EcoRII	2	372	1488									
EcoRV	1	1231										
Esp1 Fnu4HI	3	569	1079	1118								
FnuDII	0											
FseI	0											
Hael	0											
HaeIII	1	1059										
HgiAI	1	1026										
HgiCI	0											
Hgijii Hhal	2	1098	1515									
HineII	2	40	282									
HindIII	0											
Hinfi Hnal	3	469 282	1248	1641								
HpaII	0	202										
KpnI	0											
MaeI	4	680	1738	1745	1799							
MaeIII	1	945 321	951	1569								
MboI	6	70	577	714	1128	1305	1536					
McrI	0											
MfeI	1	1310										
MseI	22	158	217	281	368	395	443	454	511	658	1064	1259
		1265	1331	1349	1366	1375	1438	1522	1610	1664	1682	
		1758										
Msll MetI	2	837	1457									
MwoI	0											
NaeI	0											
NarI	0	604	69.5	705								
NCOI NdeI	3	601	625	705								
NheI	0											
NlaIII	9	294	313	605	629	709	824	842	1159	1456		
NlaIV Nati	0											
NUI NUI	0											
NspBII	1	1006										
NspI	0											
PacI	0	24										
PmaCI	1 0	34										

US 8,734,808 B2

17

	NT 1											
Enzyme	Number of Cuts			[В	ase 5' te	o Cleava	age Site	s]		
PmeI	0											
PpuMI	0											
PshAI	1	35										
PstI	1	279										
PvuI	0											
Pvull	1	1006										
Rsal	6	135	212	711	761	1385	1691					
RsrII	0											
Sacl	0											
Sach	0											
Sall	0											
SanDi	0											
Saul	0											
Scar	2	274	1400									
SCIFI	2	374	1490									
Saul	1	2020	601	625	705							
Seci	4	328	001	023	705							
SEXAI	3	65	275	701								
SIEI	3	03	213	/91								
SIII	0											
SgrAI	0											
Smal	0											
Smill	1	108										
SnaBI	0	198										
SnaDi	1	670										
Sphi	0	079										
Spll	0											
SrfI	Ő											
Sse8387I	Ő											
Sse86471	Ő											
SsnI	ž	1257	1701									
Stul	õ	1237	1701									
Styl	4	328	601	625	705							
Swal	2	159	218	020	,							
TagI	õ	105	210									
TatI	2	210	759									
TfiI	3	469	1248	1641								
TseI	3	568	1078	1117								
sp45I	0											
Tsp4CI	2	34	462									
TspEI	25	154	160	237	261	317	389	437	730	747	784	879
1		926	1310	1328	1409	1423	1482	1495	1518	1565	1600	
		1646	1678	1715	1750							
TspRI	0											
Tth111I	0											
VspI	2	658	1664									
XbaI	0											
XcmI	1	1241										
XhoI	0											
XhoII	4	70	577	714	1305							
XmaIII	0											
XmnI	0											

TABLE 1-continued

Restriction sites that will be particularly useful for inserting nuclei acid sequences into the VP-2 gene include EcoRI, HpaI, and PstI. This is because these restriction sites are commonly engineered in the multiple cloning sites of most 55 recombinant vectors.

Autonomous parvoviruses are generally limited to packaging a total of amount of DNA equal to about 105% of their wild DNA content. Thus, preferred transgenes or portions thereof (i.e. heterologous or foreign nucleotide sequences, 60 nucleic acid sequences which do not naturally occur in parvoviruses, etc.) for inclusion in the RPV gene delivery vehicles of the invention are generally from about 9 to about 60 nucleotides in length. For example, sequences of about 60 nucleotides or less (e.g. about 10, 15, 20, 25, 30, 35, 40, 45, 50 or 55 nucleotides) can be readily accommodated. Such sequences generally encode sequences of interest (e.g. small

proteins, polypeptides, peptides, etc.) comprising from about 3 to about 20 amino acids e.g. about 3, 4, 5, 6, 7, 8, 9, 19, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 amino acids. In one embodiment, peptides up to 12 amino acids encoded by 36 nucleotides are accommodated by the RPV. However, in other embodiments, significantly more nucleic acid sequences are inserted by replacing portions of the VP-2 protein by the foreign DNA. For example, in some embodiments, from about 50 to 550 amino acids are replaced, e.g. about 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, or even 550 (the entire VP-2 gene) are replaced by foreign DNA, with retention or addition of the required inverted repeats.

Sequences of interest encoded by the heterologous nucleotide sequences may include, for example, one or more epitopes or antigenic regions of interest from a protein of interest, examples of which include but are not limited to:

65

surface proteins of important pathogens of dogs, cats and other species; cassettes of linear epitopes of canine distemper virus, canine adenovirus and/or canine influenza virus surface protein; multiple peptides selected from hemagglutinin of canine distemper virus and canine influenza virus; the ami- 5 nogenic epitope of canine parvovirus; the amino epitope of canine adenovirus (derived from the penton protein), etc. Additional epitopes of some important canine viruses have been defined. (See, for example, Sugai et al, *Microbiol Immunol.* 53(12):667-74, 2009; Jung et al, *J Vet Sci.* 6(1):21-4, 10 2005; Ghosh et al, *Immunology* 104(1):58-66, 2001; etc.).

In other embodiments, the encoded polypeptide/peptide has a function or properties other than (or in addition to) that of eliciting an immune response, e.g. cytotoxicity to tumor cells, targeting properties, etc.

In some embodiments of the invention, the RVP gene delivery vehicle may be further genetically engineered to permit targeting of the RVP to a cell type or cell types of interest. Like other carnivore parvoviruses (and also rodent arena viruses), raccoon parvoviruses enter cells by binding to cell 20 surface transferrin receptors (TFR), as illustrated schematically in FIG. 5A. By manipulating and mutating the cognate transferrin binding sites of RPV (anti-receptor sites), (e.g. proteins VP-1, VP-2 and/or VP-3), it is possible to alter the host range of the virus and/or the cell type which the RPV can 25 infect. For example, such genetically engineered RPV are able to bind to transferrin receptors and infect cells of species other than raccoon (including humans), and can thus be used as gene delivery vehicles in species other than canines. Alternatively, RPVs with altered host ranges can be identified and 30 selected during passage of the virus in cells from a variety of species. There are generally two ways to achieve targeting specificity, transductional targeting and transcriptional targeting. Transductional targeting involves the selective uptake of the vector into the cells of interest, for example, dividing 35 cells of canine cancers, intestinal cells, etc. This may occur by genetically modifying the virus to contain and express factors necessary for binding to and infecting a cell of interest, e.g. ligands (e.g. cognate ligands, which may be proteins) which bind receptors located exclusively (or almost exclusively) on 40 the surface of the cell of interest, and which cause or permit uptake of the virus by the cell of interest. For example, transductional targeting with an RPV is generally carried out by genetically engineering the RPV to express on its outer surface a cognate ligand of a receptor type that is found only on 45 the targeted cell. In transcriptional targeting, although the transgene may be taken by many different host cells it is transcribed only in target cells. Transcriptional targeting with an RPV is generally carried out by including a cell specific and/or inducible promoter in the construct to drive expression 50 of the heterologous sequences of interest only (or almost exclusively) in the targeted cells.

A general strategy for preparing autonomous parvovirus vectors has been described by Maxwell et al., (Methods 28 (2002) 168-181), and RPV viral vector production can be 55 adapted as described therein, e.g. similarly to production of the LuIII vector. Briefly, a productive cell line such as CRFK is used. Producer cells (such as the 324K cell line) are transfected with supercoiled forms of a recombinant RPV plasmid containing transgene sequences of interest, together with one 60 or more plasmids expressing the required helper functions for excision, amplification and packaging of the virus particles e.g. NS1, NS2, VP1, VP-2, etc. This process is illustrated schematically in FIG. **5**B. Strategies for making recombinant plasmids for canine and feline parvoviruses have been 65 described (Austin et al., *Journal of General Virology* (1996), 77, 1787-1792) and the same or similar strategies are used to

20

make recombinant RPV plasmids. The RPV vectors are harvested from cell extracts prepared several days after transfection.

Virotherapy for tumors is now being explored (see, for example, Rommelaere et al, Cytokine & Growth Factor Reviews 21:185-195, 2010; and U.S. Pat. No. 7,179,456 to Rommelaere et al, the complete contents of which are herein incorporated by reference). However, there is a need for novel viruses that can carry out the targeted attacks. The RPV of the invention can be used in this manner, i.e. as tumor therapy agent. The use of the RVP in this manner generally involves the identification of a subject with a tumor in which the RVP is able to replicate, and administration of the RVP to the subject that bears the tumor. Administration may be carried out in any suitable manner, including but not limited to intravenously, by injection directly into the tumor (i.e. intratumoral administration), etc. RPV virotherapy is mainly intended for a single-administration (e.g. "one-shot") treatment. This is because once the subject (e.g. a dog) receives the RPV, it will respond immunologically by creating antibodies to the RPV. Thus, the possibility of giving an effective second shot is less likely. Thus, the one dose that is administered has a very high parvovirus concentration. For example, the dose will range from at least about 10 million to 100 million virus particles per dose per dose. Only RPV can be administered in an adult dog to cause no diarrhea and kill residual tumor cells in a diffuse tumor. Only RPV (but not, for example, CPV-2 or FPLV) can be used at these very high doses and yet cause no clinical disease in the recipient, i.e. RPV will not cause diarrhea in a tumor bearing dog or cat.

Subjects with tumors which may be treated in this manner include but are not limited to, for example, dogs, cats, pet minks, raccoons and humans, the therapy being highly efficacious for dogs and cats. The subject may be a juvenile or an adult. Types of tumors which may be treated with the RPV of the invention include but are not limited to: for dogs, histiocytic sarcoma (which are diffuse and for which no successful treatment is currently available), pancreatic cancers, lymphomas, gliomas, and osteosarcoma, etc.; for cats, the tumors listed above and also hepatobiliary tumors, etc.

In one embodiment, the RPV that is used to treat the tumor is genetically engineered to specifically target the tumor cells so that viral binding to the tumor cells is selective or, preferably, specific. This use of the RVP of the invention may also be carried out, for example, in adult dogs, since even if they have been previously vaccinated against CPV, the antibodies they carry will not fully neutralize the RPV of the invention.

The invention also provides diagnostic kits for the detection of the raccoon parvoviruses described herein. Such kits may include oligonucleotide primers specific for amplifying (e.g. by polymerase chain reaction) the nucleic acid sequences disclosed herein. Alternatively, such kits may include antibodies (e.g. monoclonal or polyclonal) that bind selectively or specifically to unique antigenic determinants displayed by the parvoviruses, e.g. for Asp at position 300 of the VP-2 protein. Methods of detecting the RPVs are also encompassed. Such methods generally involve obtaining a biological sample from a subject suspected of being infected with an RPV of the invention, and detecting the presence or absence of an RPV of the invention in the sample, e.g. using oligonucleotide primers specific for amplifying (e.g. by polymerase chain reaction) the nucleic acid sequences disclosed herein, and/or by using antibodies specific for the RPVs disclosed herein. Methods of diagnosing infection of an animal with the RPVs (e.g. MoAb that react with RPV and raccoon kidney cell culture) using these methods are also provided.

The foregoing examples serve to illustrate the invention but should not be construed as limiting in any way.

EXAMPLES

Example 1

Isolation of a Novel Parvovirus from a Raccoon

The common North American raccoon (Procvon lotor) is a nocturnal omnivore that has adapted well to urban habitats. Raccoons prefer to live near river banks that provide shelter and food; in urban setting they may interact with dogs and cats. Raccoons are susceptible to some diseases that affect carnivores, especially canine distemper, rabies, canine adenovirus, leptospirosis, influenza A and parvovirus. There are previous reports of the detection and characterization of parvovirus from raccoons in Canada (Barker et al., 1983) and the USA (Nettles et al., 1980); however, there are no published reports in the past two decades. It has also been reported that raccoons are not susceptible to canine parvovirus type 2 (CPV-2) (Appel and Parrish, 1982). However, as disclosed 20 herein evidence is provided that suggests that this is not the case. Moreover, the present disclosure describes the isolation of a parvovirus genetically related to CPV-2 in a rescued raccoon at a wildlife rehabilitation facility in the USA.

A two-month-old male raccoon was admitted to wildlife 25 rehabilitation in Arkansas, USA. The raccoon was clinically normal at the time of admission. After 4 weeks, the raccoon developed loss of appetite and severe diarrhea. The raccoon was treated with enrofloxacin (Baytril, Bayer) administered once a day, and fluids were administered subcutaneous. 30 Despite this treatment, the animal died. Two other raccoons had previously died in the same facility one after showing similar clinical signs. The raccoon was submitted to the Oklahoma Animal Disease Laboratory for postmortem examination. 35

On gross examination, the oral mucosa and sclera were pale to white, the abdomen was distended and the perineum and anal areas were covered with thin 'runny' yellow fluid. The animal weighed 1.5 kg and had adequate skeletal muscle and body fat stores, consistent with acute death. The stomach 40 contents were mucoid yellow with yellow granular material. The small intestine was dilated and red, with rough serosa. The contents of both the small and large intestines were runny, mucoid and yellowish, with yellow material coating a smooth glassy mucosa. 45

Histopathological examination revealed that the spleen was congested and the follicles were depleted of lymphocytes with lymphoid necrosis. Diffuse pulmonary edema was evident in the lungs. Sections of small intestine had widespread crypt necrosis with sloughing of the villi and collapse of the 50 remaining lamina propria. A few areas revealed scattered, large, hyperchromatic regenerating crypt epithelial cells. The intestinal luminal surface was coated with fibrin, neutrophils, bacteria and some hemorrhage. Immunohistochemistry for CPV revealed rare positive signals, which were evaluated as 55 equivocal. The microscopic appearance of the intestine was consistent with viral enteritis.

Fresh sections of the intestine and tongue were examined by the fluorescent antibody test with monoclonal antibody 3B10 (fluorescein isothiocynate-labeled anti-CPV conjugate; 60 MVRD) (Kapil et al., 2007). Both tissues were found to be negative. Based on the histopathological findings, there were few enterocytes remaining in the intestine that could have contained virus. The general PCR for carnivore parvovirus, as developed by Desario et al., (2005), was carried out twice on 65 the intestinal contents, and was strongly positive both times. To further support the virology findings, the intestinal con-

tents were inoculated onto cultured Crandall-Reese feline kidney (CRFK) cells (ATCC).

For virus isolation, the sample of intestine was prepared as a 10 percent w/v suspension and freeze-thawed once. The suspension was extracted with an equal volume of chloroform. After centrifugation at 10,000 g for five minutes, the supernatant was filtered through a 0.22 µm syringe filter. The filtrate was inoculated onto CRFK cells approximately 45 minutes after plating in the flasks. The cells were observed daily for 5 days, and on the fifth day cytopathology characterized by rounding and detachment of cells was observed. The cells were freeze-thawed twice and subjected to a hemagglutination test with porcine erythrocytes. The sample showed hemagglutinating activity, that parvovirus had replicated in the cell culture. The presence of the virus was further confirmed by the general parvovirus PCR (Desario et al., 2005) on the cell culture supernatant, followed by sequencing of the amplicon.

Three independent sequences were obtained from the same specimen (AR-08071304). All the sequences were the same. Two were derived from the fresh sample of intestinal contents, and the third was derived from the isolate cultured in CRFK cells. The three sequences were subjected to BLASTN analysis and CLUSTALW analyses (National Center for Biotechnology Information). Based on sequence analysis, the raccoon parvovirus was found to be genetically most closely related to CP-V2.

To completely sequence the viral protein-2 (VP-2) of the raccoon parvovirus, two additional PCR amplifications were performed as described by Decaro et al., (2008). The complete VP-2 sequence of the raccoon parvovirus was subjected to BLAST analysis. The sequence had highest identity with CPV (98 percent); and had an identity of 97 percent with isolates of feline panleukopaenia virus, another raccoon parsovirus (M24005), and mink enteritis virus (FIG. **6**).

In experimental studies, raccoons have been found to be susceptible to mink enteritis virus and feline parvovirus (Barker et al., 1983). The raccoon isolate described here had biological and sequence homology with CPV-2. Approximately 500 base pairs of the partial sequence of VP-2 had highest homology (99 percent) with CPV-2. The codons at positions 426 and 555 helped classification of the raccoon isolate as CPV-2 (Desario et al., 2005). Moreover, codons 564 and 568 were also consistent with CPV-2 (Truyen et al., 1994). Codon 564 in the raccoon isolate was AGT, as it is in canine parvovirus; this codon 564 is AAT in feline parvovirus and the previous raccoon isolate (M24005). Codon 568 is GGT in both CPV-2 and the present raccoon isolate; however, it is GCT in feline parvovirus and the previous raccoon isolate (M24005).

Similarly, based on the amino acid sequence of VP-2, the raccoon parvovirus may be considered to be a unique variant of CPV-2. At amino acid position 80 of VP-2, the raccoon isolate had arginine, the same as VP-2 from CPV isolates; in contrast, feline parvovirus has lysine at amino acid position 80. At amino acid position 87, the raccoon isolate had a leucine residue, like the variants of CPV-2, CPV-2a and CPV-2b; both FPV and CPV-2 have methionine at amino acid position 87. At amino acid position 93, the raccoon isolate had asparagine, like all CPVs, while FPV has lysine at this amino acid position. At amino acid position 101, the raccoon isolate had threonine, similar to CPV-2a and CPV-2b; both FPV and CPV-2 had isoleucine at amino acid position 101. At amino acid position 103, the raccoon parvovirus had alanine, like CPV-2a and CPV-2b, whereas FPV and CPV-2 have valine at amino acid position 103. The raccoon parvovirus had threonine at amino acid position 232, serine at amino acid position

297 and aspartic acid at amino acid position 300; FPV and CPV-2 have alanine at amino acid position 300, while CPV-2a and CPV-2b have glycine at amino acid position 300. At amino acid position 305, the raccoon isolate had aspartic acid. like FPV and CPV-2; CPV-2a and CPV-2b have tyrosine at this amino acid position. The raccoon parvovirus had asparagine at amino acid position 426, like FPV, CPV-2, and CPV-2a. At amino acid position 555, the raccoon isolate had valine, like most CPV genotypes and FPV; however, CPV-2a has isoleucine at amino acid position 555. At amino acid position 564, the raccoon isolate had serine, like all CPV isolates; FPV has asparagine at this amino acid position. At amino acid position 568, the raccoon parvovirus had glycine, like CPV-2; FPV has alanine at this amino acid position. Thus, based on the complete VP-2 sequence of the raccoon parvovirus isolate (CAR-08071304), it was considered to be a unique genetic variant of CPV-2. A summary of these differences is shown in FIG. 7.

Notably, amino acid residue 232 is Thr in the raccoon isolate and amino acid residue 300 is Asp. These two amino acid residues in the VP-2 protein of RPV are thus distinct from the VP-2 proteins of all other known CPV isolates and may be responsible for the unique properties (e.g. antibody reactivity) of this RPV. In particular, substitution of amino acid residue 300 from Gly (uncharged, side chain=H) to Asp (negative charge at neutral pH, side chain=CH₂COO⁻) alters the antigenic properties of the virus. Without being bound by theory, it is likely that this change also affects the receptor binding properties of the virus.

The previous raccoon isolate M24005 was isolated in the USA in 1979. The present isolate was antigenically different from current circulating CPV-2 isolates in USA, based on its lack of reactivity with the monoclonal antibody A3B10

24

Due to the likely interactions of raccoons with companion animals in urban areas of the USA, this finding is epidemiologically significant. Based on a published survey from Canada (Barker et al., 1983), raccoons may be frequently exposed to CPV-2. In the study of Barker et al., (1983), 25 of out of 112 raccoons (22.4 percent) tested in Canada were seropositive; with most animals positive at hemagglutinationinhibition titres equal to or greater than 1:256. Raccoons have also been implicated in the spillover of FPV to zoo animals in urban environments (Junge et al., 2007). Raccoons may be exposed to CPV-2 due to their omnivorous nature and feeding habits. The role of urban raccoons in the transmission of CPV-2 to other species requires further investigation. There is no published evidence of natural CPV-2 infection in raccoons. However, based on a hemagglutination assay, CPV-2 was implicated in a captive raccoon in the USA based upon hemagglutination of porcine erythrocytes at pH 7.2 (Nettles et al., 1980); the genome sequence was not determined in that study.

Example 2

Reactivity of Carnivore Parvoviruses with Commercially Available Monoclonal Antibodies

Testing of the new isolate was carried out using four additional commercially available antibodies, CPV1-24, CPV1-13 and CPV1-5 (Custom Monoclonal Antibodies, Sacramento, Calif.). These canine parvovirus-2 antibodies are commercially available for routine diagnosis of canine parvovirus-2. The results are presented in Table 2. As can be seen, the antibodies react similarly with CPV, FPLV and RPV. This shows that some immunogenic epitopes are conserved among these viruses.

TABLE 2

	Antibody te	sting of three a	<u>dditional r</u>	accoon isola	tes	
Isolate description			C	PV Monocle	nal Antibod	ies
440 Codon		Assigned #	CPV1-2	CPV1-24	CPV1-13	CPV1-5
GCA	CPV-2c	08060974	_	-	±	+
GCA	CPV-2c	08090386	-	-	±	+
ACA	CPV-2c	07030244	-	-	±	+
ACA	CPV-2c	07080797	-	-	±	+
ACA	CPV-2c	07051347	-	-	+	+
	CPV-2b	08070185	-	-	±	+
	CPV-2b	07030847	-	-	±	+
	FPV	08120386	-	-	±	±
	FPV	08061339	-	-	-	-
	RPV, pH 7.2	08071304	-	-	±	+
	RPV, pH 6.5	08071304	-	-	±	+

(Kapil et al., 2007). Virus isolation-positive tissue was negative by direct fluorescent antibody testing. On the basis of the 55 histological findings, few enterocytes were present lining the intestines when the raccoon died. The lack of reactivity with A3B10 was not surprising, because this monoclonal antibody reacts with a major antigenic domain of CPV-2 (Wikoff et al., 1994). The intestinal contents were negative for parasitic ova 60 by direct examination and fecal floatation test. The intestinal contents were negative for parasitic ova by direct examination and the faecal flotation test, and negative on ELISA for Clostridium difficile toxin. Streptococcus dysgalatiae subspecies equisimilis and Streptococcus uberis were isolated from a sample of lymph node by routine bacteriological examina- 65 tion. Secondary bacterial contaminants or their toxins may have contributed to the death of the raccoon.

Example 3

Reactivity of Three Other Raccoon Parvoviruses with a Panel of Monoclonal Antibodies

In addition to the Arkansas raccoon parvovirus 08071304 described above, three additional parvoviruses of raccoon origin but from Illinois were tested with respect to reactivity with the 3B10 MoAb (VMRD, WA) using fluorescent antibody testing. 3B10 MoAb is specific for CPV-2 and is frequently used to identify and/or diagnose parvovirus infection in dogs. All three viruses tested negative with 3B10 MoAb. Thus, as was the case for the RPV described in Examples 1 and 2, no epitopes of these three additional raccoon viruses were recognized by this CPV-2 MoAb antibody.

These results demonstrate that these three parvoviruses, like RPV, also have a mutation at amino acid position 300 of VP-2, since these intestinal tissue samples did not react with the 3B10 MoAb, which is known to react with an epitope that includes the amino acid at position 300 in other parvoviruses. ⁵

Example 4

Sequencing of Gene Encoding The VP-2 Proteins of the Three Additional Raccoon Parvoviruses

The gene encoding the VP-2 protein of each of the three viruses described in Example 3 was partially sequenced and 15 GTTGCGCCTAATTATTGTCAAAATGAATGAATGAATGATCTGGTCAATTATTGTAAAA

Inverse-Complement for 10071191-A, nucleotide positions 1216 to 1773 (i.e. 557 nucleotides in length):

(SEQ ID NO: 5) TCAGGAAGATATCCAGAAGGAAGTGGATTGGATTCAAAATATTAACTTTAA CCTTCCTGTAACAAATGATAATGTATTGCTACCAACAGATCCAATTG GAGGTAAAACAGGAATTAACTATACTAATATATTTAATACTTATGGT CCTTTAACTGCATTAAATAATGTACCACCAGTTTATCCAAATGGTCA AATTTGGGATAAAGAATTTGATACTGACCACCAGTTTATCCAAATGGTCA AATTTGGGATAAAGAATTGTCACAAAATAGATCCAGGACTTCATG TAAATGCACCATTTGTTGTCAAAATAATTGTCCTGGTCAATTATTT GTAAAAGTTGCGCCTAATTTAACAAATGAATATGATCCTGATGCATC TGCTAATATGTCAAGAATTGTAACAAATGAATATGATCCTGATGCAACT GTAAAATTAGTATTTAAAGCTAAACTAAGAGCCTCTCATACTTGGAAAG GTAAATTAGTATTTAAAGGTAAACTAAGGACAAATTTAACTATGT CCCAATTCAACAAATGAGTATTAATGTAGAAAAATCTCAACT ACCAAGTAATATTGGAGGTATGAAAATTGTATAGAAAAAATCTCAACT AGCACCTAGAAAATTATTTAACAACTTAGCTAGTCTTAT

Inverse-Complement for 10071191-B, nucleotide positions 1214 to 1779 (565 nucleotides in length):

26

Inverse-Complement for 10071191-C, nucleotide positions 1216 to 1774 (556 nucleotides in length):

 5
 (SEQ ID NO: 7)

 TCAGGAAGATATCCAGAAGGAGATTGGATTGAAAATATTAACTTTAAC
 7)

 GGTAAAACAGGAATTAACTATAGTATTGCTACCAACAGATCCAAATGGA
 6

 10
 TTAACTGCATTAAATAATGTACCAACAGACCAAATGGTCCAA
 7

 11
 TGGGATAAAGAATTTGATACTGACCAACAGACCTAGTTAAAT
 7

 12
 GCACCATTTGTTTGCAAAATGATCGACCAGGTTTATCCAAAATGGTCAAATT
 7

 13
 GCACCATTTGTTTGTCAAAATAATTGTCCTGGTCAATTATTGTAAAA
 7

 14
 GTTGCGCCTAATTTAACAAATGAATGATCGTGGTCAATTATTGTAAAAA
 7

 15
 GTTGCGCCTAATTTAACAAATGAATGATCTGAGATGCAATCTGCAATTAA
 7

 16
 GTTGCGACCTAATTGTAACTAACGAATTATGATCCAGGAAAGGTAAATTA
 7

 17
 GTATTTAAAGAAATGTAACTAAGAGCCTCTCATACTGGGAAAGGTAAATTA
 7

 17
 GTATTTAAAGGTAAACTAAGAGCCTCTCATACTTGGAAACCAATTCGAA
 7

 17
 GTATTTAAAGGTAAACTAAGAATTGTAACCAATTTAGAACAAGGAAAGGTAAATTA
 7

 17
 GTATTTAAAGGTAAACTAAGAGCCTCTCATACTGGAAAGGTAAATTA
 7

 17
 GTATTTAAAGGTAAACTAAGAGACCAATTTAGAACAAGGAAAGGTAAATTA
 7

 17
 GTATTTAAATGTAAGTAAACTAAGAAGCCAATTTAGGAAAGGTAAATTA
 7

 17
 GTATTTAAAGGTAAACTAAGAGACTAAGGACCAATTTAACTAAGGAAAGGTAAATTA
 7

 17
 GTATTTAAAGGAAATTGTAAGAAAATTGAACCAATTTAACTAAGGAACAAGGTAAAA

ATTATATTAACAACTTAGCTAGGCGTAT

These are partial VP-2 sequences of RPV from Chicago, 25 Ill. Thus, amino acid residue 300 was not sequenced. However, the lack of reactivity with 3B10 MoAb (see Example 3) indirectly supports that the mutation at amino acid position 300 is also present in these viruses. Alignment of the four sequences (namely the full length 08071304 with the three 30 partial sequences 10071191-1,2,3) and revealed some differences. The first significant change is that at nucleotide 1216, of 08071304 has an "A", whereas all three newly identified RPVs (10071191A, B and C) have a "T". All four sequences are identical from nucleotides 1217 to 1261. However, at ³⁵ nucleotide position 1262 of 10071191-B, there is a "G" instead of an "A" which is present in the other 3 sequences. From nucleotides 1263 through 1268, all the sequences are identical. However, at nucleotide position 1269, sequence 10071191-B has a "G" instead of a "T" which is present in the $_{40}$ other three sequences. Similarly, at nucleotide positions 1272 and 1273, 10071191-B has "GG" instead of "AA" which is present in the other three sequences. From nucleotides 1274 through 1292 all four sequences are identical. However, at nucleotide position 1293 all sequences have an "A" but 10071191-B has a "G". From nucleotide 1294 through 1300 45 all sequences are identical, but then at nucleotide position 1301, 10071191-B has a "G" rather than an "A". From nucleotide 1302 through 1349 all sequences are identical, but at nucleotide position 1350 all sequences have a "T" but 10071191-2 has a "G". From nucleotide position 1351 all 50 sequences are identical up to nucleotide position 1706.

Based on this we conclude that one isolate, 10071191-B, has a base point mutation to "G" in several locations of the VP-2 gene.

Experiments conducted with respect to culturing these 3 55 new RPVs showed that 10071199-A can be propagated in cell culture, and that 10071199-C is highly cytopathic in the CRFK cell line.

Example 5

Phylogenetic Comparison of the New Raccoon Parvovirus Isolates 0808294, 10071191-1, 10071191-2 and 10071191-3 with Other Known Carnivore Parvoviruses

65

60

To construct a more extensive phylogenetic tree of carnivore parvoviruses, the full length sequence encoding the

VP-2 protein of new raccoon parvovirus isolate 0808294, the partial sequences encoding the VP-2 protein of raccoon isolates 10071191-1, 10071191-2 and 10071191-3 VP-2, and several other CPV-2 parvovirus sequences (available from the National Center for Biotechnology Information (NCBI) web-⁵ site) were compared. All sequences were first converted to FASTA formats and then aligned using the CLUSTAL/W program of the MEGA 4.1 software. The tree was constructed using Bootstrap analysis and the hierarchical clustering method UPGMA (Unweighted Pair Group Method with ¹⁰ Arithmetic Mean).

The results are depicted in FIG. **8**, which shows the NCBI GI numbers of each isolate. As can be seen, the Arkansas RPV (08071304) clusters with the well characterized canine parvoviruses. Thus, evolutionarily it is a canine parvovirus-2 like ¹⁵ virus.

Example 6

Basic Local Alignment Search Tool (BLAST) Analysis of Full Length 08071304 RPV VP-2 Sequence

The 1801 nucleotide sequence of 08071304 RPVVP-2 was analyzed using BLAST analysis. The results showed that this ²⁵ sequence exhibits high homology with other carnivore parvoviruses.

The first 7 highest identities (99%) were with canine parvoviruses. Surprisingly several 98% identities were obtained with feline panluekopaenia. There was 98% identity score 30 with a raccoon parvovirus (M24005.1) and there was also 98% identity with mink enteritis virus. This is unusual for parvovirus VP-2 sequences, in that such high levels of identity are not usually observed amongst several different carnivore parvoviruses. Normally, a canine parvovirus aligns and 35 shows high identity only with other canine parvoviruses, feline panleukopaenia shows the highest identity with other feline panleukopaenia viruses, etc. It thus appears that 08071304 is a chimeric carnivore parvovirus. This observation is further supported by the amino acid comparisons 40 shown in FIG. 7. In the past, carnivore parvoviruses were easy to classify based on the amino acid sequence of VP-2. Interestingly, the RPV (08071304) of the invention does not follow the linear VP-2 DNA convention and rules. At some places, as shown in FIG. 7, the amino acid that is at a critical 45 epitope is like that of FPV or MEV (e.g. at amino acid position 305). However, at amino acid position 93, it is like CPV. Thus, 08071307 RPV is unique genetically, antigenically and with respect to amino acid sequence. This virus would likely provide a common carnivore parvovirus for overcoming 50 maternal derived immunity in several species.

REFERENCES

- APPEL, M. J. & C. R. PARRISH. (1982). Raccoons are not 55 susceptible to canine parvovirus. Journal of American Veterinary Medical Association 18, 489.
- AUSTIN et al. (1997). Tropic determinant for canine parvovirus and feline panleukopenia virus functions through the capsid protein VP-2, Journal of General Virology 78, 925- 60 928.
- BARKER, I. K., POVEY, R. C., & D. R. Vogt. (1983). Response of mink, skunk, red fox, and raccoon to inoculation with mink virus enteritis, feline panleukopenia and canine parvovirus and prevalence of antibody to parvovirus 65 in wild carnivores in Ontario. Canadian Journal of Comparative Medicine 47, 188-197.

- DECARO, N., DESARIO, C., MICCOLUPO, A., CAM-POLO, M., PARISI, A., MARTELLA, V., AMOR ISCO, F., LUCENTE, M. S., LAVAZZA, A., AND BUONAVOGLIA, C. 2008. Genetic analysis of feline panleukopaenia viruses from cats with gasteroenteritis. Journal of General Virology 89, 2290-2298.
- DESARIO, C., DECARO, N., CAMPOLO, M., CAVALLI, A., CIRONE, F., ELIA, G, MARTELLA, V., LORUSSO, E., CAMERO, M., & BUONAVOGLIA, C. 2005. Canine parvovirus infection: which diagnostic test for virus? Journal of Virological Methods 126, 179-185.
- GHOSH et al., (2001). Identification of canine helper T-cell epitopes from the fusion protein of canine distemper virus, Immunology 104(1):58-66.
- JUNG et al, (2005). Induction of castration by immunization of male dogs with recombinant gonadotropin-releasing hormone (GnRH)-Canine distemper virus (CDV) T helper cell epitope p35, J Vet Sci. 6(1):21-4
- JUNGE, R. E., BAUMAN, K., KING, M., & GOMPPER, M. E. (2007). Serologic assessment of exposure to viral pathogens and *Leptospira* in an urban raccoon (*Procyon lotor*) population inhabiting a large zoological park. Journal of Zoological Wildlife Medicine 38, 18-26.
- KAPIL, S., COOPER, E., LAMM, C., MURRAY, B., REZA-BEK, G., JOHNSTON III, L., CAMPBELL, G., & JOHNSON, B. (2007). Canine parvovirus types 2c and 2b in North American dogs in 2006 and 2007. Journal of Clinical Microbiology 45, 4044-4047.
- NETTLES, V. F., PEARSON, J. E., GUSTAFSON, G. A., & BLUE, J. L. (1980). Parvovirus infection in translocated raccoons. Journal of American Veterinary Medical Association 177, 787-789.
- PARRISH, C. R., & KAWAOKA, Y. (2005). The origins of new pandemic viruses: the acquisition of new host ranges by canine parvovirus and influenza A viruses. Annual Review of Microbiology 59, 553-586.
- ROMMELAERE et al, (2010) Cytokine & Growth Factor Reviews 21:185-195.
- TRUYEN, U., AGBANDGE, M., & PARRISH C. L. (1994). Characterization of the feline host range and a specific epitope of feline panleukopenia virus. Virology 200, 494-503.
- WIKOFF, W. R., WANG, G., PARRISH, C. R., CHENG, R. H., CHENG, M. L., STRASSHEIM, T., BAKER, S., & ROSSMAN, M. G. (1994). The structure of a neutralized virus: canine parvovirus complexed with neutralizing antibody fragment. Structure 2, 595-597.
- SUGAI et al, (2009). Epitope mapping of canine distemper virus phosphoprotein by monoclonal antibodies. Microbiol ImmunoI. 2009 53(12):667-74.
- QU et al., (2005) J. Biol. Chem. 280: 23:29568-29595.

While the invention has been described in terms of its preferred embodiments, those skilled in the art will recognize that the invention can be practiced with modification within the spirit and scope of the appended claims. Accordingly, the present invention should not be limited to the embodiments as described above, but should further include all modifications and equivalents thereof within the spirit and scope of the description provided herein.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 7 <210> SEQ ID NO 1 <211> LENGTH: 1800 <212> TYPE: DNA <213> ORGANISM: Raccoon parvovirus <220> FEATURE: <221> NAME/KEY: misc_feature <222> LOCATION: (1274)..(1274) <223> OTHER INFORMATION: n is a, c, q, or t <400> SEQUENCE: 1 60 atgagtgatg gagcagttca accagacggt ggtcaacctg ctgtcagaaa tgaaagagct 120 acaggatetg ggaacgggte tggaggeggg ggtggtggtg gttetggggg tgtggggatt 180 tctacqqqta ctttcaataa tcaqacqqaa tttaaatttt tqqaaaacqq atqqqtqqaa atcacagcaa actcaagcag acttgtacat ttaaatatgc cagaaagtga aaattataga 240 300 agagtggttg taaataattt agataaaact gcagttaacg gaaacatggc tttagatgat actcatgcac aaattgtaac accttggtca ttggttgatg caaatgcttg gggagtttgg 360 tttaatccag gagattggca actaattgtt aatactatga gtgagttgca tttagttagt 420 tttgaacaag aaatttttaa tgttgtttta aagactgttt cagaatctgc tactcagcca 480 ccaactaaag tttataataa tgatttaact gcatcattga tggttgcatt agatagtaat 540 aatactatgc catttactcc agcagctatg agatctgaga cattgggttt ttatccatgg 600 aaaccaacca taccaactcc atggagatat tattttcaat gggatagaac attaatacca 660 teteataetg gaactagtgg cacaceaaca aataeataee atggtaeaga teeagatgat 720 gttcaatttt atactattga aaattctgtg ccagtacact tactaagaac aggtgatgaa 780 tttgctacag gaacattttt ttttgattgt aaaccatgta gactaacaca tacatggcaa 840 acaaatagag cattgggctt accaccattt ctaaattctt tgcctcaatc tgaaggagat 900 actaactttg gtgatatagg aattcaacaa gataaaagac gtggtgtaac tcaaatggga 960 aatacaaact atattactga agctactatt atgagaccag ctgaggttgg ttatagtgca 1020 ccatattatt cttttgaggc gtctacacaa gggccattta aaacacctat tgcagcagga 1080 cggggggggg cgcaaacaga tgaaaatcaa gcagcagatg gtgatccaag atatgcattt 1140 ggtagacaac atggtcaaaa aactaccaca acaggagaaa cacctgagag atttacatat 1200 atagcacatc aagatacagg aagatatcca gaaggagatt ggattcaaaa tattaacttt 1260 aacctteetg taanaaatga taatgtattg etaceaacag ateeaattgg aggtaaaaca 1320 ggaattaact atactaatat atttaatact tatggtcctt taactgcatt aaataatgta 1380 ccaccagttt atccaaatgg tcaaatttgg gataaagaat ttgatactga cttaaaacca 1440 agacttcatg taaatgcacc atttgtttgt caaaataatt gtcctggtca attatttgta 1500 aaagttgcgc ctaatttaac aaatgaatat gatcctgatg catctgctaa tatgtcaaga 1560 1620 attgtaactt actcagattt ttggtggaaa ggtaaattag tatttaaagc taaactaaga gcctctcata cttggaatcc aattcaacaa atgagtatta atgtagataa ccaatttaac 1680 tatgtaccaa gtaatattgg aggtatgaaa attgtatatg aaaaatctca actagcacct 1740 agaaaattat attaacatac ttactatgtt tttatgttta ttacatatca actagcacca 1800

<210> SEQ ID NO 2 <211> LENGTH: 1752 <212> TYPE: DNA -continued

<213 > ORGANISM: Raccoon parvovirus	
<pre><220> FEATURE: <221> NAME/KEY: misc_feature <222> LOCATION: (1274)(1274) <223> OTHER INFORMATION: n is a, c, g, or t</pre>	
<400> SEQUENCE: 2	
atgagtgatg gagcagttca accagacggt ggtcaacctg ctgtcagaaa tgaaagagct	60
acaggatetg ggaacgggte tggaggeggg ggtggtggtg gttetggggg tgtggggatt	120
tctacgggta ctttcaataa tcagacggaa tttaaatttt tggaaaacgg atgggtggaa	180
atcacagcaa actcaagcag acttgtacat ttaaatatgc cagaaagtga aaattataga	240
agagtggttg taaataattt agataaaact gcagttaacg gaaacatggc tttagatgat	300
actcatgcac aaattgtaac accttggtca ttggttgatg caaatgcttg gggagtttgg	360
tttaatccag gagattggca actaattgtt aatactatga gtgagttgca tttagttagt	420
tttgaacaag aaatttttaa tgttgtttta aagactgttt cagaatctgc tactcagcca	480
ccaactaaag tttataataa tgatttaact gcatcattga tggttgcatt agatagtaat	540
aatactatgc catttactcc agcagctatg agatctgaga cattgggttt ttatccatgg	600
aaaccaacca taccaactcc atggagatat tattttcaat gggatagaac attaatacca	660
tctcatactg gaactagtgg cacaccaaca aatacatacc atggtacaga tccagatgat	720
gttcaatttt atactattga aaattctgtg ccagtacact tactaagaac aggtgatgaa	780
tttgctacag gaacattttt ttttgattgt aaaccatgta gactaacaca tacatggcaa	840
acaaatagag cattgggctt accaccattt ctaaattctt tgcctcaatc tgaaggagat	900
actaactttg gtgatatagg aattcaacaa gataaaagac gtggtgtaac tcaaatggga	960
aatacaaact atattactga agctactatt atgagaccag ctgaggttgg ttatagtgca	1020
ccatattatt cttttgaggc gtctacacaa gggccattta aaacacctat tgcagcagga	1080
cggggggggag cgcaaacaga tgaaaatcaa gcagcagatg gtgatccaag atatgcattt	1140
ggtagacaac atggtcaaaa aactaccaca acaggagaaa cacctgagag atttacatat	1200
atagcacatc aagatacagg aagatatcca gaaggagatt ggattcaaaa tattaacttt	1260
aaccttcctg taanaaatga taatgtattg ctaccaacag atccaattgg aggtaaaaca	1320
ggaattaact atactaatat atttaatact tatggtcctt taactgcatt aaataatgta	1380
ccaccagttt atccaaatgg tcaaatttgg gataaagaat ttgatactga cttaaaacca	1440
agacttcatg taaatgcacc atttgtttgt caaaataatt gtcctggtca attatttgta	1500
aaagttgcgc ctaatttaac aaatgaatat gatcctgatg catctgctaa tatgtcaaga	1560
attgtaactt actcagattt ttggtggaaa ggtaaattag tatttaaagc taaactaaga	1620
gcetetcata ettggaatee aatteaacaa atgagtatta atgtagataa eeaatttaae	1680
tatgtaccaa gtaatattgg aggtatgaaa attgtatatg aaaaatctca actagcacct	1740
agaaaattat at	1752
<210> SEQ ID NO 3 <211> LENGTH: 599 <212> TYPE: PRT <213> ORGANISM: Raccoon parvovirus <220> FEATURE: <221> NAME/KEY: misc_feature <222> LOCATION: (425)(425) <223> OTHER INFORMATION: Xaa can be any naturally occurring amino	acid

<400> SEQUENCE: 3

-continued

I	Met 1	Ser	Asp	Gly	Ala 5	Val	Gln	Pro	Asp	Gly 10	Gly	Gln	Pro	Ala	Val 15	Arg
i	Asn	Glu	Arg	Ala 20	Thr	Gly	Ser	Gly	Asn 25	Gly	Ser	Gly	Gly	Gly 30	Gly	Gly
(Gly	Gly	Ser 35	Gly	Gly	Val	Gly	Ile 40	Ser	Thr	Gly	Thr	Phe 45	Asn	Asn	Gln
	Thr	Glu 50	Phe	Lys	Phe	Leu	Glu 55	Asn	Gly	Trp	Val	Glu 60	Ile	Thr	Ala	Asn
:	Ser 65	Ser	Arg	Leu	Val	His 70	Leu	Asn	Met	Pro	Glu 75	Ser	Glu	Asn	Tyr	Arg 80
i	Arg	Val	Val	Val	Asn 85	Asn	Leu	Asp	Lys	Thr 90	Ala	Val	Asn	Gly	Asn 95	Met
j	Ala	Leu	Asp	Asp 100	Thr	His	Ala	Gln	Ile 105	Val	Thr	Pro	Trp	Ser 110	Leu	Val
i	Aap	Ala	Asn 115	Ala	Trp	Gly	Val	Trp 120	Phe	Asn	Pro	Gly	Asp 125	Trp	Gln	Leu
:	Ile	Val 130	Asn	Thr	Met	Ser	Glu 135	Leu	His	Leu	Val	Ser 140	Phe	Glu	Gln	Glu
	Ile 145	Phe	Asn	Val	Val	Leu 150	Гла	Thr	Val	Ser	Glu 155	Ser	Ala	Thr	Gln	Pro 160
1	Pro	Thr	Lys	Val	Tyr 165	Asn	Asn	Asp	Leu	Thr 170	Ala	Ser	Leu	Met	Val 175	Ala
1	Leu	Asp	Ser	Asn 180	Asn	Thr	Met	Pro	Phe 185	Thr	Pro	Ala	Ala	Met 190	Arg	Ser
(Glu	Thr	Leu 195	Gly	Phe	Tyr	Pro	Trp 200	Lys	Pro	Thr	Ile	Pro 205	Thr	Pro	Trp
j	Arg	Tyr 210	Tyr	Phe	Gln	Trp	Asp 215	Arg	Thr	Leu	Ile	Pro 220	Ser	His	Thr	Gly
	Thr 225	Ser	Gly	Thr	Pro	Thr 230	Asn	Thr	Tyr	His	Gly 235	Thr	Asp	Pro	Asp	Asp 240
,	Val	Gln	Phe	Tyr	Thr 245	Ile	Glu	Asn	Ser	Val 250	Pro	Val	His	Leu	Leu 255	Arg
	Thr	Gly	Asp	Glu 260	Phe	Ala	Thr	Gly	Thr 265	Phe	Phe	Phe	Asp	Cys 270	Lys	Pro
(Сув	Arg	Leu 275	Thr	His	Thr	Trp	Gln 280	Thr	Asn	Arg	Ala	Leu 285	Gly	Leu	Pro
1	Pro	Phe	Leu	Asn	Ser	Leu	Pro	Gln	Ser	Glu	Gly	Asp	Thr	Asn	Phe	Gly
j	Asp	Ile	Gly	Ile	Gln	Gln	Asp	Lys	Arg	Arg	Gly	Val	Thr	Gln	Met	Gly
i	Asn	Thr	Asn	Tyr	Ile	Thr	Glu	Ala	Thr	Ile	Met	Arg	Pro	Ala	Glu	320 Val
(Gly	Tyr	Ser	Ala	325 Pro	Tyr	Tyr	Ser	Phe	330 Glu	Ala	Ser	Thr	Gln	335 Gly	Pro
1	Phe	Lys	Thr	340 Pro	Ile	Ala	Ala	Gly	345 Arg	Gly	Gly	Ala	Gln	350 Thr	Asp	Glu
i	Asn	Gln	355 Ala	Ala	Asp	Gly	Asp	360 Pro	Arg	Tyr	Ala	Phe	365 Gly	Arg	Gln	His
(Gly	370 Gln	Lys	Thr	Thr	Thr	375 Thr	Gly	Glu	Thr	Pro	380 Glu	Arg	Phe	Thr	Tyr
	385 Ile	Ala	His	Gln	Asp	390 Thr	Gly	Arq	Tyr	Pro	395 Glu	Gly	Asp	Trp	Ile	400 Gln
;	Asn	Ile	Asn	Phe	405 Asp	Lev	Pro	Val	- Xaa	410 Asr	Asn	Asn	Val	Leu	415 Leu	Pro
-				420		Lou	- 10		425		·P			430	Lu	0

-continued

Thr	Asp	Pro 435	Ile	Gly	Gly	ГЛа	Thr 440	Gly	Ile	Asn	Tyr	Thr 445	Asn	Ile	Phe	
Asn	Thr 450	Tyr	Gly	Pro	Leu	Thr 455	Ala	Leu	Asn	Asn	Val 460	Pro	Pro	Val	Tyr	
Pro 465	Asn	Gly	Gln	Ile	Trp 470	Asp	Lys	Glu	Phe	Asp 475	Thr	Asp	Leu	Lys	Pro 480	
Arg	Leu	His	Val	Asn 485	Ala	Pro	Phe	Val	Cys 490	Gln	Asn	Asn	Суз	Pro 495	Gly	
Gln	Leu	Phe	Val 500	Lys	Val	Ala	Pro	Asn 505	Leu	Thr	Asn	Glu	Tyr 510	Asp	Pro	
Asp	Ala	Ser 515	Ala	Asn	Met	Ser	Arg 520	Ile	Val	Thr	Tyr	Ser 525	Asp	Phe	Trp	
Trp	Lys 530	Gly	Lys	Leu	Val	Phe 535	Lys	Ala	Lys	Leu	Arg 540	Ala	Ser	His	Thr	
Trp 545	Asn	Pro	Ile	Gln	Gln 550	Met	Ser	Ile	Asn	Val 555	Asp	Asn	Gln	Phe	Asn 560	
Tyr	Val	Pro	Ser	Asn 565	Ile	Gly	Gly	Met	Lys 570	Ile	Val	Tyr	Glu	Lys 575	Ser	
Gln	Leu	Ala	Pro 580	Arg	ГÀа	Leu	Tyr	His 585	Thr	Tyr	Tyr	Val	Phe 590	Met	Phe	
Ile	Thr	Tyr 595	Gln	Pro	Ser	Thr										
<212 <213	2> TY 3> OF	(PE : RGANI	PRT SM:	Raco	coon	par	vovi	rus								
<222 <222 <222	L> NZ 2> LC 3> C1	AME/H CATI CHER	CE: CEY: ION: INFO	miso (429 DRMA	c_fea 5) FION	ature (425) : Xaa	e) a car	ı be	any	nati	ıral	ly o	ccuri	ring	amino ac	id
<222 <222 <222 <223	D> FF L> NZ 2> LC 3> OI D> SF	AME/H DCATI THER EQUEN	KE: KEY: LON: INFO NCE:	miso (425 DRMA 4	c_fea 5) FION	ature (425) : Xaa	e) a car	ı be	any	natı	urali	ly o	ccuri	ring	amino ac	id
<222 <222 <222 <223 <400 Met 1	L> NA 2> LC 3> OJ D> SH Ser	AME/I DCATI THER EQUEN	KE: (EY: ION: INF(NCE: Gly	mis (429 DRMA 4 Ala 5	c_fea 5) TION Val	ature (425) : Xaa Gln	e) a car Pro	n be Asp	any Gly 10	natu Gly	ural: Gln	ly oo Pro	ccurr Ala	ring Val 15	amino ac Arg	id
<222 <222 <222 <222 <400 Met 1 Asn	5 FI 1 > NA 2 > LC 3 > OT 3 > OT 5 Ser Glu	AME/I DCAT THER EQUEN Asp Arg	E: (EY: ION: INF(NCE: Gly Ala 20	misc (429 DRMA 4 Ala 5 Thr	c_fea 5) TION Val Gly	ature (425) : Xaa Gln Ser	e) Pro Gly	n be Asp Asn 25	any Gly 10 Gly	natu Gly Ser	Gln Gly	ly o Pro Gly	Ala Gly 30	Val 15 Gly	amino ac Arg Gly	id
<222 <222 <222 <400 Met 1 Asn Gly	Gly	AME/I DCAT: THER EQUEN Asp Arg Ser 35	CEY: ION: INFC ICE: Gly Ala 20 Gly	misc (429 DRMA 4 Ala 5 Thr Gly	c_fea 5) TION Val Gly Val	Gln Gly	Pro Gly Ile 40	Asp Asn 25 Ser	any Gly 10 Gly Thr	natu Gly Ser Gly	Gln Gly Thr	ly o Pro Gly Phe 45	Ala Gly 30 Asn	Val 15 Gly Asn	amino ac Arg Gly Gln	id
<222 <222 <222 <400 Met 1 Asn Gly Thr	Glu Glu Glu Glu Glu	AME/I DCAT THER EQUEN Asp Arg Ser 35 Phe	CE: (CY: (ON: INFC NCE: Gly Ala 20 Gly Lys	miso (429 DRMA 4 Ala 5 Thr Gly Phe	C_fea 5) TION Val Gly Val Leu	Gln Gly Glu Glu	Pro Gly Ile 40 Asn	Asp Asp 25 Ser Gly	any Gly 10 Gly Thr Trp	natu Gly Ser Gly Val	Gln Gly Thr Glu 60	Pro Gly Phe 45 Ile	Ala Gly 30 Asn Thr	Val 15 Gly Asn Ala	amino ac Arg Gly Gln Asn	id
<pre><222 <222 <222 <400 Met 1 Asn Gly Thr Ser 65</pre>	<pre>S PA L> NA L> NA S PA L> NA S PA S PA S PA S PA S PA S PA S PA S P</pre>	And / MME / H AME / H CCAT: THER CQUEN Asp Arg Ser 35 Phe Arg	(E: (EY: (ION: INFC (IVCE: Gly Ala 20 Gly Lys Leu	misc (425 DRMA: 4 Ala 5 Thr Gly Phe Val	C_fea 5) TION Val Gly Val Leu His 70	Gln Gly Glu Leu	Pro Gly Ile 40 Asn	Asp Asp 25 Ser Gly Met	any Gly 10 Gly Thr Trp Pro	natu Gly Ser Gly Val Glu 75	Gln Gly Thr Glu 60 Ser	Pro Gly Phe 45 Ile Glu	Ala Gly 30 Asn Thr Asn	Val 15 Gly Asn Ala Tyr	amino ac Arg Gly Gln Asn Arg 80	id
<pre><222:</pre> <222: <pre></pre>	<pre>S PA 1 > NA 2 > LC 2 > LC 3 > OT 3 > OT 0 > SE Glu Glu 50 Ser Val</pre>	Ang DCAT THER EQUEN Asp Arg Ser 35 Phe Arg Val	(E: (EY: (ON: INFC (CE: Gly Ala 20 Gly Lys Leu Val	misc (422) RMA: 4 Ala 5 Thr Gly Phe Val Asn 85	c_fes 5) TION Val Gly Val Leu His 70 Asn	Glu Gln Ser Gly Leu Leu	Pro Gly Ile 40 Asn Asn Asp	Asp Asp 25 Ser Gly Met Lys	any Gly 10 Gly Thr Trp Pro Thr 90	natu Gly Ser Gly Val Glu 75 Ala	Gln Gly Thr Glu 60 Ser Val	Pro Gly Phe 45 Ile Glu Asn	Ala Gly 30 Asn Thr Asn Gly	Val 15 Gly Asn Ala Tyr Asn 95	amino ac Arg Gly Gln Asn Arg 80 Met	id
<pre><222</pre> <222<222<400 Met 1 Asn Gly Thr Ser 65 Arg Ala	<pre> > NA > NA 2> LC 3> OT)> SF Ser Glu Glu Glu Gly Ser Val Leu</pre>	Arg CATG CAT: THER Asp Arg Ser 35 Phe Arg Val Asp	(E: (EY: (INFC) NCE: Gly Ala 20 Gly Lys Leu Val Asp 100	misc (422 DRMA: 4 Ala 5 Thr Gly Phe Val Asn 85 Thr	c_fes 5) TION Val Gly Val Leu His 70 Asn His	Glu Ser Glu Leu Ala	Pro Gly Ile 40 Asn Asp Gln	Asp Asn 25 Ser Gly Met Lys Ile 105	any Gly 10 Gly Thr Trp Pro Thr 90 Val	natu Gly Ser Gly Val Glu 75 Ala Thr	Gln Gly Thr Glu 60 Ser Val Pro	Pro Gly Phe 45 Ile Glu Asn Trp	Ala Gly 30 Asn Thr Gly Ser 110	Val 15 Gly Asn Ala Tyr Asn 95 Leu	amino ac Arg Gly Gln Asn Arg 80 Met Val	id
<pre><222 <222 <222 <400 Met 1 Asn Gly Thr Gly Thr Ser 65 Arg Ala Asp</pre>	<pre>>>> File NZ >>> LC >>> C >>> SE Glu Glu Glu Ser Val Leu Ala</pre>	Alon ME/I DCAT: THER EQUEN Asp Arg Ser 35 Phe Arg Val Asp Asp 115	(E: (EY: (CN: INFC (CE: Gly Ala 20 Gly Lys Leu Val Asp 100 Ala	misso (422 DRMA: 4 Ala 5 Thr Gly Phe Val Asn 85 Thr Trp	c_fes 5) TION Val Gly Val Leu His 70 Asn His Gly	Glu (425) (425) Ser Glu Ser Gly Leu Leu Ala Val	Pro Gly Ile 40 Asn Asn Asp Gln Trp 120	Asp Asn 25 Ser Gly Met Lys Ile 105 Phe	any Gly 10 Gly Thr Trp Pro Thr 90 Val Asn	natu Gly Ser Gly Val Glu 75 Ala Thr Pro	Gln Gly Thr Glu 60 Ser Val Pro Gly	Pro Gly Phe 45 Ile Glu Asn Trp Asp 125	Ala Gly 30 Asn Thr Asn Gly Ser 110 Trp	Val 15 Gly Asn Ala Tyr Asn 95 Leu Gln	amino ac Arg Gly Gln Asn Arg 80 Met Val Leu	id
<pre><222 <222 <222 <400 Met 1 Asn Gly Thr Gly Thr Ser 65 Ala Asp Ile</pre>	<pre>>>>>>>>>>>>>>>>>>>>>>>>>>>>>>>>>>>></pre>	Ang MAC/I CAT: THER Asp Arg Ser 35 Phe Arg Val Asp Asp Asp Asn 115 Asn	(E: (EY: (CN: INFC (CE: Gly Ala 20 Gly Lys Leu Val Asp 100 Ala Thr	misc (422 DRMA: 4 Ala 5 Thr Gly Phe Val Asn 85 Thr Trp Met	c_fes 5) TION Val Gly Val Leu His 70 Asn His Gly Ser	Glu Ser Gly Leu Ala Glu 135	Pro Gly Ile 40 Asn Asn Gln Trp 120 Leu	Asp Asp 25 Ser Gly Met Lys Ile 105 Phe His	any Gly Oly Thr Trp Pro Thr 90 Val Asn Leu	natu Gly Ser Gly Val Glu 75 Ala Thr Pro Val	Gln Gly Thr Glu 60 Ser Val Pro Gly Ser 140	ly od Pro Gly Phe 45 Ile Glu Asn Trp 125 Phe	Ala Gly 30 Asn Thr Asn Gly Ser 110 Trp Glu	Val 15 Gly Asn Ala Tyr Asn 95 Leu Gln Gln	amino ac Arg Gly Gln Asn Arg 80 Met Val Leu Glu	id
<pre><222</pre> <222<222<222 <pre><400</pre> Met 1 Asn Gly Thr Gly Thr Ser 65 Arg Ala Asp Ile Ile 145	<pre>>> NZ >> NZ >> LC >> OT >> SF Glu Gly Glu 50 Ser Val Leu Ala Val 130 Phe</pre>	Ang MAC/I NAE/I CAT: THER Asp Arg Arg Val Asp Asn 115 Asn Asn	(E: (EY: (CY: (INFC) (I	misc (4225) DRMA: 4 Ala 5 Thr Gly Phe Val Asn 85 Thr Trp Met Val	C_feas 5) TION Val Gly Val Leu His 70 Asn His Gly Ser Leu 150	Glu Ser Gly Glu S5 Leu Ala Val Glu 135 Lys	Pro Gly Ile 40 Asn Asn Gln Trp 120 Leu Thr	Asp Asn 25 Ser Gly Met Lys Ile 105 Phe His Val	any Gly 10 Gly Thr Trp Pro Thr 90 Val Asn Leu Ser	natu Gly Ser Gly Val Glu Thr Pro Val Glu 155	Gln Gly Thr Glu 60 Ser Val Pro Gly Ser 140 Ser	ly of Pro Gly Phe 45 Ile Glu Asn Trp 125 Phe Ala	Ala Gly 30 Asn Thr Asn Gly Ser 110 Trp Glu Thr	Val 15 Gly Asn Ala Tyr Asn 95 Leu Gln Gln	amino ac Arg Gly Gln Asn Arg 80 Met Val Leu Glu Pro 160	iđ
<pre></pre>	<pre>S Fi = NZ > Z > LC > Ser Glu Gly Glu 50 Ser Val Leu Ala Nal 130 Phe Thr</pre>	Alon ME/I NE/I CAT: THER Asp Arg Ser 35 Phe Arg Val Asp Val Asn 115 Asn Lys	(E: (EY: (CY: (INFC) (I	miss (4225) DRMA: 4 Ala 5 Thr Gly Phe Val Asn 85 Thr Trp Met Val Xal Trr 165	c_fes 5) TION Val Gly Val Leu His 70 Asn His Gly Ser Leu 150 Asn	Glu (425) (425) (425) (425) (42) (42) (42) (42) (42) (42) (42) (42	Pro Gly Ile 40 Asn Asn Asn Gln Trp 120 Leu Thr Asp	Asp Asp 25 Ser Gly Met Lys Ile 105 Phe His Val Leu	any Gly Thr Trp Pro Thr 90 Val Asn Leu Ser Thr 170	natu Gly Ser Gly Val Glu 75 Ala Thr Pro Val Glu 155 Ala	Gln Gly Thr Glu 60 Ser Val Pro Gly Ser 140 Ser Ser Ser	ly of Pro Gly Phe 45 Ile Glu Asn Trp 125 Phe Ala Leu	Ala Gly 30 Asn Thr Asn Gly Ser 110 Trp Glu Thr Met	Val 15 Gly Asn Ala Tyr Leu Gln Gln Gln Val 175	amino ac Arg Gly Gln Asn Arg 80 Met Val Leu Glu Pro 160 Ala	iđ

			180					185					190		
Glu	Thr	Leu 195	Gly	Phe	Tyr	Pro	Trp 200	Lys	Pro	Thr	Ile	Pro 205	Thr	Pro	Trp
Arg	Tyr 210	Tyr	Phe	Gln	Trp	Asp 215	Arg	Thr	Leu	Ile	Pro 220	Ser	His	Thr	Gly
Thr 225	Ser	Gly	Thr	Pro	Thr 230	Asn	Thr	Tyr	His	Gly 235	Thr	Asp	Pro	Asp	Asp 240
Val	Gln	Phe	Tyr	Thr 245	Ile	Glu	Asn	Ser	Val 250	Pro	Val	His	Leu	Leu 255	Arg
Thr	Gly	Asp	Glu 260	Phe	Ala	Thr	Gly	Thr 265	Phe	Phe	Phe	Asp	Cys 270	Lys	Pro
Суз	Arg	Leu 275	Thr	His	Thr	Trp	Gln 280	Thr	Asn	Arg	Ala	Leu 285	Gly	Leu	Pro
Pro	Phe 290	Leu	Asn	Ser	Leu	Pro 295	Gln	Ser	Glu	Gly	Asp 300	Thr	Asn	Phe	Gly
Asp 305	Ile	Gly	Ile	Gln	Gln 310	Asp	Lys	Arg	Arg	Gly 315	Val	Thr	Gln	Met	Gly 320
Asn	Thr	Asn	Tyr	Ile 325	Thr	Glu	Ala	Thr	Ile 330	Met	Arg	Pro	Ala	Glu 335	Val
Gly	Tyr	Ser	Ala 340	Pro	Tyr	Tyr	Ser	Phe 345	Glu	Ala	Ser	Thr	Gln 350	Gly	Pro
Phe	Lys	Thr 355	Pro	Ile	Ala	Ala	Gly 360	Arg	Gly	Gly	Ala	Gln 365	Thr	Asp	Glu
Asn	Gln 370	Ala	Ala	Asp	Gly	Asp 375	Pro	Arg	Tyr	Ala	Phe 380	Gly	Arg	Gln	His
Gly 385	Gln	Lys	Thr	Thr	Thr 390	Thr	Gly	Glu	Thr	Pro 395	Glu	Arg	Phe	Thr	Tyr 400
Ile	Ala	His	Gln	Asp 405	Thr	Gly	Arg	Tyr	Pro 410	Glu	Gly	Asp	Trp	Ile 415	Gln
Asn	Ile	Asn	Phe 420	Asn	Leu	Pro	Val	Xaa 425	Asn	Asp	Asn	Val	Leu 430	Leu	Pro
Thr	Asp	Pro 435	Ile	Gly	Gly	ГЛа	Thr 440	Gly	Ile	Asn	Tyr	Thr 445	Asn	Ile	Phe
Asn	Thr 450	Tyr	Gly	Pro	Leu	Thr 455	Ala	Leu	Asn	Asn	Val 460	Pro	Pro	Val	Tyr
Pro 465	Asn	Gly	Gln	Ile	Trp 470	Asp	Lys	Glu	Phe	Asp 475	Thr	Asp	Leu	Lys	Pro 480
Arg	Leu	His	Val	Asn 495	Ala	Pro	Phe	Val	Cys 490	Gln	Asn	Asn	Cys	Pro 495	Gly
Gln	Leu	Phe	Val	Гуз	Val	Ala	Pro	Asn	Leu	Thr	Asn	Glu	Tyr	Asp	Pro
Asp	Ala	Ser	500 Ala	Asn	Met	Ser	Arg	505 Ile	Val	Thr	Tyr	Ser	Aab	Phe	Trp
Trp	Lys	515 Gly	Lys	Leu	Val	Phe	520 Lys	Ala	Lys	Leu	Arg	525 Ala	Ser	His	Thr
- Trn	530 Asp	Pro	- T16	Glr	Gln	535 Met	Ser	TIA	Aan	Val	540 Asp	Agn	Gln	Phe	Asn
545			110	- -	550	net	201	116	-	555	 Jer	- 116			560
Tyr	Val	Pro	Ser	Asn 565	⊥le	GIY	GIY	Met	Lуз 570	ile	Val	Tyr	Gíu	Lуя 575	Ser

Gln Leu Ala Pro Arg Lys Leu Tyr 580

<210> SEQ ID NO 5 <211> LENGTH: 557 <212> TYPE: DNA <213> ORGANISM: Raccoon parvovirus <400> SEQUENCE: 5 tcaggaagat atccagaagg agattggatt caaaatatta actttaacct tcctgtaaca 60 aatgataatg tattgctacc aacagatcca attggaggta aaacaggaat taactatact 120 aatatattta atacttatgg tootttaact goattaaata atgtaccaco agtttatooa 180 aatggtcaaa tttgggataa agaatttgat actgacttaa aaccaagact tcatgtaaat 240 gcaccatttg tttgtcaaaa taattgtcct ggtcaattat ttgtaaaagt tgcgcctaat 300 ttaacaaatg aatatgatcc tgatgcatct gctaatatgt caagaattgt aacttactca 360 gatttttggt ggaaaggtaa attagtattt aaagctaaac taagagcctc tcatacttgg 420 aatccaattc aacaaatqaq tattaatqta qataaccaat ttaactatqt accaaqtaat 480 attggaggta tgaaaattgt atagaaaaat ctcaactagc acctagaaaa ttatattaac 540 aacttagcta gtcttat 557 <210> SEQ ID NO 6 <211> LENGTH: 565 <212> TYPE: DNA <213> ORGANISM: Raccoon parvovirus <400> SEQUENCE: 6 ggtcaggaag atatccagaa ggagattgga ttcaaaatat taactttagc cttccggtgg 60 caaatgataa tgtattgctg ccaacaggtc caattggagg taaaacagga attaactata 120 ctaatatatt taatacgtat ggtcctttaa ctgcattaaa taatgtacca ccagtttatc 180 caaatggtca aatttgggat aaagaatttg atactgactt aaaaccaaga cttcatgtaa 240 atgcaccatt tgtttgtcaa aataattgtc ctggtcaatt atttgtaaaa gttgcgccta 300 atttaacaaa tqaatatqat cctqatqcat ctqctaatat qtcaaqaatt qtaacttact 360 420 cagatttttg gtggaaaggt aaattagtat ttaaagctaa actaagagcc tctcatactt qqaatccaat tcaacaaatq aqtattaatq taqataacca atttaactat qtaccaaqta 480 atattggagg tatgaaaatt gtatagaaaa attctcaaac tagcacctag aaaattatat 540 aacaacttta cttctggtgg gcggg 565 <210> SEQ ID NO 7 <211> LENGTH: 556 <212> TYPE: DNA <213> ORGANISM: Raccoon parvovirus <400> SEQUENCE: 7 tcaggaagat atccagaagg agattggatt caaaatatta actttaacct tcctgtaaca 60 aatgataatg tattgctacc aacagatcca attggaggta aaacaggaat taactatact 120 aatatattta atacttatgg tootttaact goattaaata atgtaccaco agtttatooa 180 aatggtcaaa tttgggataa agaatttgat actgacttaa aaccaagact tcatgtaaat 240 gcaccatttg tttgtcaaaa taattgtcct ggtcaattat ttgtaaaagt tgcgcctaat 300

CODE	п.	nuad	

				-contir	nued	
ttaacaaatg	aatatgatcc	tgatgcatct	gctaatatgt	caagaattgt	aacttactca	360
gatttttggt	ggaaaggtaa	attagtattt	aaagctaaac	taagagcctc	tcatacttgg	420
aatccaattc	aacaaatgag	tattaatgta	gataaccaat	ttaactatgt	accaagtaat	480
attggaggta	gaaaattgta	tagaaaaatc	tcaactagca	cctagaaaat	tatattaaca	540
acttagctag	gcgtat					556

40

I claim:

1. A raccoon parvovirus (RPV) comprising the characteristics of i) Parvovirus-2 (Raccoon) of ATCC NO. PTA-11400, or ii) progeny of Parvovirus-2 (Raccoon) of ATCC NO. PTA-11400, wherein said characteristics include that said RPV comprises a nucleic acid encoding a VP-2 protein, said nucleic acid comprising nucleotide sequence SEQ ID NO: 1.

2. An immunogenic composition comprising a nucleic acid comprising the nucleotide sequence SEQ ID NO: 1, or a portion of SEQ ID NO: 1; wherein said portion of SEQ ID NO: 1 encodes an antigenic region of a VP-2 protein comprising both amino acid residue Thr at position 232 and amino acid residue Asp at position 300 of SEQ ID NO: 3, wherein said nucleic acid is present in a killed or attenuated parvovirus virion or low passage raccoon parvovirus (RPV).

3. The immunogenic composition of claim 2 wherein said killed parvovirus virion comprises the characteristics of i) Parvovirus-2 (Raccoon) having the ATCC NO. PTA-11400, or ii) progeny of Parvovirus-2 (Raccoon) of ATCC NO. PTA-11400, wherein said characteristics include that said RPV comprises a nucleic acid encoding a VP-2 protein, said nucleic acid comprising the nucleotide sequence SEQ ID NO: 1

4. The immunogenic composition of claim 2, wherein said attenuated parvovirus virion is present in a solid carrier suitable for supralingual dissolution.

5. The-immunogenic composition of claim 2, wherein said immunogenic composition is suitable for subcutaneous administration.

6. A method of eliciting an immune response in an animal against parvovirus infection, comprising the step of administering to said animal an immunogenic composition comprising a nucleic acid comprising the nucleotide sequence SEQ

ID NO: 1, or a portion of SEQ ID NO: 1, wherein said portion of SEQ ID NO: 1 encodes an antigenic region of a VP-2 protein comprising both amino acid residue Thr at position 232 and amino acid residue Asp at position 300 of SEQ ID NO: 3.

7. The method of claim 6, wherein said nucleic acid is present in a killed or attenuated parvovirus virion or low 20 passage RPV in a killed or attenuated parvovirus virion.

8. The method of claim 7, wherein said killed parvovirus virion comprises the characteristics of i) Parvovirus-2 (Raccoon) having the ATCC NO. PTA-11400,or ii) progeny of Parvovirus-2 (Raccoon) of ATCC NO. PTA-11400, wherein said characteristics include that said RPV comprises a nucleic acid encoding a VP-2 protein, said nucleic acid comprising the nucleotide sequence SEQ ID NO: 1.

9. The method of claim 7, wherein said attenuated parvovirus virion is present in a solid carrier suitable for supralingual dissolution.

10. The method of claim 6, wherein said immunogenic composition is suitable for subcutaneous administration.

11. The method of claim 9, wherein said animal is a puppy.

12. An expression vector comprising a nucleotide sequence 35 encoding the amino acid sequence of SEQ ID NO: 3, or a portion of SEQ ID NO: 3; wherein said portion of SEQ ID NO: 3 is an antigenic region of a VP-2 protein comprising both amino acid residue Thr at position 232 and amino acid residue Asp at position 300 of SEQ ID NO: 3,

wherein said expression vector is a recombinant expression vector or a killed or attenuated viral vector.

13. The expression vector of claim 12, that is a recombinant viral expression vector.