



US007794734B2

(12) **United States Patent**
Ayalew et al.(10) **Patent No.:** **US 7,794,734 B2**
(45) **Date of Patent:** **Sep. 14, 2010**(54) **MANNHEIMIA HAEMOLYTICA CHIMERIC
OUTER MEMBRANE PROTEIN PLPE AND
LEUKOTOXIN EPITOPES AS A VACCINE OR
VACCINE COMPONENT AGAINST SHIPPING
FEVER**(75) Inventors: **Sahlu Ayalew**, Stillwater, OK (US);
Anthony W. Confer, Stillwater, OK
(US)(73) Assignee: **The Board of Regents for Oklahoma
State University**, Stillwater, OK (US)(*) Notice: Subject to any disclaimer, the term of this
patent is extended or adjusted under 35
U.S.C. 154(b) by 0 days.(21) Appl. No.: **11/545,069**(22) Filed: **Oct. 6, 2006**(65) **Prior Publication Data**

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Related U.S. Application Data(60) Continuation-in-part of application No. 11/235,982,
filed on Sep. 27, 2005, now Pat. No. 7,144,580, which
is a division of application No. 10/696,544, filed on
Oct. 29, 2003, now abandoned.(60) Provisional application No. 60/757,342, filed on Jan.
9, 2006, provisional application No. 60/422,305, filed
on Oct. 30, 2002.(51) **Int. Cl.****A61K 39/00** (2006.01)
A61K 39/102 (2006.01)
A61K 39/02 (2006.01)
A61K 39/38 (2006.01)
A61K 39/116 (2006.01)
A61K 38/00 (2006.01)
C07K 14/00 (2006.01)(52) **U.S. Cl.** **424/255.1; 424/192.1; 424/190.1;**
424/184.1; 424/234.1; 424/203.1; 514/2;
530/350; 530/300; 530/825; 530/806(58) **Field of Classification Search** **424/192.1,**
424/184.1, 203.1, 190.1, 234.1, 255.1; 514/2;
530/350, 300, 825, 806

See application file for complete search history.

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outer membrane proteins of *Pasteurella haemolytica* serotype 1,"
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(Continued)

Primary Examiner—S. Devi(74) *Attorney, Agent, or Firm*—Fellers, Snider, Blankenship,
Bailey & Tippens(57) **ABSTRACT**Vaccine preparations for the prevention and treatment of
bovine respiratory disease (BRD) and, in particular, its most
severe form, termed "shipping fever", are provided. The
preparations comprise chimeric proteins comprising immun-
odominant epitopes of recombinant *Mannheimia*
haemolytica outer membrane protein PlpE, and immun-
odominant epitopes of recombinant *M. haemolytica* leuko-
toxin.**13 Claims, 26 Drawing Sheets**

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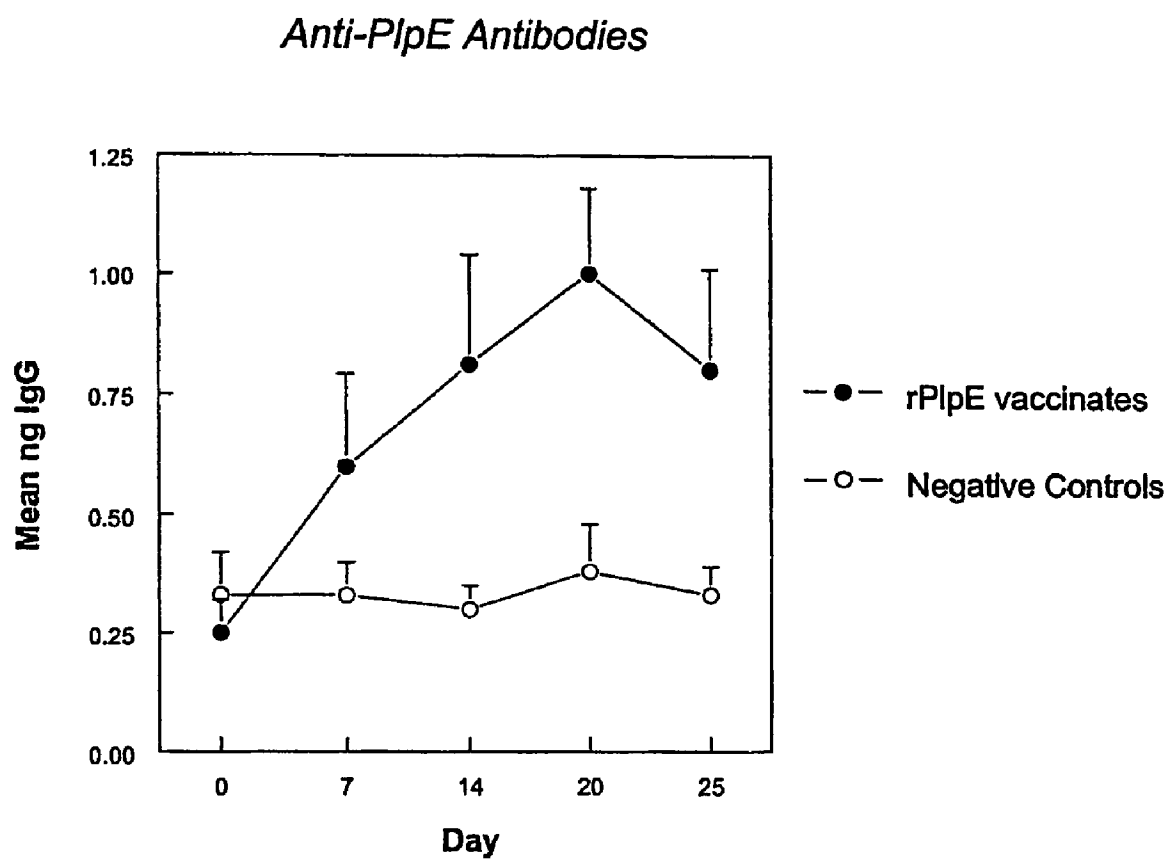
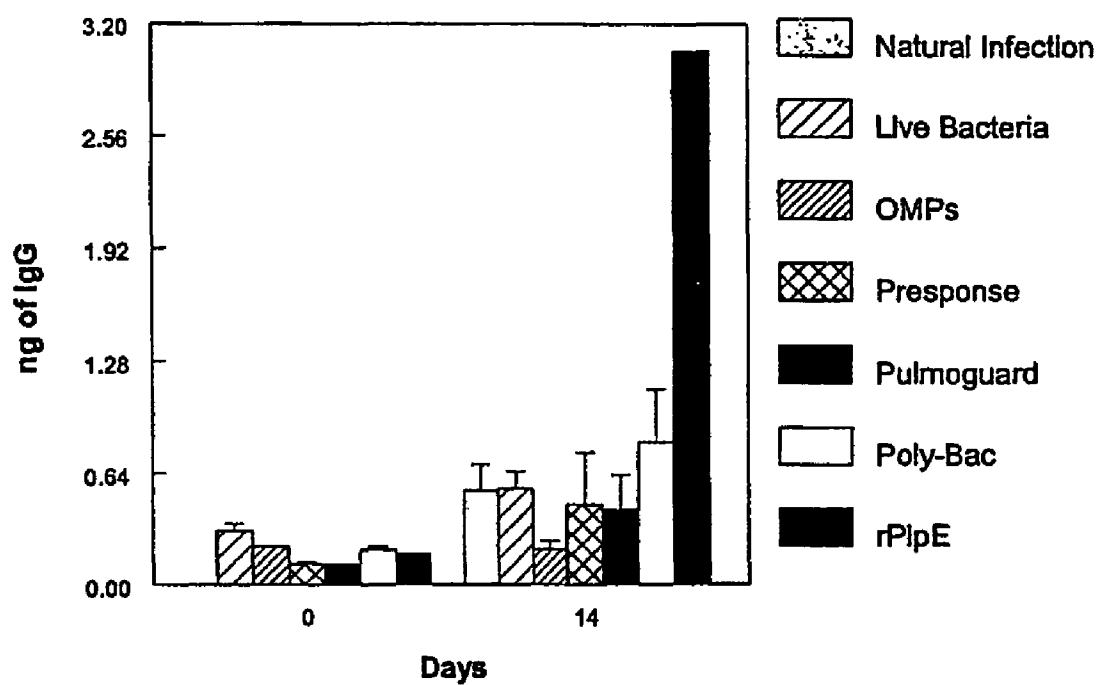
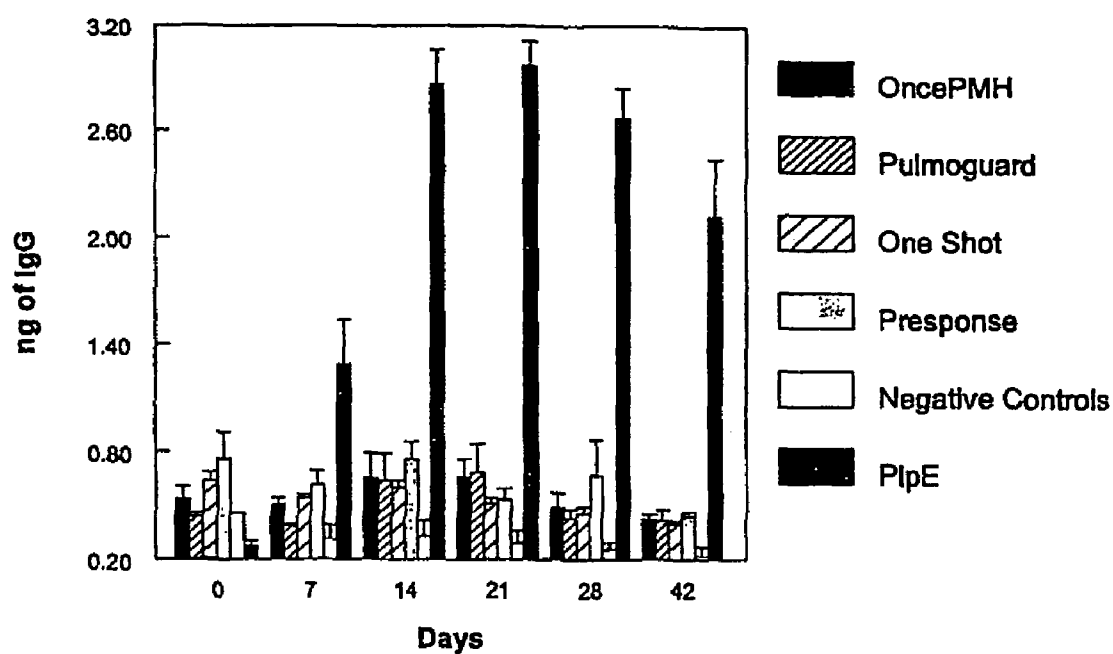


FIG. 1

Anti-PipE: Commercial *M. haemolytica* Vaccines - Exp. 1**FIG. 2**

Anti-PlpE:Commercial *M. haemolytica* Vaccines - Exp. 2**FIG. 3**

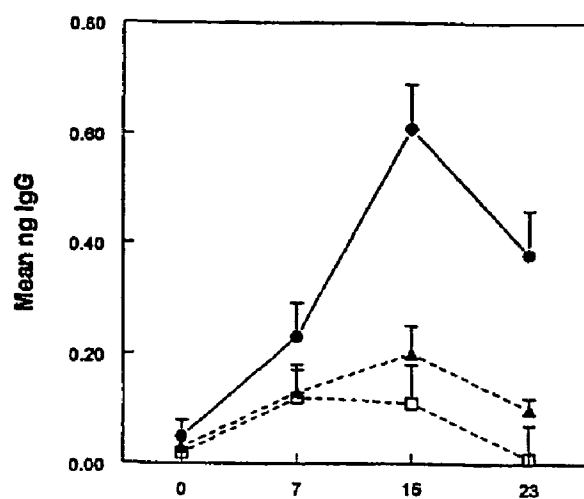


FIG. 4A

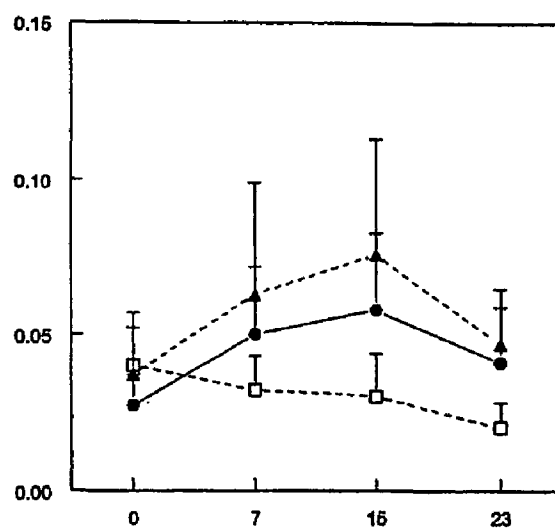


FIG. 4B

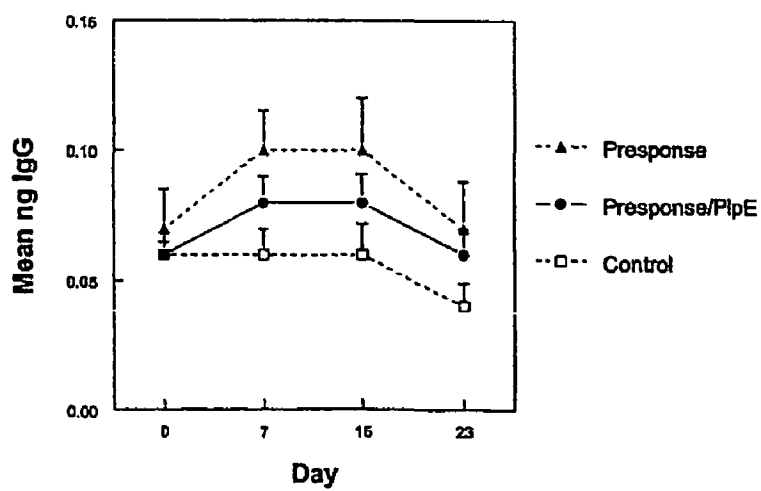


FIG. 4C

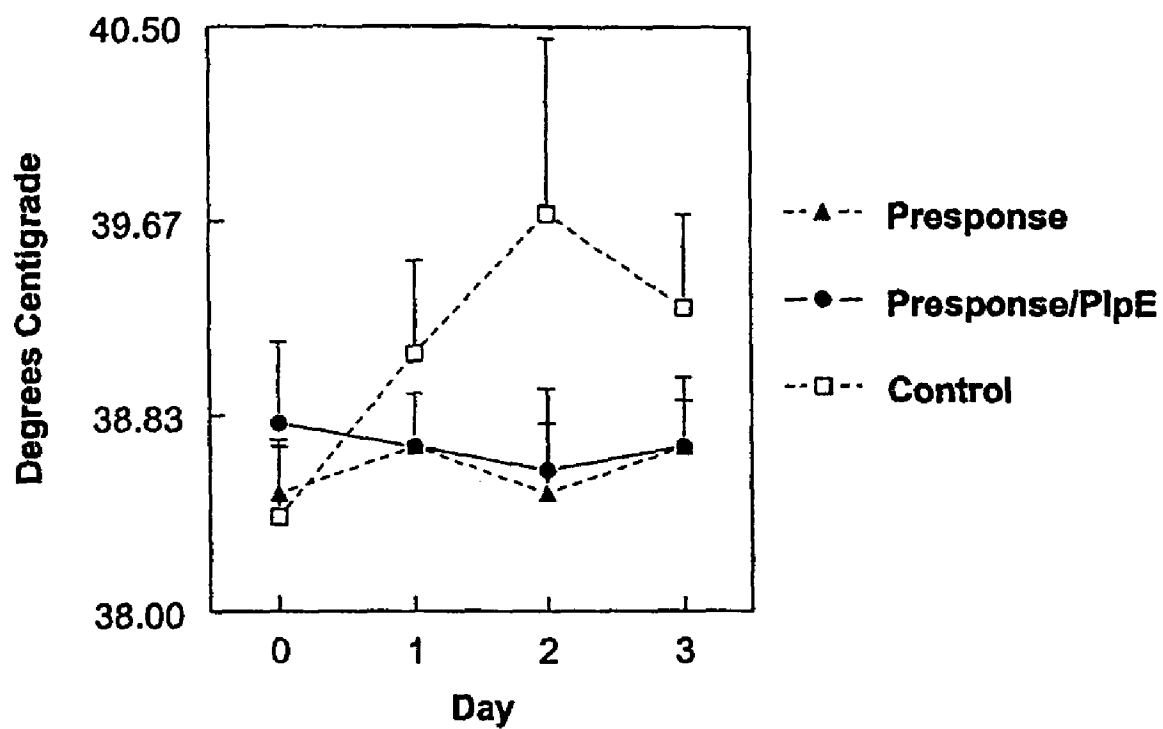
Rectal temperatures after challenge

FIG. 5

Epitopes of P1pE Determined by Peptide Array (Peptscan)

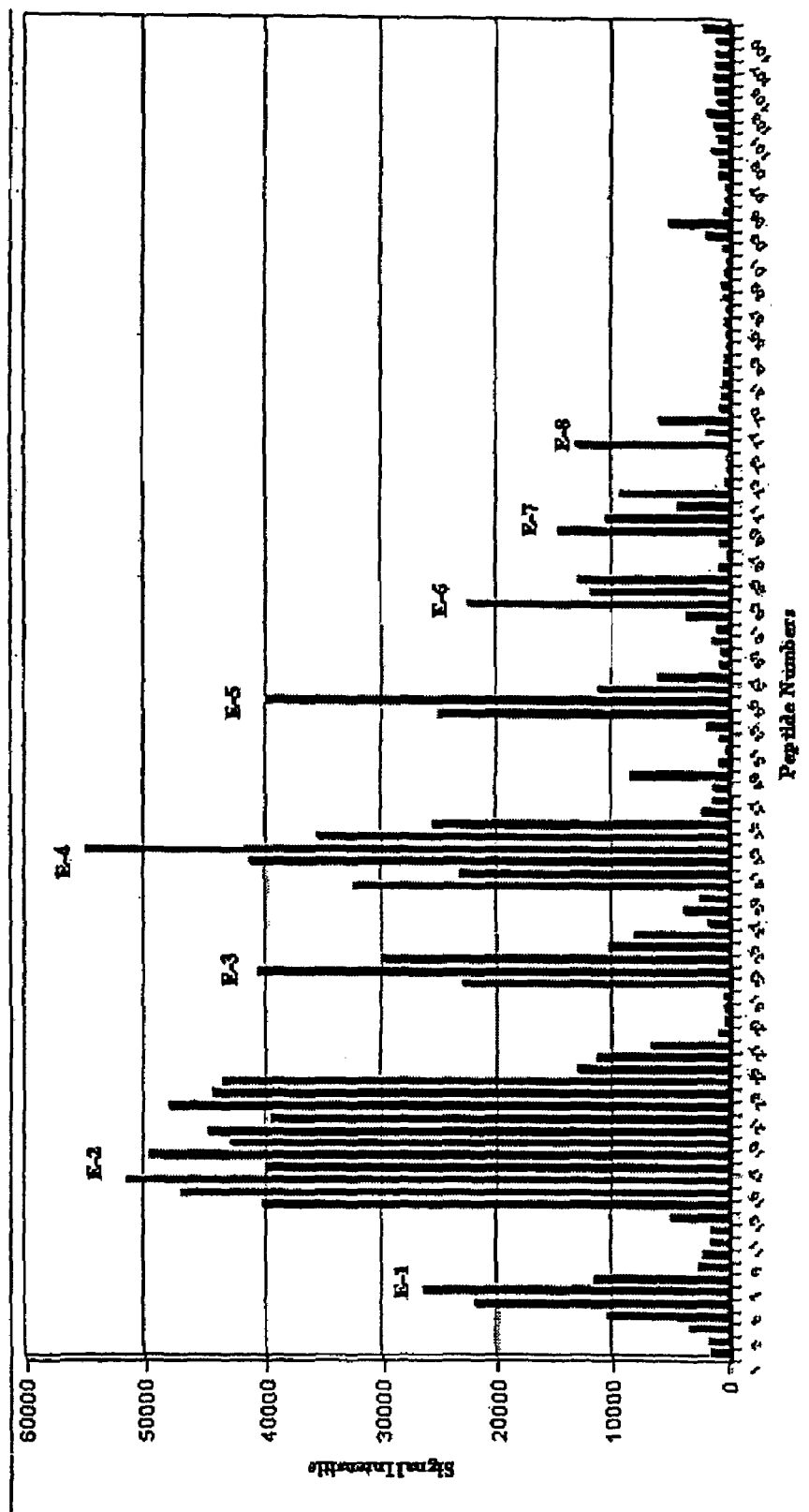


FIG. 6

A.

TPNHPKPVLPKQTQNNLQAQNVPPAQNASQAQNAPQAQNAPQAQNAPQVENAPQA

B.

ACACCGAATCACCCCAAACCAGTACTAGTACCAAAAACACAAAATAATCTTCAAGC
ACAAAATGTTCTCAGGCACAAAATGCCTCTCAGGCACAAAATGCCCCTCAGGCAC
AAAATGCTCCTCAGGCACAAAATGCTCCTCAGGTGGAAAATGCTCCTCAGGCA

Figure 7 A-B

A.

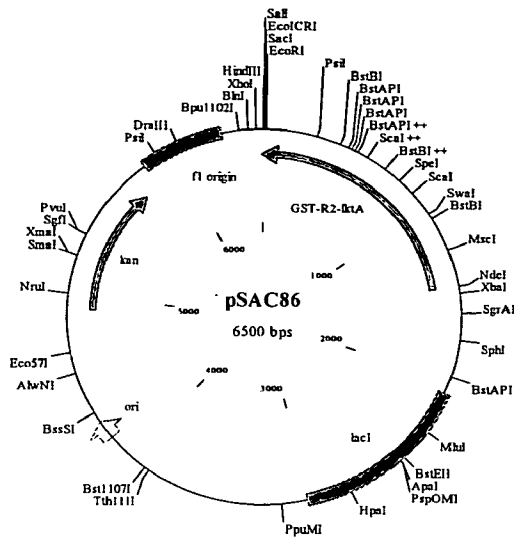
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RNVLVAPTSM

B.

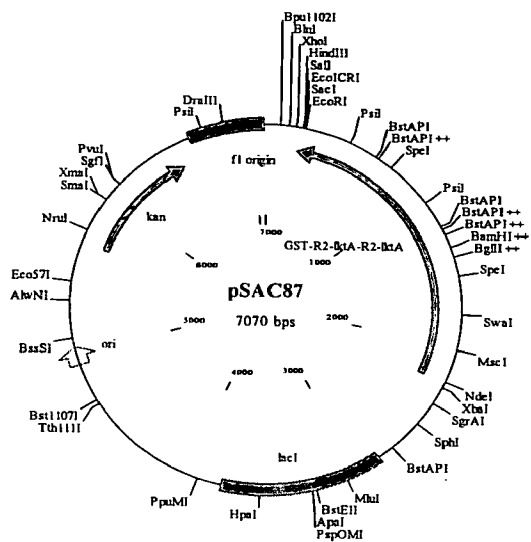
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TCGGTCAAAATGGCGAGCGGATCACCTCAAAGCAAGTTGATGATCTTATCGCAAAA
GGTAACGGCAAAATTACCCAAGATGAGCTATCAAAAGTTGTTGATAACTATGAATTG
CTCAAACATAGCAAAAATGTGACAAACAGCTTAGATAAGTTAATCTCATCTGTAAGT
GCATTTACCTCGTCTAATGATTCGAGAAATGTATTAGTGGCTCCAACCTTCAATG

Figure 8 A-B

9A



9B



9C

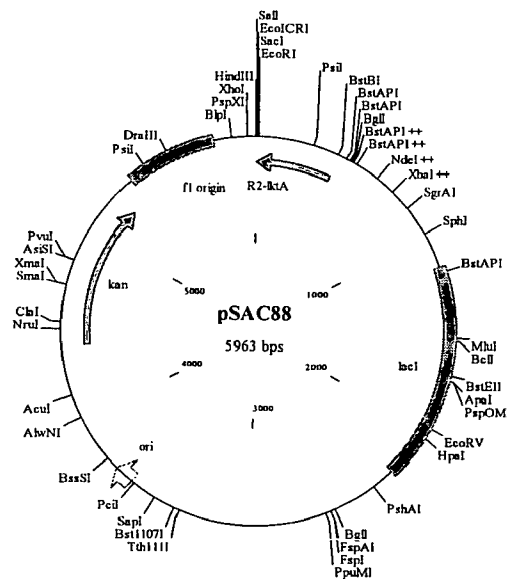
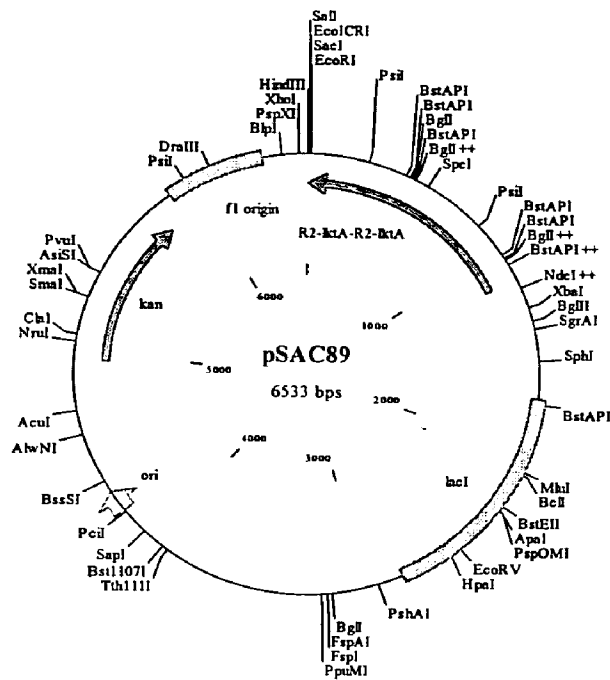


Figure 9 A-C

9D



9E

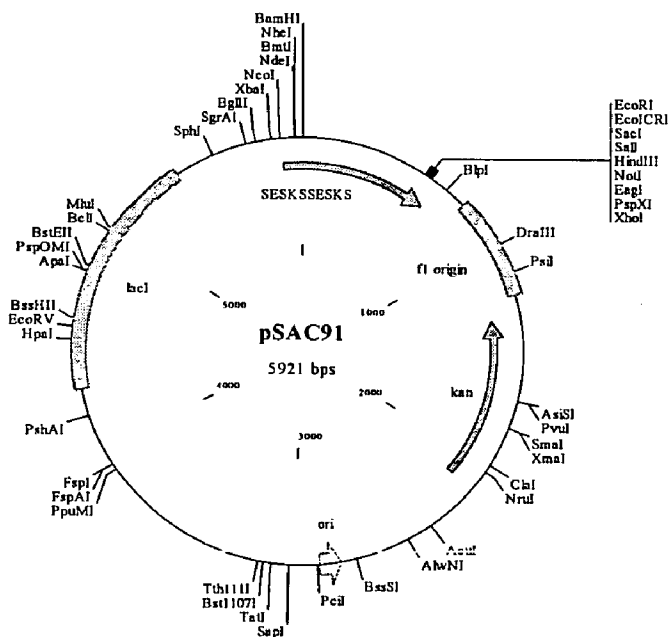


Figure 9 D-E

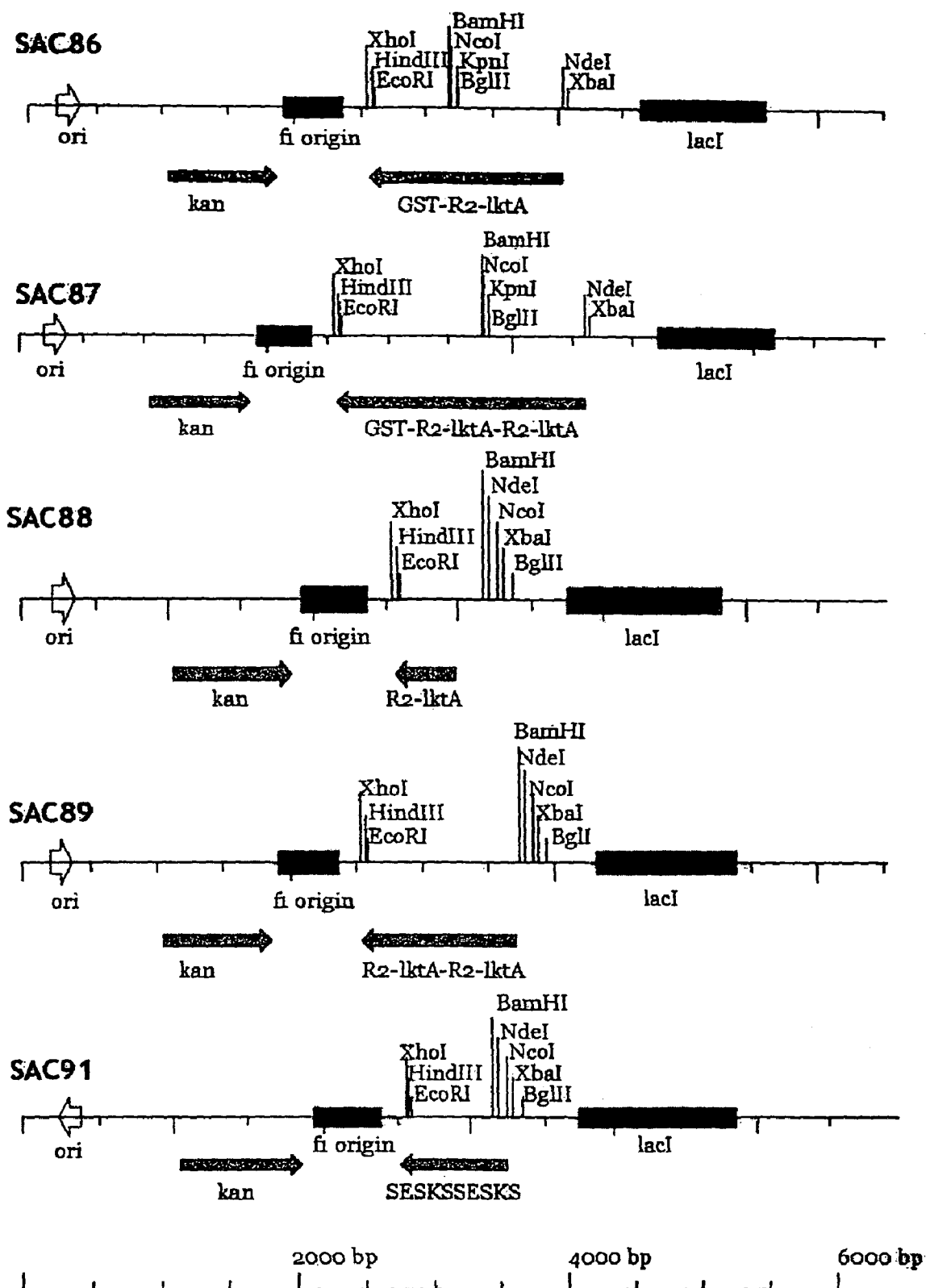


Figure 9F

A.

MSPILGYWKIKGLVQPTRLLEYLEEKYEEHLYERDEGDKWRNKKFELGLEFPNLPYYID
GDVKLTQSMAIRYIADKHNMLGGCPKERAISMLEGAVLDIRYGVSRIAYSKDFETLKV
DFLSKLPEMLKMFEDRLCHKTYLNGDHVTHPDFMLYDALDVVLYMDPMCLDAFPKLV
CFKKRIEAIPOIDKYLKSSKYIAWPLQGWWQATFGGGDHPPKSDGSTSGSGHHHHHSAG
LVPRGSTAIGMKETAATAKFERQHMDSPDLGTGGGSGDDDDKSPMGYRGSTPNHPKPV
LVPKTQNNLQAQNVPOAQNASQAQONAPQAQONAPQAQONAPOVENAPOA RS SDSN
LKDLTFEKKVKNLVITNSKKEKVTIQNWFEADFAKEVPNYKATKDEKIEEIIGQN
GERITSKQVDDLIAGNGKITQDELSKVVDNYELLKHSKNVTNSLDKLISSVSAFTSS
NDSRNVLVAPTSM RS EFELRRQALIN

Figure 10A

B.

TGTCCCCTATACTAGGTTATTGGAAAATTAAGGGCCTTGTGCAACCCACTCGACTTC
TTTTGGAATATCTTGAAGAAAAATATGAAGAGCATTTGTATGAGCGCGATGAAGGTG
ATAAATGGCGAAACAAAAAGTTTGAATTGGGTTTGGAGTTTCCCAATCTTCCTTATT
ATATTGATGGTGATGTAAATTAACACAGTCTATGGCCATCATAACGTTATATAGCTG
ACAAGCACAAACATGTTGGGTGGTTGTCCAAAAGAGCGTGCAGAGATTTCAATGCTTG
AAGGAGCGGTTTTGGATATTAGATACGGTGTTCGAGAATTGCATATAGTAAAGACT
TTGAAACTCTCAAAGTTGATTTTCTTAGCAAGCTACCTGAAATGCTGAAAATGTTTCG
AAGATCGTTTATGTCATAAAACATATTTAAATGGTGATCATGTAACCCATCCTGACT
TCATGTTGTATGACGCTCTTGATGTTGTTTTATACATGGACCCAATGTGCCTGGATGC
GTTCCCAAATAGTTTGTTTTAAAAACGTATTGAAGCTATCCCACAAATTGATAA
GTACTTGAAATCCAGCAAGTATATAGCATGGCCTTTGCAGGGCTGGCAAGCCACGTT
TGGTGGTGGCGACCATCCTCCAAAATCGGATGGTTCAACTAGTGGTTCTGGTCATCA
CCATCACCATCACTCCGCGGGTCTGGTGCCACGCGGTAGTACTGCAATTGGTATGAA
AGAAACCGCTGCTGCTAAATTCGAACGCCAGCACATGGACAGCCCAGATCTGGGTA
CCGGTGGTGGCTCCGGTGATGACGACGACAAGAGTCCCATGGGATATCGGGGATCC
ACACCGAATCACCCCAAACCAGTACTAGTACCAAAAACACAAAATAATCTTCAAGC
ACAAAATGTTCTCAGGCACAAAATGCCTCTCAGGCACAAAATGCCCCTCAGGCAC
AAAATGCTCCTCAGGCACAAAATGCTCCTCAGGTGGAAAATGCTCCTCAGGCAAGA
TCCTCTGATTCGAACTTAAAAGATTTAACATTTGAAAAAGTTAAACATAATCTTGTC
ATCACGAATAGCAAAAAAGAGAAAGTGACCATTCAAAACTGGTTCCGAGAGGCTGA
TTTTGCTAAAGAAGTGCCTAATTATAAAGCAACTAAAGATGAGAAAATCGAAGAAA
TCATCGGTCAAAATGGCGAGCGGATCACCTCAAAGCAAGTTGATGATCTTATCGCAA
AAGGTAACGGCAAAATTACCCAAGATGAGCTATCAAAAGTTGTTGATAACTATGAA
TTGCTCAAACATAGCAAAAATGTGACAAACAGCTTAGATAAGTTAATCTCATCTGTA
AGTGCATTTACCTCGTCTAATGATTGAGAAATGTATTAGTGGCTCCAACCTCAATG
AGATCCGAATTCGAGCTCCGTCGACAAGCTTTAATTAATTA

Figure 10B

A.

MSPILGYWKIKGLVQPTRLLLEYLEEKYEEHLYERDEGDKWRNKKFELGLEFPNLPYYID
GDVKLTQSMAIIRYIADKHNMLGGCPKERAIEISMLEGAVLDIRYGVSR IAYSKDFETLKV
DFLSKLPEMLKMFEDRLCHKTYLNGDHVTHPDFMLYDALDVVLYMDPMCLDAFPKLV
CFKKRIEAI PQIDKYLKSSKYIAWPLQGWQATFGGGDHPPKSDGSTSGSGHHHHHSAG
LVPRGSTAIGMKETAAAKFERQHMDSPDLGTGGGSGDDDDKSPMGYRGSTPNHPKPV
LVPKTQNNLQAQNVPOAQNASQAQNAPOAQNAPOAQNAPOVENAPOA RS SDSNL
KDLTFEKVKHNLVITNSKKEKVTIQNW FREADFAKEVPNYKATKDEKIEEII GQNG
ERITSKQVDDLIAGNGKITQDELSKVVDNYELLKHSKNVTNSL DKLISVS AFTSSN
DSRNVLVAPTSM RS TPNHPKPV LVPKTQNNLQAQNVPOAQNASQAQNAPOAQNA
POA QNAPOVEN APOA RS SDSNLKDLTFEKVKHNLVITNSKKEKVTIQNW FREAD
FAKEVPNYKATKDEKIEEII GQNGERITSKQVDDLIAGNGKITQDELSKVVDNYEL
LKHSKNVTNSL DKLISVS AFTSSNDSRNVLVAPTSM RS EFELRRQALIN

Figure 11A

B.

TGTCCTTATACTAGGTTATTGGAAAATTAAGGGCCTTGTGCAACCCACTCGACTTC
TTTTGGAATATCTTGAAGAAAAATATGAAGAGCATTGTATGAGCGCGATGAAGGTG
ATAAATGGCGAAACAAAAAGTTTGAATTGGGTTTGGAGTTTCCCAATCTTCCTTATT
ATAATTGATGGTGATGTTAAATTAACACAGTCTATGGCCATCATACGTTATATAGCTG
ACAAGCACAACATGTTGGGTGGTTGTCCAAAAGAGCGTGCAGAGATTTCAATGCTTG
AAGGAGCGGTTTTGGATATTAGATACGGTGTTCGAGAATTGCATATAGTAAAGACT
TTGAAACTCTCAAAGTTGATTTTCTTAGCAAGCTACCTGAAATGCTGAAAATGTTCTG
AAGATCGTTTATGTCATAAAACATATTTAAATGGTGATCATGTAACCCATCCTGACT
TCATGTTGTATGACGCTCTTGATGTTGTTTTATACATGGACCCAATGTGCCTGGATGC
GTTCCCAAATTAGTTTGTTTTAAAAAACGTATTGAAGCTATCCACAAAATTGATAA
GTACTTGAAATCCAGCAAGTATATAGCATGGCCTTTGCAGGGCTGGCAAGCCACGTT
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CCGGTGGTGGCTCCGGTGATGACGACGACAAGAGTCCCATGGGATATCGGGGATCC
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AAAATGCTCCTCAGGCACAAAATGCTCCTCAGGTGGAAAATGCTCCTCAGGCAAGA
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TTTTGCTAAAGAAGTGCCTAATTATAAAGCAACTAAAGATGAGAAAATCGAAGAAA
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TTGCTCAAACATAGCAAAAATGTGACAAACAGCTTAGATAAGTTAATCTCATCTGTA
AGTGCAATTTACCTCGTCTAATGATTCGAGAAATGTATTAGTGGCTCCAACCTTCAATG
AGATCCGAATTCGAGCTCCGTGACAAAGCTTTAATTAATTA

Figure 11B

A.

MGSSHHHHHHSSGLVPRGSHMASMTGGQQMGRGSTPNHPKPVLPKTONNLQAO
NVPQAQNASQAQNAPOAQNAPOAQNAPOQVENAPOA RS SDSNLKDLTFEKVKHNL
VITNSKKEKVTIQNWFEADFAKEVPNYKATKDEKIEEIIIGQNGERITSKQVDDLIA
KGNGKITQDELSKVVDNYELLKHSKNVTNSLDKLISSVSAFTSSNDSRNVLVAPTSM
RS EFELRRQALIN

B.

ATGGGCAGCAGCCATCATCATCATCACAGCAGCGGCCTGGTGCCGCGCGGCAG
CCATATGGCTAGCATGACTGGTGGACAGCAAATGGGTTCGCGGATCCACACCGAATC
ACCCCAAACCAGTACTAGTACCAAAAACACAAAATAATCTTCAAGCACAAAATGTT
CCTCAGGCACAAAATGCCTCTCAGGCACAAAATGCCCCTCAGGCACAAAATGCTCCT
CAGGCACAAAATGCTCCTCAGGTGGAAAATGCTCCTCAGGCAAGATCCTCTGATTCTG
AACTTAAAAGATTTAACATTTGAAAAAGTTAAACATAATCTTGTCATCACGAATAGC
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AGCAAAAATGTGACAAACAGCTTAGATAAGTTAATCTCATCTGTAAGTGCATTTACC
TCGTCTAATGATTTCGAGAAATGTATTAGTGGCTCCAACCTCAATGAGATCCGAATTC
GAGCTCCGTCGACAAGCTTTAATTAATTAA

Figure 12 A-B

A.

MGSSHHHHHSSGLVPRGSHMASMTGGQQMGRGSTTPNHPKPVLVPKTQNNLQAQN
VPQAQNASQAQNAPOQAQNAPOQAQNAPOVENAPOA RS SDSNLKDLTFEKKVHNLV
ITNSKKEKVTIQNWFEADFAKEVPNYKATKDEKIEEIIGQNGERITSKQVDDLI
AKGNGKITQDELSKVVDNYELLKHSKNVTNSL
DKLISSVSAFTSSNDSRNVLVAPTSM
RS TPNHPKPVLVPKTQNNLQAQNVPOQAQNASQAQNAPOQAQNAPOVENA
POA RS SDSNLKDLTFEKKVHNLVITNSKKEKVTIQNWFEADFAKEVPNYKATK
DEKIEEIIGQNGERITSKQVDDLI
AKGNGKITQDELSKVVDNYELLKHSKNVTNSL
DKLISSVSAFTSSNDSRNVLVAPTSM RS EFELRRQALIN

Figure 13A

B.

ATGGGCAGCAGCCATCATCATCATCACAGCAGCGGCCTGGTGCCGCGCGGCAG
CCATATGGCTAGCATGACTGGTGGACAGCAAATGGGTCGCGGATCCACACCGAATC
ACCCCAAACCAGTACTAGTACCAAAAAACAAAAATAATCTTCAAGCACAAAATG
TTCCTCAGGCACAAAATGCCTCTCAGGCACAAAATGCCCTCAGGCACAAAATG
CTCCTCAGGCACAAAATGCTCCTCAGGTGGAAAATGCTCCTCAGGCA *AGATCC*
TCTGATTTCGAACTTAAAAGATTTAACATTTGAAAAAGTTAAACATAATCTTGTCA
TCACGAATAGCAAAAAAGAGAAAAGTGACCATTCAAACTGGTTCCGAGAGGCT
GATTTTGCTAAAGAAGTGCCTAATTATAAAGCAACTAAAGATGAGAAAATCGAA
GAAATCATCGGTCAAAATGGCGAGCGGATCACCTCAAAGCAAGTTGATGATCTT
ATCGCAAAAGGTAACGGCAAAATTACCCAAGATGAGCTATCAAAAGTTGTTGAT
AACTATGAATTGCTCAAAACATAGCAAAAAATGTGACAAACAGCTTAGATAAGTTA
ATCTCATCTGTAAGTGCATTTACCTCGTCTAATGATTTCGAGAAATGTATTAGTG
GCTCCAACCTTCAATG *AGATCC* ACACCGAATCACCCCAAACCAGTACTAGTACC
AAAAACACAAAATAATCTTCAAGCACAAAATGTTCTCAGGCACAAAATGCCTC
TCAGGCACAAAATGCCCTCAGGCACAAAATGCTCCTCAGGCACAAAATGCTCC
TCAGGTGGAAAATGCTCCTCAGGCA *AGATCC* TCTGATTTCGAACTTAAAAGATT
TAACATTTGAAAAAGTTAAACATAATCTTGTCTATCACGAATAGCAAAAAAGAGA
AAGTGACCATTCAAACTGGTTCCGAGAGGCTGATTTTGCTAAAGAAGTGCCTA
ATTATAAAGCAACTAAAGATGAGAAAATCGAAGAAATCATCGGTCAAAATGGCG
AGCGGATCACCTCAAAGCAAGTTGATGATCTTATCGCAAAAGGTAACGGCAAAA
TTACCCAAGATGAGCTATCAAAAGTTGTTGATAACTATGAATTGCTCAAAACATA
GCAAAAATGTGACAAACAGCTTAGATAAGTTAATCTCATCTGTAAGTGCATTTA
CCTCGTCTAATGATTTCGAGAAATGTATTAGTGGCTCCAACCTTCAATG *AGATCC*
GAATTCGAGCTCCGTCGACAAGCTTTAATTAATTAA

Figure 13B

A.

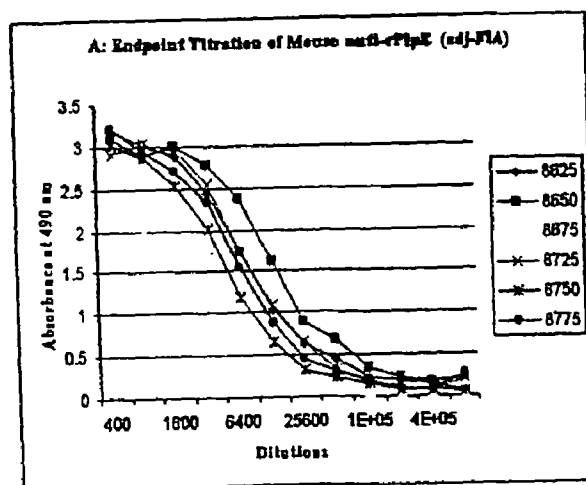
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EIIQNGERI *GGGS* SRSGGGS QAQNASQAQNAPQAQNAPQAQNAPQVENAPQAQ
NAPQVENAPQ*GGGS* FREAEFAKTIQNYVATRDDKIEEIIQNGERI *GGGS*
RSEFELRRQACGRTRAPPPPLRSGC

B.

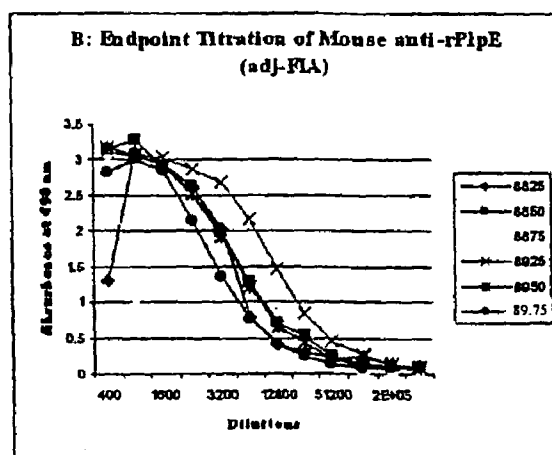
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GCACAAAATGCTCCTCAGGTGGAAAATGCTCCTCAGGCACAAAATGCTCCTCAGGT
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AATGGTGAACGGATTGGCGGTGGTGGGTGAGATCCGAATTCGAGCTCCGTGACACA
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Figure 14A-B

15A



15B



15C

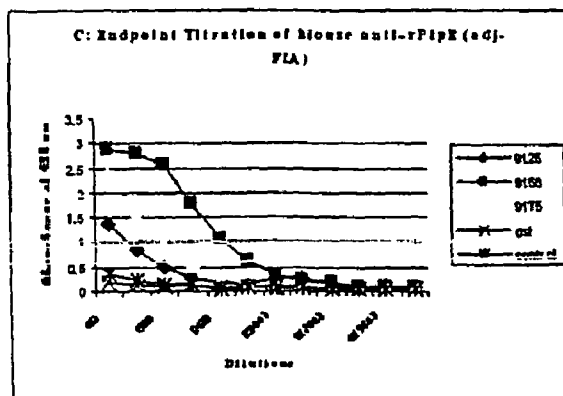
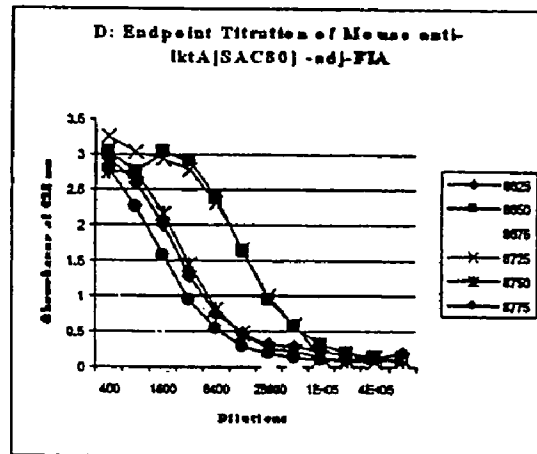
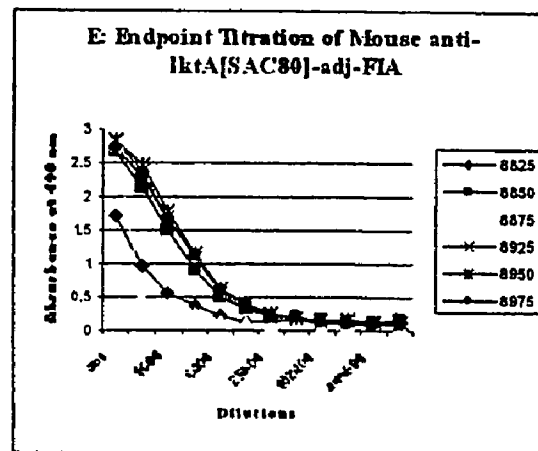


Figure 15 A-C

15D



15E



15F

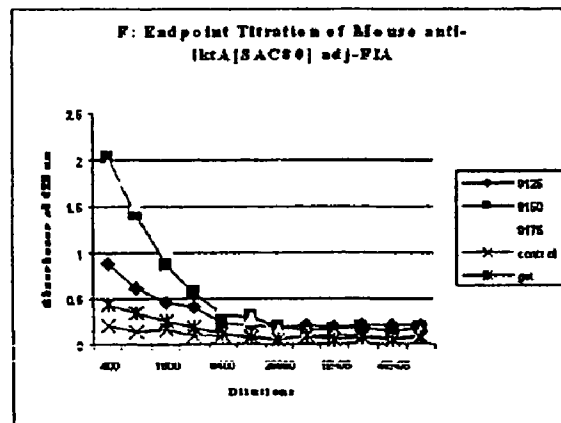


Figure 15 D-F

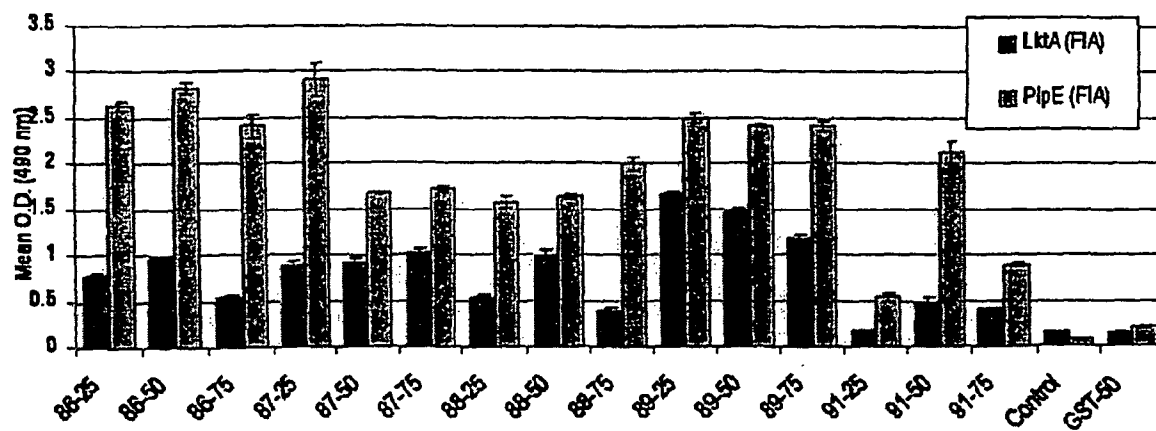
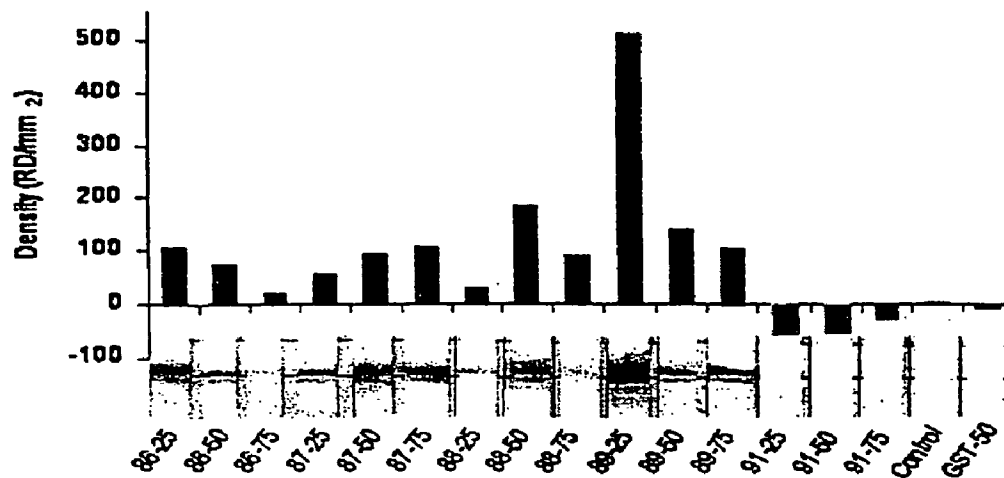


Figure 16

17A



17B

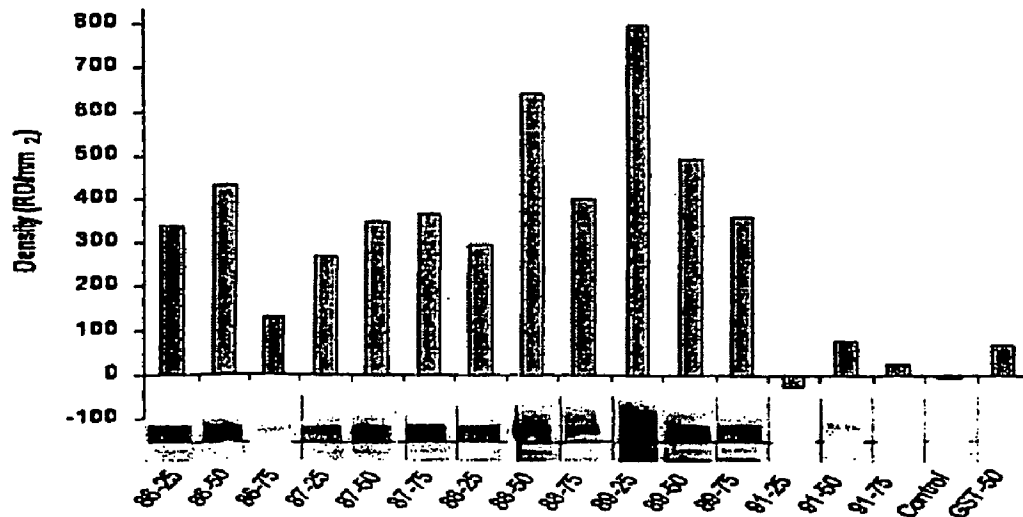


Figure 17 A-B

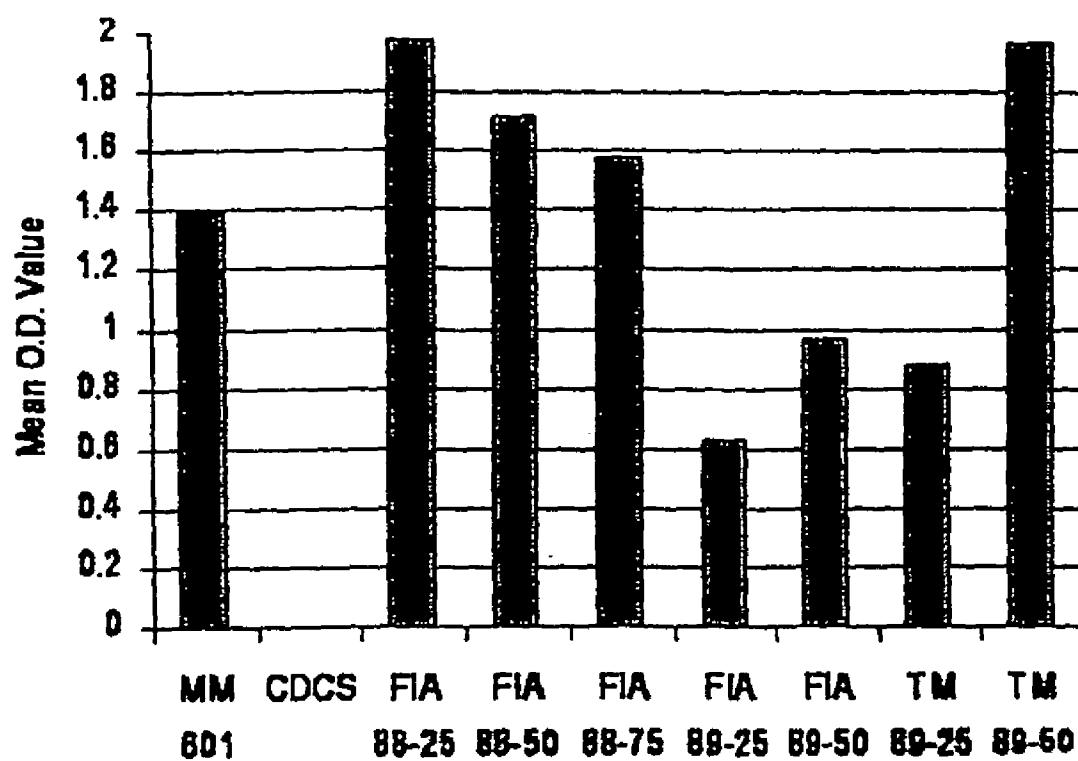


Figure 18

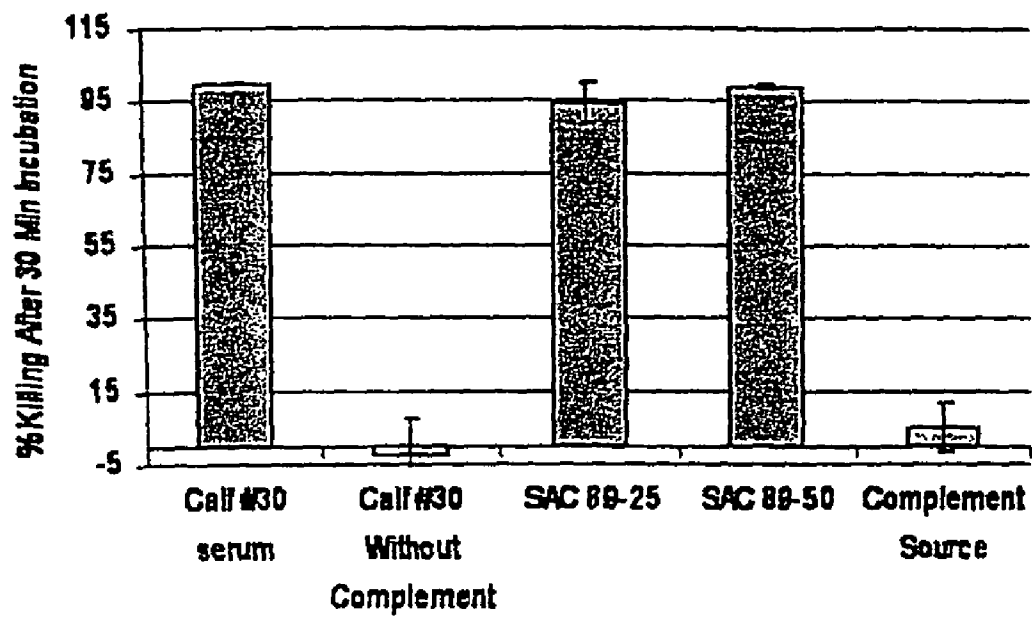
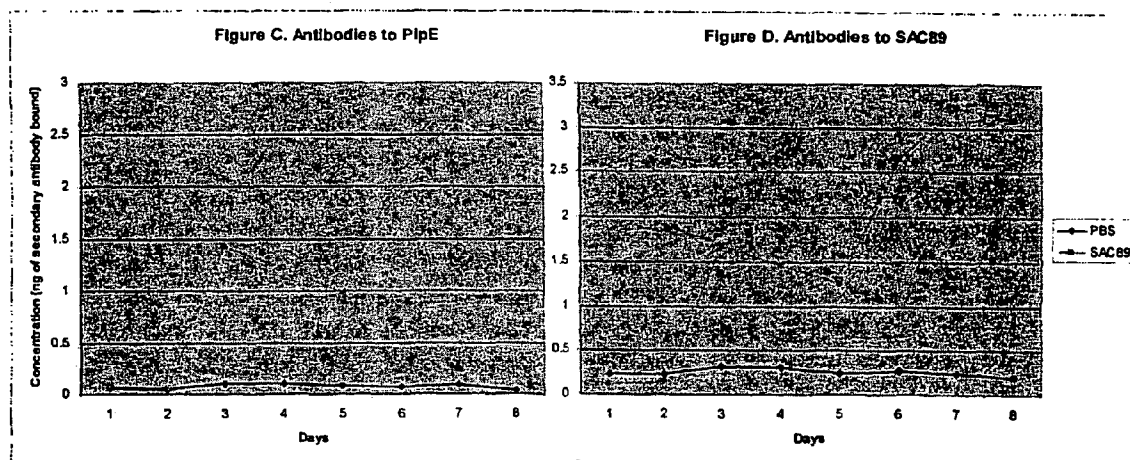
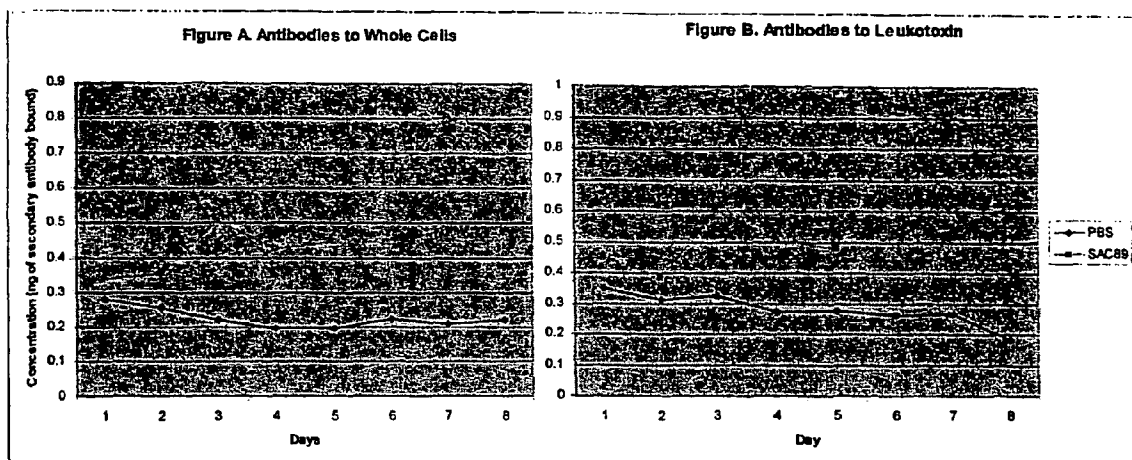


Figure 19



**MANNHEIMIA HAEMOLYTICA CHIMERIC
OUTER MEMBRANE PROTEIN PLPE AND
LEUKOTOXIN EPITOPES AS A VACCINE OR
VACCINE COMPONENT AGAINST SHIPPING
FEVER**

CROSS-REFERENCE TO RELATED
APPLICATIONS

This application claims benefit of U.S. provisional patent application 60/757,342, filed Jan. 9, 2006, and is a continuation-in-part of U.S. patent application Ser. No. 11/235,982, filed Sep. 27, 2005, now issued U.S. Pat. No. 7,144,580, which is a divisional of U.S. patent application Ser. No. 10/695,544, filed Oct 29, 2003, now abandoned, which claimed benefit of U.S. provisional patent application 60/422,305, filed Oct 30, 2002, the complete contents of each of which are hereby incorporated by reference.

This invention was made using funds from grants from the United States Department of Agriculture having grant number USDA-NRI 2002-35204-12250. The United States government may have certain rights in this invention.

BACKGROUND OF THE INVENTION

1. Field of the Invention

The present invention relates generally to the prevention of bovine respiratory disease (BRD) and, in particular, its most severe form, termed "shipping fever". More specifically, the present invention relates to the use of chimeric proteins comprising immunodominant epitopes of recombinant *Mannheimia haemolytica* outer membrane protein PlpE, and immunodominant epitopes of recombinant *M. haemolytica* leukotoxin as a vaccine or vaccine component against shipping fever.

2. Background

BRD is the major cause of beef cattle morbidity and mortality and of economic losses to the beef cattle industry. The cost of BRD to beef cattle producers approaches \$1 billion annually. BRD arises from the interaction of numerous contributing factors including physical stresses associated with weaning, shipment, inclement weather, and overcrowding coupled with viral and bacterial infections. The end result in severe cases is colonization of the lungs with pathogenic bacteria resulting in severe pneumonia. *Pasteurella multocida*, *Haemophilus somnus* and *Mannheimia* (formerly *Pasteurella*) *haemolytica* are associated with bovine pneumonia.

However, *Mannheimia haemolytica* serotype 1(S1) is by far the most important and commonly isolated bacterial pathogen in development of the often-fatal fibrinous pleuropneumonia in beef cattle known as pneumonic pasteurellosis or "shipping fever".

Prevention and control of shipping fever in feedlots is currently partially addressed by three different mechanisms: antibiotic treatment upon arrival of cattle at the feedlot, antibiotic therapy for sick cattle, and vaccination against BRD viruses and *M. haemolytica*. The extensive use of antibiotics to control shipping fever increases the possibility of antibiotic residues in meat and the development of drug-resistant bacteria in cattle, including those bacteria with potential impact on human health such as *Salmonella* and *Escherichia coli* 0157:H7.

Viral and bacterial vaccines for the control of shipping fever have been used for many years. Despite their availability, the disease continues to be a major bovine health problem. Attempts to develop a vaccine include the following:

U.S. Pat. No. 5,055,400 to Lo et al., (Oct. 8, 1991) discloses the *Pasteurella haemolytica* leukotoxin gene and protein.

U.S. Pat. No. 5,476,657 (Potter, Dec. 19, 1995) and U.S. Pat. No. 5,871,750 (Potter, Feb. 16, 1991) disclose vaccines comprising *Pasteurella haemolytica* leukotoxin or truncated forms of the leukotoxin.

U.S. Pat. No. 5,708,155 (Potter et al., Jan. 13, 1998) and U.S. Pat. No. 6,797,272 (Potter et al., Sep. 28, 2004) disclose vaccines comprised of chimeras of *Pasteurella haemolytica* leukotoxin and an antigen such as somatostatin (SRIF), gonadotropin releasing hormone (GnRH), or rotavirus viral protein 4 (VP4).

U.S. Pat. No. 5,238,823 (Potter et al., Aug. 24, 1993), U.S. Pat. No. 5,273,889 (Potter et al., Dec. 28, 1993), U.S. Pat. No. 5,594,107 (Potter et al., Jan. 14, 1997) and U.S. Pat. No. 6,096,320 (Potter et al., 5 Aug. 1, 2000), disclose vaccines of chimeric proteins comprising *Pasteurella haemolytica* leukotoxin or an antigenic fragment thereof, and a cytokine such as gamma-interferon or interleukin-2.

U.S. Pat. No. 5,871,750 (Potter, Feb. 16, 1999) discloses vaccine compositions comprising a truncated *Pasteurella haemolytica* leukotoxin. Other *Pasteurella haemolytica* cell surface antigenic proteins are also disclosed (fimbrial protein, plasmin receptor protein, and 50K outer membrane protein).

U.S. Pat. No. 5,723,129 (Potter et al., Mar. 3, 1998), U.S. Pat. No. 5,837,268 (Potter et al., Nov. 17, 1998), U.S. Pat. No. 6,022,960 (Potter et al., Feb. 8, 2000) and U.S. Pat. No. 6,521,746 (Potter et al., Feb. 18, 2003) disclose vaccines comprised of chimeras of leukotoxin and gonadotropin releasing hormone (GnRH) multimers.

U.S. Pat. No. 6,475,754 (Bemis et al., Nov. 5, 2002) discloses an antigenic chimeric protein comprising fimbrial protein of *Bordetella bronchiseptica* and leukotoxin of *M. haemolytica*.

Immunity against *M. haemolytica* is thought to be primarily through production of serum antibodies that neutralize the secreted leukotoxin (LKT) and antibodies against surface antigens. The specific surface antigens that are important in stimulating host immunity to *M. haemolytica* are not known; however, several studies point towards the importance of outer membrane proteins (OMPs). Pandher et al. demonstrated 21 surface-exposed immunogenic outer membrane proteins in *M. haemolytica* S1 using protease treatment and Western blotting. (Pandher K, Murphy G L, Confer A W. Identification of immunogenic, surface-exposed outer membrane proteins of *Pasteurella haemolytica* serotype 1. Vet Microbiol 1999; 65: 215-26) High antibody responses to outer membranes, as measured by ELISA, and to several specific OMPs, as measured by quantitative Western Blotting, consistently correlated with resistance to challenge with virulent *M. haemolytica* S1 (Confer A W, McCraw R D, Durham J A, Morton R J, Panciera R J. Serum antibody responses of cattle to iron-regulated outer membrane proteins of *Pasteurella haemolytica* A1. Vet Immunol Immunopathol 1995; 47: 101-10 and Mosier D A, Simons K R, Confer A W, Panciera R J, Clinkenbeard K D. *Pasteurella haemolytica* antigens associated with resistance to pneumonic pasteurellosis. Infect Immun 1989; 57: 711-6). Vaccination of cattle with OMP-enriched cellular fractions, from *M. haemolytica* S1 significantly enhanced resistance of cattle against experimental challenge in the absence of antibodies to LKT. (Morton R J, Panciera R J, Fulton R W, Frank G H, Ewing S A, Homer J T, Confer A W. Vaccination of cattle with outer membrane protein-enriched fractions of *Pasteurella haemolytica* and resistance against experimental challenge exposure. Am J Vet Res 1995; 56: 875-879) However, the

extraction procedure for bacterial outer membranes is time consuming and expensive, making use of purified OMPs as a component of a *M. haemolytica* vaccine impractical due to cost. Thus, it can be appreciated that the identification of specific, surface exposed immunogenic *M. haemolytica* OMPs that would stimulate strong antibody responses is highly desirable. Cloning and expression of the appropriate gene(s) and production of recombinant OMP could then be achieved inexpensively.

One of the *M. haemolytica* OMPs to which high antibody responses correlated with resistance against experimental challenge is a major 45 kDa OMP. Prior studies were undertaken to clone and characterize that protein. In 1998, Pandher et al. reported the cloning, sequencing and characterization of the gene for the major 45-kDa *M. haemolytica* S1 outer membrane lipoprotein, designated PlpE. (Pandher K, Confer A W, Murphy G L. Genetic and immunologic analyses of PlpE, a lipoprotein important in complement-mediated killing of *Pasteurella haemolytica* serotype 1. Infect Immun 1998; 66: 5613-9, which publication is incorporated herein by reference) PlpE was found genetically to have 32-35% similarity to an immunogenic lipoprotein, OmlA, demonstrated in *Actinobacillus pleuropneumoniae* serotypes 1 and 5. Affinity-purified, anti-PlpE antibodies recognized an OMP in all serotypes of *M. haemolytica* except in serotype 11. In addition, PlpE was determined to be surface-exposed, and in complement-mediated killing assays, a significant reduction was observed in killing of *M. haemolytica* when bovine immune serum that was depleted of anti-PlpE antibodies was used as the source of antibody, suggesting that antibodies against PlpE may contribute to host defense against the bacterium.

Because of the economic constraints of the cattle industry, bovine vaccines must be low in cost. Therefore, current *M. haemolytica* vaccines are crude, usually consisting of a culture supernatant, which contains *M. haemolytica* leukotoxin and sloughed surface proteins, and/or the killed bacterium. Perino and Hunsaker reviewed published field studies on commercial *M. haemolytica* vaccines and found that efficacy could be established in only 50% of the trials. (Bov Practitioner 1997; 31: 59-66). There is thus an ongoing need for improvement in *M. haemolytica* vaccines, and for the development of improved methods and compositions for protecting cattle against shipping fever.

SUMMARY OF THE INVENTION

In connection with the present invention, the gene for *M. haemolytica* outer membrane protein PlpE was cloned and the recombinant PlpE (rPlpE) was purified and used in immunological and vaccination studies. It was discovered that adjuvanted rPlpE was highly immunogenic in cattle, and vaccination of cattle with 100 µg of rPlpE markedly enhanced resistance against experimental challenge with virulent *M. haemolytica*. It was also discovered that the addition of rPlpE to a commercial *M. haemolytica* vaccine significantly enhanced ($p < 0.05$) protection afforded by the vaccine against experimental challenge.

Thus, in one aspect of the present invention there are provided vaccine compositions comprising rPlpE or conservatively modified variants thereof separately or which may optionally be combined with adjuvant to enhance the protection efficacy of vaccine preparations against BRD and/or shipping fever, wherein the vaccine composition further comprises a pharmaceutically acceptable carrier or diluent. The rPlpE also may optionally be combined with other immunogens and/or existing commercially available vaccines to form an augmented vaccine composition, wherein the vaccine

composition further comprises a pharmaceutically acceptable carrier or diluent and adjuvant.

In another aspect of the invention there are provided methods for inducing an immune response in cattle to provide immune protection against BRD and/or shipping fever, the method comprising administering to an at-risk animal an effective amount of a vaccine composition comprising rPlpE or conservatively modified variants thereof alone or in combination with an adjuvant and/or other immunogens to provide a means to reduce the risk of BRD, wherein the vaccine composition further comprises a pharmaceutically acceptable carrier or diluent.

Most of the structure of an OMP molecule would play no significant role in inducing protective immune responses, because extended portions of the molecule are buried, unexposed, in the outer membrane. Instead, immunity can be attributed to only short, surface-exposed epitopes of these proteins. Identification of such surface-exposed epitopes as protective antigens in animal models has been the target of peptide vaccine design strategies for various pathogenic bacteria. Because of *M. haemolytica* PlpE's potential as an important immunogen, studies were undertaken to characterize surface-exposed and immunologically important epitopes of PlpE and to produce and test recombinant epitopes corresponding thereto. Thus, in another aspect of the invention there are provided immunologically important epitopes of rPlpE for use in vaccines and related methodologies.

In another aspect of the invention, chimeric proteins are provided which contain one or more copies of an immunodominant epitope of *M. haemolytica* rPlpE in combination with an immunodominant epitope of *M. haemolytica* leukotoxin.

The present invention provides an immunogenic composition which includes at least one chimeric protein comprising: one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE, and one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* leukotoxin (LKT); and a physiologically compatible carrier. In one embodiment, the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE is R2 and is represented by SEQ ID NO: 19. In one embodiment, the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT is mLKT A and is represented by SEQ ID NO: 21. In one embodiment, the at least one chimeric protein further comprises a leader sequence, for example, the glutathione-S-transferase leader sequence. The at least one chimeric protein may further comprises one or more spacer peptides. In one embodiment of the invention, the at least one chimeric protein comprises two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE and two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT.

The invention further provides a chimeric protein which comprises: one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE, and one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* leukotoxin (LKT). In one embodiment, the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE is R2 as represented by SEQ ID NO: 19. In one embodiment, the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT is mLKT A as represented by SEQ ID NO: 21. In another embodiment, the chimeric protein further comprises a leader sequence, which may be, for example, the glutathione-S-transferase leader sequence. In another embodiment, the chimeric protein comprises one or more spacer peptides. In one embodiment of the invention, the chimeric protein comprises

two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE and two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT.

The invention also provides a vaccine preparation which comprises at least one chimeric protein comprising: one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE, and one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* leukotoxin (LKT); and a physiologically compatible carrier. In one embodiment, the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE is R2 as represented by SEQ ID NO: 19. In one embodiment of the invention, the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT is mLKT A as represented by SEQ ID NO: 21. In another embodiment, the at least one chimeric protein further comprises a leader sequence, one example of which is the glutathione-S-transferase leader sequence. In another embodiment, the at least one chimeric protein further comprises one or more spacer peptides. In other embodiments, the at least one chimeric protein comprises two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE and two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT.

In yet other embodiments, the vaccine preparation of also includes an adjuvant.

The present invention also provides a method of eliciting an immune response to *Mannheimia haemolytica* in a mammal. The method comprises the step of administering to the mammal at least one chimeric protein comprising: one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE, and one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* leukotoxin (LKT). In one embodiment, the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE is R2 as represented by SEQ ID NO: 19. In one embodiment of the invention, the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT is mLKT A as represented by SEQ ID NO: 21. In another embodiment, the at least one chimeric protein further comprises a leader sequence, one example of which is the glutathione-S-transferase leader sequence. In another embodiment, the at least one chimeric protein further comprises one or more spacer peptides.

In another embodiment, the at least one chimeric protein comprises two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE and two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT. In one embodiment of the invention, the mammal is bovine.

The invention also provides a method of vaccinating cattle to prevent or attenuate disease symptoms caused by *Mannheimia haemolytica*. The method comprises the step of administering to the cattle at least one chimeric protein comprising: one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE, and one or more copies of an immunodominant epitope of recombinant *Mannheimia haemolytica* leukotoxin (LKT). According to the method, the at least one chimeric protein is administered in an amount sufficient to prevent or attenuate disease symptoms caused by *Mannheimia haemolytica*. In one embodiment, the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE is R2 as represented by SEQ ID NO: 19. In one embodiment of the invention, the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT is mLKT A as represented by SEQ ID NO: 21. In another

embodiment, the at least one chimeric protein further comprises a leader sequence, one example of which is the glutathione-S-transferase leader sequence. In another embodiment, the at least one chimeric protein further comprises one or more spacer peptides. In addition, in one embodiment, the at least one chimeric protein comprises two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* PlpE and two copies of the immunodominant epitope of recombinant *Mannheimia haemolytica* LKT.

A better understanding of the present invention, its several aspects, and its advantages will become apparent to those skilled in the art from the following detailed description, taken in conjunction with the attached figures, wherein there is described the preferred embodiment of the invention, simply by way of illustration of the best mode contemplated for carrying out the invention.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a graph depicting anti-PlpE antibody response of 6 cattle vaccinated with 100 µg of rPlpE on day 0.

FIG. 2 is a bar graph depicting anti-PlpE antibody responses of cattle that spontaneously seroconverted to *M. haemolytica* (Natural Infection), vaccinated with commercial vaccines, *M. haemolytica* outer membranes (OMP), or live *M. haemolytica*.

FIG. 3 is a bar graph depicting anti-PlpE antibodies for cattle vaccinated with commercial *M. haemolytica* vaccines or 100 µg of rPlpE.

FIG. 4 A-C is a series of graphs depicting anti-PlpE (A), anti-*M. haemolytica* leukotoxin (B), and anti-*M. haemolytica* whole cells (C) in cattle vaccinated with PRESPONSE, PRE-SPONSE plus 100µg of rPlpE, or nonvaccinated.

FIG. 5 is a graph depicting rectal temperatures of calves after challenge.

FIG. 6 is a graphical depiction of bovine antibody against surface exposed components of PlpE that was affinity purified with intact *M. haemolytica* cells and used to probe a peptide array. Densitometric analysis demonstrated a total of 8 distinct antigenic regions (E1-8) in PlpE with E2 being the largest and E4 having the highest densitometric signal.

FIG. 7 A-B. A, amino acid sequence of R2 immunodominant region (epitope) of rPlpE (SEQ ID NO: 19); B, DNA sequence encoding R2 (SEQ ID NO: 20).

FIG. 8 A-B. A, amino acid sequence of mLKTA (SEQ ID NO: 21); B, DNA sequence encoding mLKTA (SEQ ID NO: 22).

FIG. 9 A-F. Schematic representations of plasmids encoding the chimeric proteins of the invention. A, plasmid encoding SAC86; B, plasmid encoding SAC 87; C, plasmid encoding SAC88; D, plasmid encoding SAC89; E, plasmid encoding SAC91; F, linearized versions of the plasmids.

FIG. 10 A-B. A, amino acid sequence of SAC86 chimera (SEQ ID NO: 23). Sequences of spacer peptides are shown in italics; sequences from PlpE are shown in bold with a solid underline; and sequences from LKT are shown in bold with a dotted underline; B, nucleic acid sequence encoding SAC86 chimera (SEQ ID NO: 24).

FIG. 11 A-B. A, amino acid sequence of SAC87 chimera (SEQ ID NO: 25). Sequences of spacer peptides are shown in italics; sequences from PlpE are shown in bold with a solid underline; and sequences from LKT are shown in bold with a dotted underline; B, nucleic acid sequence encoding SAC87 chimera (SEQ ID NO: 26).

FIG. 12 A-B. A, amino acid sequence of SAC88 chimera (SEQ ID NO: 27). Sequences of spacer peptides are shown in italics; sequences from PlpE are shown in bold with a solid

underline; and sequences from LKT are shown in bold with a dotted underline; B, nucleic acid sequence encoding SAC88 chimera (SEQ ID NO: 28).

FIG. 13 A-B. A, amino acid sequence of SAC89 chimera (SEQ ID NO: 29); B, nucleic acid sequence encoding SAC89 chimera (SEQ ID NO: 30). For both A and B, sequences of spacer peptides are shown in italics; sequences from PlpE are shown in bold with a solid underline; and sequences from LKT are shown in bold with a dotted underline. FIA=Freund's Incomplete Adjuvant; TM=TiterMax®.

FIG. 14 A-B. A, amino acid sequence of SAC91 chimera (SEQ ID NO: 31). Sequences of spacer peptides are shown in italics; sequences from PlpE are shown in bold with a solid underline; and sequences from LKT are shown in bold with a dotted underline; B, nucleic acid sequence encoding SAC91 chimera (SEQ ID NO: 32).

FIG. 15 A-F. Data from endpoint titrations of mouse antibodies against rPlpE and LKTA. A, using 25, 50 or 75 µg of SAC86 and SAC87 proteins; B, using 25, 50 or 75 µg of SAC88 and SAC89 proteins.

FIG. 16. Summary of antibody responses to PlpE and LKT via single dilution ELISA.

FIG. 17 A-B. Quantification of Western blot data using densitometric analysis. A, blots against LKT; B, blots against PlpE.

FIG. 18. LKT neutralization activity of murine anti-chimeric immune sera.

FIG. 19. Bactericidal activity of murine anti-SAC89 hyper-immune sera compared to anti-PlpE hyper-immune calf serum.

FIG. 20 A-D. Antibody responses of cattle vaccinated with PBS plus adjuvant or 100 µg of SAC89 plus adjuvant. Data are expressed as nanograms of immunoglobulin binding to the antigen in ELISAs. A, antibodies to whole cells; B, antibodies to Leukotoxin; C, antibodies to PlpE; D, antibodies to SAC89.

DETAILED DESCRIPTION OF THE INVENTION

The invention is not limited in its application to the details of the embodiments and steps described herein. The invention is capable of other embodiments and of being practiced or carried out in a variety of ways. It is to be understood that the phraseology and terminology employed herein is for the purpose of description and not of limitation.

In accordance with the present invention there are provided new vaccine preparations against BRD and shipping fever through the use of discrete recombinant PlpE and subunits of rPlpE containing immunoprotective regions. In one aspect, only rPlpE or immunoprotective and functional regions thereof are utilized as the antigenic component of the vaccine. In another aspect, rPlpE or subunits thereof are utilized in combination with other antigen components, such as leukotoxin (LKT).

In yet another aspect, the invention provides chimeras (both proteins and the nucleic acids that encode them) of the major surface-exposed epitope of PlpE (designated herein as either "E2" or "R2") and a minimal leukotoxin A fragment ("mLKTA"). In some embodiments, the chimeras include a leader sequence such as the GST leader sequence. In some embodiments, the antigenic regions (R2 and MLKTA) are present in multiple copies in the chimera., the multiple copies being separated by spacer peptides.

Another aspect of the present invention relates to methods useful to reduce the risk of BRD and shipping fever in cattle and prevent or attenuate biological transmission of the disease among cattle populations.

The experiments described and non-limiting examples provided hereinafter demonstrate that cattle immunized with rPlpE and/or epitopes thereof, or with chimeric proteins that include the immunodominant PlpE regions and mLKTA epitopes (particularly in multiple copies) are unexpectedly better protected against infection following challenge with *M. haemolytica* than cattle immunized with existing commercially available vaccines.

EXAMPLE 1

Immunogenicity of rPlpE and Vaccine Preparation

Studies were undertaken to determine the immunogenicity of outer membrane lipoprotein PlpE from *M. haemolytica* S1, to determine if commercial vaccines stimulate antibodies to it, and examine if addition of recombinant PlpE to a commercial *M. haemolytica* vaccine would augment vaccine-induced immunity.

1. Materials and Methods

1.1. Bacterial Culture.

M. haemolytica S1 Oklahoma Strain was used for serology antigen preparation and for challenge of animals. Frozen stock cultures were plated onto brain-heart infusion (BHI) and grown at 37° C. in a 5% CO₂ environment for 18 hours. An isolated colony from each was propagated in 10 ml BHI broth with rotatory shaking at 120 oscillations/min. for 18 hours at 37° C. 100 µl of suspension was added to 1 L of BHI broth and grown overnight. The bacteria were sedimented by centrifugation at 6000 ×g for 15 minutes, washed in 125 ml sterile phosphate buffered saline (PBS) and re-centrifuged as above 6000 ×g for 15 minutes. The bacteria were re-suspended in PBS and adjusted spectrophotometrically to a final concentration of approximately 1.0×10⁹ CFU/ml (optical density of A₆₀₀=0.65).

1.2. Cloning and Purification of PlpE

The truncated form of plpE lacking the sequence encoding the putative signal peptide was amplified from pB4522 (Pandher et. al., 1998, supra) with the help of a forward primer starting 58 nucleotides into the 5'-end and priming into the open reading frame of plpE and a reverse primer which is complementary to the 3'-end of the gene. The amplimer was cut with BamHI and HindIII and ligated into an expression vector, pRSETA, cut with the same restriction enzymes. Competent *E. coli* DH5α were transformed with the ligation mixture and transformants were plated on Luria-Bertani (LB) agar plates with 50 µg/ml of ampicillin. Transformants were screened and appropriate subclones were identified. Plasmid DNA isolated from such subclones was submitted to the Oklahoma State University Core Facility where the nucleotide sequence was determined by the ABI Model 3700 (BioSciences) automated DNA sequencing system (SEQ ID NO: 1). Once the nucleotide sequence of a representative subclone was compared to that deposited in the GenBank (AF059036), the recombinant plasmid was introduced into BL21 (DE3) pLysS by transformation to express and purify rPlpE (SEQ ID NO: 2).

The expression of rPlpE was done according to the protocol recommended by the manufacturer of the vector and the expression host (Invitrogen, Calif.). Briefly, single colonies of BL21 (DE3) pLysS harboring the truncated plpE in pRSETA, were inoculated into appropriate volumes of LB broth with 50 µg/ml ampicillin and 34 µg/ml chloramphenicol. The culture was incubated at 37° C. until A₆₀₀=0.5 was attained at which time the synthesis of the recombinant pro-

tein was induced by adding IPTG (1 mM final concentration) and the induction was continued for at least 3 hours. In order to purify rPlpE, the culture was harvested and lysed by sonication. The cellular debris was then removed by centrifugation and the recombinant protein was loaded onto an affinity column packed by Pro-Bond nickel-chelating resin that selectively binds recombinant proteins with 6 histidine residues (His-Tag) at either the N- or carboxy-terminus. In this instance, the His-Tag is at the N-terminus. The recombinant protein bound to the resin was then eluted with either a low pH buffer or competition with imidazole.

The purity of each preparation was determined by SDS-PAGE followed by Coomassie stain and Western blot with murine anti-PlpE ascites fluid.

1.3... Serology

Antibodies to formalin-killed *M. haemolytica* whole bacterial cells (WC), to LKT, and to rPlpE were determined by enzyme-linked immunosorbent assay (ELISA). For WC preparation, *M. haemolytica* S1 were prepared from a washed 24 hour culture by suspending cells in 0.4% formalinized saline at a concentration determined spectrophotometrically to be 1.850 OD₆₅₀. LKT was prepared from supernatant from a 3-hour culture of *M. haemolytica* S1 grown in RPMI-1640 medium at 37° C. in a shaking incubator. The LKT was partially purified by precipitation with 40-60% ammonium sulfate. The precipitate was resuspended in 3M guanidine containing 59 mM NaHPO₄ and 100 mM NaCl. By SDS-PAGE of the LKT preparation, one intensely staining band was identified at 105 kDa and confirmed to be LKT on a Western blot using an anti-LKT monoclonal antibody. Leukotoxin activity was 10⁻⁴ LKT units per ml. The 2-keto-3-deoxyoctonate concentration was 7.5 µg per mg of protein.

Wells of 96-well microtiter plates were coated with WC at an optical density reading equivalent to 10⁸ CFU of a 24-hour culture, with LKT at 50 ng per well, or with rPlpE at 50 ng per well. Sera were diluted in PBS-Tween 20 containing 1% BSA. ELISA for detection of serum antibodies to PlpE was done in the first immunogenicity study using serum dilutions ranging from 1:400-1:819,200. Otherwise, sera were tested against various antigens at dilutions of 1:800 for WC, 1:1600 for LKT, and 1:1600 for rPlpE, which were in the linear range of established dilution curves. The extent of antibody binding was detected using a 1:400 dilution of horseradish peroxidase-conjugated, affinity purified rabbit anti-bovine IgG (Kirkegaard & Perry Laboratories, Gaithersburg, Md.). Antibody responses are expressed as ng of immunoglobulin binding based on a set of IgG standards on each plate.

1.4. Animals

A total of 82 normal healthy beef calves (Hereford or Angus/Hereford cross) of mixed sex were used. The calves were weaned at around 6-8 months of age. All calves were vaccinated with 7-way Clostridial vaccine and leptospiral vaccine, and treated with anthelmintic 30 days prior to the study. The calves received free choice native grass hay supplemented with grain ration throughout the study. All animal studies were done following using protocols approved the University Institutional Animal Care and Use Committee (Protocol #182).

1.5. Anti-PlpE Responses with *M. haemolytica* Vaccines

To determine if vaccination of cattle with commercial or experimental *M. haemolytica* vaccines stimulate anti-PlpE antibodies, two studies were done. The first experiment was a retrospective study using sera from 18 cattle from previous vaccine studies. Serum antibodies to PlpE were determined on samples from the day of vaccination (day 0) and from day

14. On day 0, three calves each were vaccinated subcutaneously with one of the following commercial vaccines: *P. haemolytica* Toxoid, BRSV-BVD-IBR-PI₃ Vaccine (PYRAMID™ 4/PRESPONSE®; Fort Dodge Laboratories), *P. haemolytica-multocida* Bacterin-Toxoid (PULMO-GUARD™ PH-M, Boehringer Ingelheim), *P. haemolytica-multocida-Salmonella typhimurium* Bacterin-Toxoid (POLY-BAC B® 1, Texas Vet Labs). Three calves were each vaccinated with 2 mg of an *M. haemolytica* outer membrane preparation in Freund's incomplete adjuvant or 10⁹ CFU of live *M. haemolytica*. In addition, sera were analyzed from three non-vaccinated calves that spontaneously seroconverted to *M. haemolytica* based on positive antibody responses to WC and LKT.

The second vaccine experiment was a prospective study designed to follow the anti-PlpE antibodies for 42 days after a single dose of a commercial *M. haemolytica* vaccine or rPlpE. Thirty calves were divided equally among 6 groups and vaccinated subcutaneously once each on day 0 with PRE-SPONSE®, *P. haemolytica* Bacterin-Toxoid (ONE SHOT™, Pfizer), an avirulent *M. haemolytica* culture (ONCE PMH®, Intervet), PULMO-GUARD™ PH-M, or 100 µg of rPlpE in commercial adjuvant (Pfizer). Five unvaccinated calves served as controls. Sera were obtained on days 0, 7, 14, 21, 28, and 42, and antibodies to WC, LKT and PlpE were determined.

1.6. Recombinant PlpE Immunogenicity Studies

To determine if rPlpE was immunogenic, one calf each was vaccinated once with either 10, 50, or 100 µg of rPlpE in a commercial proprietary adjuvant (Pfizer Inc, Lincoln, Nebr.). One calf remained as a non-vaccinated control. Sera were obtained 21 days after vaccination and evaluated for end-point antibody titers against rPlpE using serial 2-fold dilutions. Twenty-four days after the initial vaccination, each calf and a non-vaccinated calf were transthoracically challenged with 5.0x10⁹ CFU of live *M. haemolytica* from an overnight culture in accordance with established procedures. Four days later, calves were humanely killed, and lung lesion scores determined on a 20-point scale.

In a second cattle experiment, 6 cattle were vaccinated with 100 µg of rPlpE in commercial adjuvant on day 0 and 6 calves remained as non-vaccinated controls. On day 21, all cattle were challenged intrathoracically with 1x10⁹ CFU of virulent *M. haemolytica*. Calves were humanely killed on day 25, and lung lesion scores determined. Antibody responses against rPlpE and *M. haemolytica* WC were determined on days 0, 7, 14 and 21 after vaccination.

In a third cattle experiment, PRESPONSE® was obtained from the manufacturer, and 18 weanling beef steers were equally allocated among the following vaccine groups: Group 1-PRESPONSE, Group 2-PRESPONSE+100 µg PlpE, and Group 3-non-vaccinated. Cattle were vaccinated on day 0 with 2 ml of PRESPONSE (manufacturer's recommended dosage) or 2 ml of PRESPONSE mixed with 0.5 ml of PlpE (100 µg). Antibody responses to *M. haemolytica* WC, rPlpE or to LKT were determined by ELISA on days 0, 7, 15, and 23. On day 24, cattle in Groups 1, 2, & 3 were challenged transthoracically with 3.0x10⁹ CFU of *M. haemolytica*. Four days later, calves were humanely killed, and lung lesion scores determined.

1.7. Statistical Analysis

Mean rectal temperatures, antibody responses and lesion scores among the various groups were compared by Student's t tests. Mean rectal temperatures and antibody responses within groups were compared by paired t tests. Differences were considered significant when p<0.05. Linear regression

analyses were done to determine if there was a significant correlation between antibody response and lesion score.

2. Results

2.1. Recombinant PlpE Immunogenicity

In the first immunogenicity experiment that determined end-point anti-rPlpE titers in response to various doses of rPlpE, serum from the non-vaccinated calf had an end-point antibody titer of 1:400 against rPlpE. Sera from the 10, 50, and 100 µg vaccinates had titers of 1: 12,800, 1: 25,600, and 1: 25,600, respectively. Intrathoracic challenge of those calves with virulent *M. haemolytica* resulted in a lesion score of 15.5 (20 maximum severity) for the non-vaccinated control calf. Lesion scores for the 10, 50, and 100 µg-vaccinates were 4.5, 3.0, and 3.5 respectively.

In the second immunogenicity experiment, vaccination with rPlpE on day 0 stimulate a significant increase in antibodies to rPlpE and to *M. haemolytica* WC on day 7 (FIG. 1). Those responses continued to increase to a maximum on day 20 and declined insignificantly on day 25, whereas antibodies to rPlpE and to WC failed to increase for the nonvaccinated calves.

Anti-LKT antibodies did not significantly increase for either the rPlpE-vaccinated or control groups (data not shown). Mean lesion scores (standard deviation) after challenge were 7.0±3.8 for nonvaccinated controls and 4.1±3.0 for the rPlpE vaccinates, a 41.4% reduction in lesion scores. Those differences were significant at the level of $p=0.07$. When data from the first experiment were combined with these data, the mean lesion score for nonvaccinated controls was 8.2±4.7 and mean lesion score for PlpE vaccinates was 3.9±2.6 ($p<0.05$), a 52.1% reduction in lesion scores.

2.2. *M. haemolytica* Vaccines

In the first vaccine experiment, vaccination of calves with commercial vaccines, *M. haemolytica* outer membranes, and live *M. haemolytica* resulted in a nonsignificant increase in antibodies to PlpE (FIG. 2). In contrast, natural exposure to *M. haemolytica*, as indicated by spontaneous seroconversion, resulted in a significant increase in anti-PlpE antibodies. All vaccine-induced responses and natural exposure were substantially less than the antibodies produced in a calf vaccinated with 100 µg of rPlpE in commercial adjuvant. There were no significant differences among the antibody responses to rPlpE on day 14 for any of the commercial vaccine, live *M. haemolytica* vaccinated, or natural exposure groups. Antibody responses to *M. haemolytica* LKT and WC significantly increased for PULMOGUARD- and the live bacteria-vaccinated and natural exposure calves, whereas vaccination with outer membranes stimulated a significant antibody response to WC and vaccination with POLY-BAC and PRESPONSE failed to stimulate significant antibody responses to either *M. haemolytica* antigen (data not shown).

In the second vaccine experiment, vaccination of calves with one of four commercial *M. haemolytica* vaccines resulted in nonsignificant increases in antibodies to PlpE (FIG. 3).

Vaccination of calves with 100 µg of rPlpE in commercial adjuvant stimulated a significant increase in antibody responses to PlpE by day 7. That response continued to increase until it peaked on day 21 after vaccination. Vaccination with each commercial vaccine and with rPlpE resulted in significant increases in antibodies to *M. haemolytica* WC by day 7 (ONE SHOT and PRESPONSE) and by day 14 (ONCE PMH, PULMOGUARD, and rPlpE) (FIG. 2). Those responses remained significantly increased through day 14 (ONCE PMH and PRESPONSE) and day 42 (ONE SHOT,

PULMOGUARD, and rPlpE). Peak antibody responses for ONE SHOT-vaccinated cattle were significantly greater than peak responses for ONCE PMH, PRESPONSE or rPlpE vaccinates. Although antibody responses to LKT increased after vaccination with each commercial vaccine, only the responses initiated by PULMOGUARD and ONE SHOT were significantly increased beginning on day 7 through day 28. Anti-LKT antibodies did not increase for the rPlpE vaccinates. Peak anti-LKT antibody responses for PULMOGUARD-vaccinated cattle were significantly greater than peak responses for ONCE PMH, PRESPONSE or rPlpE vaccinates, whereas peak anti-LKT antibody responses for ONE SHOT-vaccinated cattle were significantly greater than peak responses for PRESPONSE vaccinates.

2.3. Augmentation of Commercial Vaccine with rPlpE

Because vaccination with commercial *M. haemolytica* vaccines stimulated low antibody responses to rPlpE, we investigated the augmentation of a commercial vaccine with rPlpE. Vaccination with PRESPONSE stimulated a significant increase in anti-rPlpE antibodies on day 15. Those responses, however, were not significantly different than were antibody responses of the nonvaccinated control calves on days 7, 15, and 23 (FIG. 4 A-C). PRESPONSE-rPlpE vaccination stimulated a significant increase in anti-rPlpE antibodies on days 7, 15 and 23, and those responses were significantly higher than responses for the PRESPONSE-vaccinated or nonvaccinated control calves. Anti-WC and anti-LKT responses were significantly increased on days 7 and 15 for the PRESPONSE- and PRESPONSE-rPlpE vaccinates. Those responses were not significantly different between those groups, whereas they were significantly greater than were anti-WC and anti-LKT antibody values for the nonvaccinated control group.

Rectal temperatures were taken on the day of challenge (day 24) and for the next 3 days (FIG. 5). Rectal temperatures remained essentially normal for all cattle except for the non-vaccinated Control group. In that group, rectal temperatures significantly increased on days 25 and 26, declining insignificantly on day 27. On days 26 and 27, mean rectal temperatures for the nonvaccinated Control group were significantly greater than for either the PRESPONSE or PRESPONSE/rPlpE groups. At necropsy, mean lung lesion scores were 7.9±3.6 for nonvaccinated controls, 3.0±1.3 for PRESPONSE-vaccinates (62.0% reduction in lesion score), and 1.1±0.9 for PRESPONSE/rPlpE vaccinates (86.1 % reduction in lesion scores). Differences between the PRESPONSE and Control and PRESPONSE/PlpE and Control lesion scores were significant. In addition, mean lesion score for the PRESPONSE/PlpE group was significantly lower than for the PRESPONSE group. There was a significant correlation ($r=-0.598$, $p<0.01$) between high serum antibody responses to rPlpE at day 23 and low lesion scores.

3. Discussion

The foregoing studies demonstrate that rPlpE is highly immunogenic for cattle and that vaccination with rPlpE can greatly enhance resistance against experimental challenge with the bacterium. The in vivo studies definitively indicate that anti-PlpE antibodies can contribute to host defense against *M. haemolytica* infection.

Vaccination of cattle with commercial *M. haemolytica* vaccines, live *M. haemolytica* or outer membranes or after prior natural exposure stimulated low antibody responses to PlpE. For those vaccines, the rise in antibodies to rPlpE as measured on various days were not significant, and even those vaccines that stimulated high antibodies to *M. haemolytica* WC and LKT still stimulated low anti-rPlpE response. Commercial vaccine-induced anti-rPlpE antibody responses were sub-

stantially lower than those stimulated by vaccination with 100 µg of rPlpE in a commercial adjuvant. This was not unexpected, because commercial vaccines vary greatly in their composition in that some are composed of culture supernatants and bacterial cell components, others contain whole bacterial cells, and one is a live mutant. A somewhat surprising finding was that calves previously vaccinated with *M. haemolytica* outer membranes in Freund's incomplete adjuvant had low antibody responses to rPlpE on day 14 (see Morton et al., supra). Therefore, although PlpE is a major outer membrane protein, its concentrations in commercial and experimental vaccines are most likely low and variable. In addition, the adjuvant used may play an important role in stimulating antibodies to PlpE.

Because commercial vaccines stimulated low antibodies to PlpE, we used rPlpE to augment the antibody response of a commercial vaccine, PRESPONSE, and demonstrated that PRESPONSE/PlpE stimulated greater protection against challenge than did PRESPONSE alone. Conlon et al. previously demonstrated that addition of recombinant LKT enhanced the efficacy of a culture supernatant vaccine and decreased clinical signs and pneumonic lesions. (Conlon J A, Shewen P E, Lo R Y. Efficacy of recombinant leukotoxin in protection against pneumonic challenge with live *Pasteurella haemolytica* A1. Infect Immun 1991; 59:587-91) Therefore, addition of one or more recombinant proteins to a *M. haemolytica* vaccine could be used by animal health companies to provide better products for protection of cattle against shipping fever.

In a recent survey, researchers found that of the *M. haemolytica* isolates from bovine respiratory disease from upper Midwestern United States were 60% A1, 26% A6 and 7% A2 with the remaining isolates from A9, A11 and untypable strains. (Al-Ghamdi G M, Ames T R, Baker J C, Walker R, Chase C C, Frank G H, Maheswaran S K. Serotyping of *Mannheimia* (*Pasteurella*) *haemolytica* isolates from the upper Midwest United States. J Vet Diagn Invest 2000; 12: 576-8) In another study, 60% of *M. haemolytica* isolates from cattle in a Texas feedyard were A1, whereas 40% were serotypes A2, A6, or A5 (Purdy C W, Raleigh R H, Collins J K, Watts J L, Straus D C. Serotyping and enzyme characterization of *Pasteurella haemolytica* and *Pasteurella multocida* isolates recovered from pneumonic lungs of stressed feeder calves. Curr Microbiol 1997; 34: 244-9). Therefore, although serotype 1 is the most common isolate from shipping fever, other serotypes play a role in the disease. Currently available *M. haemolytica* vaccines contain serotype 1 exclusively and therefore may or may not provide efficacious immunity against other serotypes. Cross serotype protection as stimulated by outer membrane vaccines or bacterins is limited. It is known that antibodies against *M. haemolytica* serotype 1 LKT will cross neutralize the toxin prepared from other serotypes. Therefore, commercial vaccines that stimulate anti-LKT antibodies should provide some cross protection against other serotypes.

However, Conlon et al. (supra) demonstrated that vaccination with recombinant LKT alone failed to stimulate protection against experimental *M. haemolytica* challenge, and Purdy et al (Purdy C W, Straus D C, Struck D, Foster G S. Efficacy of *Pasteurella haemolytica* subunit antigens in a goat model of pasteurellosis. Am J. Vet res 1993; 54:1637-47) found that vaccination of goats with LKT-impregnated agar beads stimulated incomplete immunity. Shewen and Wilkie (Shewen P E, Wilkie B N. Vaccination of calves with leukotoxic culture supernatant from *Pasteurella haemolytica*. Can J Vet Res 1988; 52:30-6) demonstrated that immunity to *M. haemolytica* was directed against both surface antigens and

LKT. The actual surface antigen of importance in stimulating protections is not known for sure; however, studies indicate that it is most likely outer membrane proteins and not capsular polysaccharide or lipopolysaccharide. Pandher et al. (Supra) demonstrated the presence of a PlpE-like protein in outer membrane of all *M. haemolytica* serotypes except serotype 11, an uncommon isolate from shipping fever. There was some variation in molecular masses among the various proteins. With the current findings, demonstrating immunogenicity of rPlpE and augmentation of a commercial vaccine that stimulates anti-LKT antibodies, the previous demonstration of a PlpE-like protein in most serotypes and the anti-LKT antibodies can neutralize LKT from other serotypes, the addition of rPlpE to a commercial vaccine that stimulates anti-LKT antibodies will enhance cross serotype protection in shipping fever.

EXAMPLE 2

Characterization of rPlpE epitopes

Additional studies were undertaken to characterize surface-exposed and immunologically important epitopes of rPlpE.

1. Materials and Methods

1.1 Construction and Purification of Truncated Forms of rPlpE

Three additional rPlpE proteins carrying varying degrees of deletions were constructed in pET28 and purified according to the method described above. The first of these was obtained by using plpBM-1 (5'-CTTGGATCCCCAAGCA-CAAAATGTT-3') (SEQ ID NO: 3), a primer that primes 84 bp into the 5' end of plpE thus introducing a deletion of 28 amino acids into the N-terminus end of rPlpE (rPlpEAN28); and the second by plpBM-2 (5'-CCTGGATCCCCAAGCA-GAGGTTACT-3') (SEQ ID NO: 4), which primes 228 bp into the 5' end of plpE introducing a 76 amino acid deletion in the N-terminus of rPlpE (rPlpEAN76); and the third with plpBM-3 (5'-ATTGGATCCAATGCTGAACAACCTC-3') (SEQ ID NO: 5) that primes 450 bp into 5' end of plpE introducing a deletion of 150 amino acids into the N-terminus in of rPlpE (rPlpEAN150). The reverse primer in all instances was plpEER, (5'-GACTGAATTCT-TATTTTTTCTCGCTAACCATTA-3') (SEQ ID NO: 6).

1.2. Production of Polyclonal Mouse Ascites

Three female, CFW mice were immunized 3 times with 50 µg of complete or truncated rPlpE diluted by half in RIBI (Corixa Corp, Seattle, Wash.) adjuvant. The first immunization was done subcutaneously (SC). Subsequent immunizations were done intraperitoneally (IP). A test-bleed was performed and the serum screened for antibodies to rPlpE by ELISA. The response was moderate, so two additional immunizations were performed IP. The mice were then injected with approximately 2×10⁶ sarcoma cells (ATCC cat # TIB-66). Between 7 and 10 days after sarcoma injection, the mice started producing ascites. Ascites fluid was removed from each mouse three times; mice were then euthanized by barbiturate overdose.

1.3. Preparation of Affinity Columns and Purification of Anti-PlpE Antibodies

Purified rPlpE was coupled to NHS-activated SEPHAROSE™ 4 Fast Flow (Amersham Biosciences, Upsala, Sweden) according to the manufacturer's recommendation. Briefly, 3-7 mg of rPlpE in PBS was mixed with 2 ml bed volume of washed and equilibrated NHS-activated

SEPHAROSE™ 4 Fast Flow in an Econo Column (BioRad, Hercules, Calif.), incubated at 4° C. overnight at which time the non-reacted groups were blocked by 0.1 M Tris pH 8.0, and washed with alternating high and low pH buffers, Tris, pH 8.0 and acetate buffer pH 4.0, respectively.

Several affinity columns were prepared with rPlpE carrying varying degrees of truncation from the N-terminus.

Anti-rPlpE antibodies against specific regions of PlpE were purified using the affinity columns described above. The Econo-Column with NHS-activated SEPHAROSE™ coupled to an rPlpE of interest was fitted with a Flow adaptor according to the recommendation of the manufacturer (BioRad, Hercules, Calif.). The affinity column was equilibrated by applying Dulbecco's Phosphate Buffered Saline (DPBS) at a flow rate of 1 ml/min. Hyperimmune serum produced by immunizing calves with the intact rPlpE was mixed with DPBS in a ratio of 1 to 10 and passed through Nalgene 0.45 µm PES filters (Nalge, Rochester, N.Y.). The filtered serum was then applied to the equilibrated column via peristaltic pump at a flow rate of 1 ml/min. The flow thru was re-applied to the column several times to re-extract the serum by connecting the flow through to the reservoir of the initial serum. The column was then washed with DPBS. The complete removal of nonspecific proteins was determined with the help of the UV monitor attached to a chart recorder. Once there was no indication of nonspecific protein in the flow through, the specifically bound antibody was eluted with 100 mM Glycine Buffer (100 mM Glycine, 140 mM NaCl, pH 3.0) by collecting fractions in microfuge tubes containing 1/10 vol of 1 M Tris-HCl, pH 8.0. The absorbance of each fraction was determined at 280 nm. Those fractions that had a reading at least 2-3 times the background were pooled and dialyzed overnight against DPBS at 4° C. in a Slide-A-Lyzer® Dialysis Cassette (Pierce, Rockford, Ill.). The concentration of affinity purified antibody was determined with BCA Protein Assay Kit (Pierce, Rockford, Ill.). More specific antibodies against rPlpE with 28, 76 and 150 amino acids deletions on their N-termini, rPlpEAN28 (pSAC63), rPlpEAN76 (pSAC64) and rPlpEAN150 (pSAC65), respectively, were purified as described.

Antibodies against regions of PlpE that are exposed on the surfaces of *M. haemolytica* cells were purified as described by Turbyfill et al., (1998). Briefly, intact *M. haemolytica* cells from the late log phase were incubated with hyperimmune bovine sera immunized by rPlpE or anti-PlpE mouse ascites diluted in PBS on ice for 2-4 hr with gentle agitation. The cells were spun down and washed several times with PBS. The antibodies bound to the surface were eluted by resuspending and agitating the cells in 0.1M Glycine, 140mM NaCl, pH 3.0 for at least 30 minutes. The cells were centrifuged at 13,000×g, and the eluted antibodies were collected in the supernatant which was neutralized immediately by adding 1/10 volume of 1 M Tris, pH 8.0.

1.4. Epitope Mapping of PlpE by Peptide Array (Pepscan)

A peptide array comprising a total of 109 overlapping 13-mer peptides with 3 amino acid offsets was custom made by Sigma-Genosys LP (The Woodlands, Tex.). Briefly the synthesis of peptides was performed on cellulose membranes in which hydroxyl functions of a commercially available filter paper are derivatized with 9-fluorenylmethoxycarbonyl-B-alanine (Fmoc-B-A1a) with subsequent removal of the Fmoc group. The synthesis areas were defined by spotting a Fmoc-B-alanine-pentafluorophenyl ester solution to distinct areas on the membrane. Blocking the remaining amino functions between spots provided discrete reaction sites on the membrane for further standard solid phase peptide synthesis using

amino acid pentafluorophenyl esters. Peptides remained covalently attached to the cellulose membrane by the C-terminus and have a free N-terminus.

The peptide array was probed with anti-PlpE hyperimmune sera as follows. Prior to blotting, membranes with the custom spots were blocked with SuperBlock® Dry Blend (Pierce, Rockford, Ill.) blocking buffer in TBS, pH 7.4. The membrane was then incubated in blocking buffer containing a primary antibody at a dilution of 1:1000 to 1:5000 for an hour. Following several washes with TBS, pH 7.4, supplemented with 0.05% Tween-20, 0.2% Triton®-X-100 (TBSTT), the membrane was incubated in Superblock containing a goat anti-bovine or anti-mouse secondary antibody conjugated to Horse Radish Peroxidase (KPL, Gaithersburg, Md.) at dilutions 1:100,000 to 1:200,000 for one hour. The membrane was washed several times with TBSTT. The peptide array was incubated with SuperSignal® West Pico Chemiluminescent Substrate working solution (0.125 ml/cm²) for 5 minutes, placed in plastic membrane protector and exposed to a CL-X POSURE™ (Pierce, Rockford, Ill.) X-Ray film for varying durations of time. The X-Ray film was then developed in a Konica SRX-101A Medical Film Processor (Taiwan). The developed X-Ray film was scanned by Arcus 1200 Agfa scanner (Taiwan), and scanned images were analyzed using Gene Pix® Pro 4.0 (Axon Instruments, Union City, Calif.). Signal intensities were defined as median pixel intensity following subtraction of local median background signal. The peptide array was stripped with RESTORE™ Western Blot Stripping Buffer (Pierce, Rockford, Ill.) according to the procedure recommended by the manufacturer before it was probed with a different anti-PlpE antibody. This was repeated several times with anti-PlpE antibodies obtained from different sources or purified in varieties of ways.

2. Results

2.1. Epitope Mapping of PlpE

The determination of the epitope map of PlpE was attempted in two steps. The first approach involved the localization of the general area of immunogenic or immunodominant epitopes by deleting specific regions of PlpE from both the N-Terminus and C-Terminus by PCR with the help of specific primers. A total of 6 plasmid constructs carrying the plpE gene with varying degrees of deletions were made. The cloning of three of these constructs that carry deletions from the N-termini of PlpE, (pSAC63, pSAC64, and pSAC65) is described above. Three additional plasmids viz., pSAC30, pSAC31, and pSAC32 that carry 106(rPlpEAC106), 96(rPlpEAC96) and 86(rPlpEAC86) amino acid deletions on the C-terminus of PlpE, respectively, were designed and constructed. The reverse primers used to introduce these deletions on the 3' end of the plpE gene in the latter constructs were HNplp-1 (5'-GATAAGCTTTTACCGTGC-GCAAATTC-3') (SEQ ID NO: 7), HNplp-2(5'-AAAAAGC-TTTATTTAATTCTACATC-3') (SEQ ID NO: 8), and HNplp-3 (5'-TTTAAGCTTTTATATACTTCCTTGAGC-3') (SEQ ID NO: 9), respectively, and a forward primer plpEBH, (5'-GTCAGGATCCTGCGGAGGAAGCGGTAGC-3') (SEQ ID NO: 10).

Amplimers were cut with BamHI and HindIII and cloned into pET28 or pRSETA cut with the same enzymes. Following confirmation of the identity of putative clones by both restriction analysis and sequencing, plasmids from true clones were introduced into BL21 (DE3) by transformation where the truncated forms of PlpE were overexpressed and purified according to the protocol described earlier. The 6 truncated forms of rPlpE and the intact form were separated on a 12.5% SDS-PAGE for Western analysis. Hyperimmune

serum from calves immunized with the intact rPlpE was used as primary antibodies and goat anti-bovine alkaline Phosphatase conjugated antibodies as secondary antibodies. Densitometric analysis of the respective bands in a Western blot in which the same amount of the recombinant proteins were loaded onto an SDS-PAGE and probed with hyperimmune serum from a calf that was immunized with rPlpE clearly showed that there are significant differences amongst recombinant proteins carrying deletions in the intensity of their reaction to the hyperimmune serum. Accordingly there is no difference in the intensity of binding between rPlpE and mutants with the deletions from the C-terminus viz., pSAC30, pSAC31, and pSAC32 that carry 106 (rPlpEAC106), 96 (rPlpEAC96) and 86 (rPlpEAC86) amino acid deletions on the C-terminus of PlpE, respectively. The binding capacity of mutants carrying deletions on their N-termini decreases with increasing deletions. There is no appreciable difference between rPlpE and pSAC63 (rPlpEΔN28) with 28 amino acid deletions on the N-terminus. The reactivity of pSAC64 (rPlpEΔN76), which carries a deletion of 76 amino acids on the N-terminus, drops to 63%, which is a 37% loss in signal intensity, when compared to rPlpE. Further deletion into the N-terminus as seen in pSAC65 (rPlpEΔN150) reduces the binding capacity of the truncated proteins by 60%. These findings clearly suggest that the region between residues 28 and 76 from the N-terminus of PlpE carries a stretch of amino acids with possible epitope(s) that may be responsible for invoking the immune response elicited when rPlpE is used as a vaccine.

2.2. Fine Mapping of Epitopes on PlpE

Putative antigenic regions in PlpE were identified by using the MACVECTOR™ 7.0 software that employed algorithms such as antigenic index, hydrophilicity and surface probability. However, the identification of epitopes was done with a peptide array comprising 109 overlapping 13-mer peptides that were synthesized by the chemistry described earlier. The peptides were covalently bound to derivatized cellulose membrane by the C-terminus and have a free N-terminus. Anti-PlpE hyperimmune antibodies purified by any number of the methods described earlier were used to probe the peptide array. The custom spots were stripped and probed several times. When bovine antibody against surface exposed components of PlpE that was affinity purified with intact *M. haemolytica* cells was used to probe the peptide array a total of 8 distinct regions (E1-8) were identified (FIG. 6). Epitope 1 (PNHPKPVLVPKTQNNL) (SEQ ID NO: 11) spans 3 peptides; epitope 2 (QNASQAQNAPQAQNAPQAQNAPQVENAPQAQNAPQVENAPQAE) (SEQ ID NO: 12), 11 peptides; epitope 3 (GSFDKIGSVKLNK) (SEQ ID NO: 13), 3 peptides; epitope 4 (KLGTPPKFDKVS GKKIIEE) (SEQ ID NO: 14), 6 peptides; epitope 5 (LIRRSDDLFGYGY) (SEQ ID NO: 15), 3 peptides; epitope 6 (ADKFSQYFVVYDE) (SEQ ID NO: 16), 3 peptides; epitope 7 (NISDKLTATYRKK) (SEQ ID NO: 17), 2 peptides; and epitope 8 (PHTKEFAARISKL) (SEQ ID NO: 18). Approximately the same set of epitopes, albeit with decreasing intensities, were picked up when whole serum obtained from cows with a naturally high anti-PlpE antibody titer that were also challenged with live *M. haemolytica* was used. The signal intensities of all of the epitopes with the exception of epitope 2 were much lesser in this blot than in the earlier. The purification of IgG from the latter serum with Protein G affinity columns did not alter the above result in that exactly the same putative epitopes were identified suggesting that IgG was the class of immunoglobulins involved in this immune response. When whole hyperimmune serum from calves immunized

with rPlpE was used to probe the stripped peptide array, the same set of peptides mentioned above were identified once again confirming the binding capacity of the above indicated stretches of amino acids along PlpE. On the other hand, when sera from calves that were given live *M. haemolytica* were used to probe the peptide array, epitope 2 was the only one that was picked up. According to the manufacturers of the custom spots, non-specific binding of the antibody-enzyme conjugate may occur to peptides that contain combinations of basic amino acids. When goat anti-bovine-HRP, the secondary antibody used in this project, was used to probe the spots, epitopes 1, 3, 4, 7, and 8 were picked up. The same sets of epitopes were identified when the array was probed with rabbit anti-bovine-HRP, showing putative epitopes 1, 3, 4, 7, and 8 were not true epitopes. In order to identify spots that would non-specifically bind bovine immunoglobulins, serum from colostrum deprived new born calf was used to probe the array. Interestingly, in addition to the putative epitopes identified by the secondary antibody-enzyme conjugates, i.e. 1, 3, 4, 7, and 8, epitopes 5 and 6 exhibited reactivity to bovine immunoglobulins. Epitope 2 was the only one that did not react to both the serum from the colostrum deprived calf and secondary antibody-enzyme conjugate showing that this epitope is the only one responsible for inducing the specific immune response when calves were either vaccinated with rPlpE or *M. haemolytica*.

A closer examination of epitope 2 shows that this is part of the region identified as having 8 imperfect repeats of hexapeptides (Pandher et. al., 1998). The 11 peptides (#13 through 23) identified here as epitope 2 comprise the last 4 residues of the 2nd repeat described by Pandher et al., (1998) and the rest of the repeats i.e., repeats 3 through 8 with the exception of the 1st hexapeptide. A feature of these 11 peptides is the lack of uniformity in their binding capacity as evidenced by the variation in their signal intensities. Peptides #15, 17, and 19 exhibit the highest signal intensities followed by #s 21 and 23. The first five residues of the N-termini of these peptides are QNAPQ (SEQ ID NO: 33) with the exception of #21 in which the first glutamine is replaced by glutamate. It is worthwhile noting that both glutamine and asparagine are positively charged, with hydrophobicity index of -0.91 and -0.92, respectively. The remaining 6 peptides in epitope 2 have proline at their N-termini instead of glutamine and this may account for their relatively lower signal intensity in the peptide array. The relatively high signal intensities exhibited by peptides 15, 17, and 19 may reflect the manner in which these epitopes are presented to the immune system under natural condition on the surfaces of *M. haemolytica* cells and the inherent immunogenic nature of these stretches of amino acids. The fact that epitope 2 contains a significant number of prolines at defined intervals which are usually indicators of turns, has an unusually high number of very basic residues such as glutamine, asparagine and glutamate which are hydrophilic, with high surface probability and 8 repeats are features that are usually associated with regions of protein that are associated with being immunogenic. Moreover, computer analysis of the deduced amino acid sequence of epitope 2 with algorithms such as Parker's antigenicity, Kyte/Doolittle hydrophilicity, surface probability and Chou Fasman D structure indices show that the stretch of amino acids has a moderately high antigenicity, fairly hydrophilic, contains fairly high number of amino acids with very high surface probability and is characterized by series of turns

the final 3 mice per subgroup were bled on day 42. Antibody responses to PlpE and LkTA were determined by ELISA

TABLE 2

Experimental design of murine chimeric protein immunization study			
Group #	Plasmid Designation	Description of Insert	Name of Recombinant Protein
1	pSAC86	G-R2-LKTA	SAC86
2	pSAC87	G-2(R2-LKTA)	SAC87
3	pSAC88	R2-LKTA	SAC88
4	pSAC89	2(R2-LKTA)	SAC89
5	pSAC91	G-S-R2-S-LKTA-S	SAC91
6	Control	Titermax ® adjuvant only	n/a

G = GST leader peptide

R2 = immunodominant epitope of PlpE

LkTA = section of the C terminus of LkTA that includes the leukotoxin neutralizing epitope

S = spacer peptide

The endpoint titration data when rPlpE is used as a ligand is shown in FIG. 15A-C. (The nomenclature in the figures designates the protein and its concentration used in the vaccine, e.g. "8825" or "88-25" indicates mice vaccinated with 25 µg of SAC88.) A comparison of the immune responses to rPlpE indicates that SAC 86, 87, 88 and 89 are all highly immunogenic. However, this is not the case with SAC91, the chimera that includes spacer peptides. As can be seen, the chimeras that contain 2 copies of the epitopes from the two proteins (i.e. SAC87 and SAC89) were better immunogens, as evidenced by the high titer of the anti-rPlpE antibodies in the sera of mice that were vaccinated with these two recombinant proteins.

Antibody titers against mlkTA is shown in FIG. 15D-F. As can be seen, SAC 86, SAC87, SAC88 and SAC89 elicit significantly higher antibody titers to this immunodominant epitope than does SAC91.

The results of a summary of antibody responses to the chimeric proteins as determined by a single dilution ELISA are presented in FIG. 16. As can be seen, according to this assay, SAC89 protein (containing two R2 and two mLKTA antigens) was the best overall antigen.

The response of mice to vaccination with the chimeric proteins was also quantified with Western blots, coupled with densitometric analysis of the bands. The results are given in FIGS. 17A and B, with FIG. 17A showing the results obtained for LKT and FIG. 17B showing the results obtained for PlpE. As can be seen, the SAC89 protein again appears to elicit the best overall response to both antigens.

FIG. 18 depicts LKT neutralization activity of murine anti-chimeric immune sera at a dilution of 1:16 as determined by MTT assay. A colorimetric microtitration assay was adapted to quantify the cytotoxicity of LKT to bovine neutrophils used as target cells (Vega, M. V., S. K. Maheswaran, J. R. Leininger, and T. R. Ames. 1987. Adaptation of a colorimetric microtitration assay for quantifying *Pasteurella haemolytica* A1 leukotoxin and antileukotoxin. Am J Vet Res 48:1559-1564). The viability of LKT-treated target cells was detected by use of methylthiazole tetrazolium (MTT) assay. The MTT assay is based on measuring the activity of living cells via mitochondrial dehydrogenases that cleave the tetrazolium ring of MTT, yielding purple formazan crystals which are insoluble in aqueous solutions. The crystals are dissolved in acidified isopropanol and measured spectrophotometrically. The amount of formazan formed was quantified by use of an ELISA plate reader and is directly proportional to the number of viable target cells, thus allowing its adaptation for detecting LKT-neutralization antibody titers. The mouse sera were

combined with lyophilized LKT to allow neutralization of the LKT by anti-LKT antibodies. BL-3 cells were suspended to a concentration of 2.5×10^6 cells/ml and 50 µl was added to each neutralization reaction. The cell mixture was incubated and the soluble MTT was added to each sample. MTT is converted by viable cells to an insoluble precipitate, which is then dissolved and read spectrophotometrically. The amount of precipitate formed relates to the amount of LKT neutralization that has occurred. LKT neutralizing mouse monoclonal antibody (MM601) was used as a positive control and colostrum-deprived calf serum (CDCS) was used as a negative control. As can be seen, LKT neutralization activity of anti-chimeric immune sera ranges from 45% to 142% of that of MM601.

FIG. 19 depicts bactericidal activity of murine anti-SAC-89 hyper-immune sera obtained as the result of administering either 25 µg of SAC89 (89-25) or 50 µg of SAC89 (89-50). Tests were carried out in the presence of a suitable source of complement. Tests were carried out in the presence of a suitable source of complement. Complement-mediated killing assay was done as we have previously described (Ayalew, S., A. W. Confer, and E. R. Blackwood. 2004. Characterization of immunodominant and potentially protective epitopes of *Mannheimia haemolytica* serotype 1 outer membrane lipoprotein PlpE. Infect Immun 72:7265-7274.). Briefly, *M. haemolytica* cells were grown in BHI broth and decapsulated in 1xPBS at 41° C. The cells were resuspended to an O.D.₆₀₀=0.500 and then diluted to 1:1000 in PBS for use in the assay. The mouse sera were heat treated at 56° C. to inactivate existing complement and used as the antibody sources. Colostrum-deprived calf serum was used as the source of complement in the assay. The antibodies, bovine complement, and decapsulated *M. haemolytica* cells were mixed and plated on BHIA Blood plates at T₀ and T₃₀ after incubation at 37° C. Growth was determined by counting the number of colonies present after 15-16 hours of incubation at 37° C. and 5% CO₂ and the percent killing was calculated with the formula: [(T₀ growth-T₃₀ growth)/T₀ growth]×100%

The results showed that the serum bactericidal activity of mouse anti-SAC89 hyper-immune sera is as potent as serum from a calf that was vaccinated with intact rPlpE.

EXAMPLE 4

Vaccination With SAC89 and Challenge With *Mannheimia haemolytica* Experimental design and results. Fifteen recently weaned Angus cross steers were purchased. Eight were vaccinated subcutaneously with PBS in Emulsigen-P adjuvant on day 0, whereas 7 received 100 µg of SAC89 plus adjuvant subcutaneously in the neck. The vaccine was repeated on day 28. Antibody responses against SAC89, leukotoxin, recombinant PlpE, and whole bacterial cells were significantly increased by day 14 after vaccination (FIG. 20 A-D). A decline in antibodies followed with a significantly higher response after revaccination on day 28. During the course of the study, calf #48 caught its head in a feed bunk and broke its neck. It was thus removed from the study.

On day 42, cattle were challenged transthoracically with 5×10^9 CFU of live, virulent *Mannheimia haemolytica* A1 Oklahoma strain (Panciera & Corstvet, Am J Vet Res. 1984 December; 45 (12):2532-7). Calf #25 died within 3 days after challenge due to severe pneumonia and terminal septicemia and was given the maximum score of 20 (Panciera et al. Am J Vet Res 1984 December; 45 (12):2538-42). Four days after challenge, cattle were humanely killed and lungs evaluated for lesion scores on a 20 point scale (0 being no lesion and 20

being maximum). There was a significant reduction (69.9%, $p < 0.02$) in lesion scores for SAC89 vaccinates compared to PBS/Adjuvant vaccinates (Table 3).

TABLE 3

Lesion scores after vaccination with SAC89 or PBS		
Calf No.	Vaccine	Lesion Score
8	PBS/ADJ	1.5
21	PBS/ADJ	7.5
22	PBS/ADJ	7.5
25	PBS/ADJ	20
29	PBS/ADJ	13
31	PBS/ADJ	8.5
41	PBS/ADJ	5
42	PBS/ADJ	1.5
48	PBS/ADJ	Deceased*
Mean \pm SD		8.06 \pm 6.13
3	SAC89/ADJ	1.5
7	SAC89/ADJ	1.5
12	SAC89/ADJ	0
23	SAC89/ADJ	2
30	SAC89/ADJ	2
32	SAC89/ADJ	5.5
39	SAC89/ADJ	4.5
Mean \pm SD		2.43 \pm 1.9
(69.9% reduction in lesion scores)		

*Calf died from an accident and was removed from the study.
 $P < 0.02$ between vaccinates and controls

This example shows that vaccination of cattle with a recombinant chimeric protein comprising PlpE-LKT of *M. haemolytica* provides significant protection against challenge with virulent *M. Haemolytica*. In addition, the vaccine stimulates antibodies to *M. haemolytica* LKT, whole cells, outer membrane lipoprotein PlpE and to the chimeric protein itself.

Accordingly, it can be appreciated that subunits derived from PlpE, and especially epitope 2 (i.e. R2), are useful as well in the inventive vaccine compositions and methodologies. The inclusion of such region(s) enhances the host immune response directed against relevant immunoprotective epitopes. It accordingly can be appreciated that the inventive vaccines utilize as distinct antigenic components rPlpE or subunits thereof capable of eliciting an antibody or other immune response against *M. haemolytica*. As a result, the invention encompasses proteins which may be the full length antigen, antigen fragment, antigen derivative or a fusion product of such antigen, antigen fragment or antigen derivative with another protein. In particular, chimeric proteins comprising antigenic regions of rPlpE and LKT are effective in eliciting a protective immune response against *M. haemolytica*. Proteins included within the present invention include those depicted in the Sequence Listing as well as mutants of said sequences capable of eliciting an antibody or other immune response which recognizes an epitope(s) of such amino acid sequences. In general, the polypeptides that represent the proteins of the invention will have at least from about 60 to 70% identity with the sequences presented herein, and preferably about 70 to about 80% identity, and more preferably about 80 to 90% identity, and most preferably about 90-95 or 95-100% identity.

In addition, the chimeric proteins of the invention may also include a leader sequence. In one embodiment, the leader sequence is glutathione-S-transferase (GST), the sequence of which is known and readily available (see, for example, U.S. Pat. No. 6,368,584, the contents of which is hereby incorporated by reference). However, those of skill in the art will recognize that other leader sequences exist which are also suitable for use in the invention.

In addition, other short sequences may serve as "spacer+ peptides, for example, between R2 peptide sequences in a polypeptide that comprises more than one R2 sequence, and/or between R2 and mLKT sequences in a polypeptide that comprises one or more of R2 or mLKT, or both. In one embodiment, the spacer peptide is Arg-Ser (i.e. "RS"). However, those of skill in the art will recognize that other spacer peptides may also be used in the practice of the invention, including but not limited to Gly-Gly-Gly-Ser (i.e. GGGs, SEQ ID NO: 34), and others, so long as the spacer peptides allow exposure of the antigenic epitopes to the immune system a manner that results in an immune response.

The nucleotide sequences used to generate the antigens may be inserted into any of a wide variety of expression vectors by a variety of procedures. Such procedures and others are deemed to be known by those skilled in the art. Suitable vectors include chromosomal, nonchromosomal and synthetic DNA sequences; e. g., derivatives of SV40; bacterial plasmids; phage DNAs; yeast plasmids; vectors derived from combinations of plasmids and phage DNAs, viral DNA such as baculovirus, vaccinia, adenovirus, fowl pox virus, pseudorabies, etc. The appropriate DNA sequence must be operatively linked in the vector to an appropriate expression control sequence(s) (promoter) to direct mRNA synthesis. As representative examples of such promoters, there may be mentioned LTR or SV40 promoter, the *E. coli* lac or trp, the phage lambda PL promoter and other promoters known to control expression of genes in prokaryotic and eukaryotic cells or their viruses. The expression vector also includes a non-coding sequence for a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression.

The vector containing an appropriate sequence, as well as an appropriate promoter or control sequence, may be employed to transform an appropriate host to permit the host to express the protein. Examples of host organisms and cells include bacterial strains (e. g., *E. coli*, *Pseudomonas*, *Bacillus*, *Salmonella*, etc.), fungi (e. g., yeasts and other fungi), animal or plant hosts (e.g., mouse, swine or animal and human tissue cells). The selection of the host is deemed to be within the scope of those skilled in the art.

As previously mentioned, it is also understood that the appropriate sequence present in the vector when introduced into a host may express part or only a portion of the protein which is encoded within the noted terminology, it being sufficient that the expressed protein be capable of eliciting an antibody or other immune response which recognizes an epitope(s) of the listed amino acid sequences.

The isolated polypeptides expressed by the host transformed by the vector may be harvested by methods which will occur to those skilled in the art and used in a vaccine for providing an enhanced immune response against infection with *M. haemolytica*. Vaccine preparation is easily accomplished using well known methods and techniques. An enhanced immune response is manifest by protection against infection or a decrease in severity of infection, which may be reflected in body temperature and antibody titers as described above.

The host expressing the antigen may itself be used to deliver antigen to non-human animals, by introducing killed or viable host cells that are capable of propagating in the animal. Direct incorporation of the nucleotide sequences into host cells may also be used to introduce the sequences into animal cells for expression of antigen in vivo.

Vaccine preparations are combined with physiologically acceptable carriers to form vaccines. The carrier employed in

conjunction with vaccine may be any one of a wide variety of carriers. As representative examples of suitable carriers, there may be mentioned mineral oil, synthetic polymers, etc. Carriers for vaccines are well known in the art and the selection of a suitable carrier is deemed to be within the scope of those skilled in the art. The selection of a suitable carrier is also dependent upon the manner in which the vaccine is to be administered. The preferred physiologically acceptable carrier is an adjuvant. Preferably, the inventive vaccine formulation is set to contain about 10-100, and preferably about 100, micrograms of recombinant antigens in commercially available adjuvant (Pfizer). Similar quantities of recombinant antigens would be used if added to another commercial vaccine formulation.

Examples of adjuvants that may be used in the practice of the invention include but are not limited to: aluminum hydroxide gel-based adjuvants, saponin-based adjuvants, block co-polymer-based adjuvants, water-in-oil adjuvants, and oil-in-water adjuvants. Specific examples include but are not limited to Freund's incomplete adjuvant, TiterMax®, Emulsigen®-P, Xtend II, Xtend SP, SUPERIMM®, and RIBI adjuvant.

The vaccines may be administered by a variety of routes including intravenously, intraperitoneally, intramuscularly, and subcutaneously. The preferred route of administration is subcutaneous. Alternatively, the vaccine may be administered intranasally or orally. The vaccine can be administered in a single dose or multiple doses until a protective effect is achieved.

Those of skill in the art will further recognize that in order to function as a "vaccine", inoculation with a protein of the invention (e.g. a chimeric protein) may, on the one hand, offer full protection from the development of symptoms associated with infection by *M. haemolytica* (i.e. so-called symptoms of "shipping fever"). However, a vaccine preparation can be valuable even if symptoms are not totally prevented, but are merely attenuated. Further, the vaccine preparation may be used prophylactically prior to suspected exposure to the etiological agent of the disease, or after exposure, or even after some symptoms of disease have appeared. Benefit from the immune stimulating effects of the vaccine preparation that is administered may accrue even after the onset of infection.

The vaccines of the invention may be administered alone as the sole agent for combating *M. haemolytica* infection. Alternatively, the vaccine preparations may be administered in concert with other agents such as vaccine preparations utilizing other *M. haemolytica* proteins or antigens, several commercial varieties of which are known.

According to the invention, chimeric proteins which include one or more immunodominant epitopes of recombinant PlpE in combination with one or more immunodominant epitopes of LKT, are provided as vaccinating agents. Those of skill in the art will recognize that several terms are used in the art to describe peptide and/or polypeptide sequences that elicit an immune response, and that there is sometimes overlap or inconsistency within the art with respect to the categorization of such sequences. Herein, the term "immunodominant epitope" or "immunodominant region" is intended to refer to regions (i.e. segments or portions) of a protein from a pathogenic organism that, when administered to a mammal, are capable of inducing a protective immune response to the organism in the mammal. Generally, but not always, such immunodominant regions or epitopes are highly immunogenic when tested according to methods that are known to those of skill in the art, such as those described herein.

In a preferred embodiment of the invention, the exemplary immunodominant region or epitope of PlpE that is utilized is

R2 as represented by SEQ ID NO: 19, and the exemplary immunodominant region or epitope of LKT is mLKT as represented by SEQ ID NO: 21. In addition, those of skill in the art will recognize that several modifications of these sequences can be made for any of several purposes, without compromising the ability of the chimeric protein to elicit a suitable immune response. For example, conservative amino acid substitutions can be tolerated. Conservative amino acid substitution can be defined as recognized by those of skill in the art, for example, according to the BLOSUM62 matrix, described by Henikoff and Henikoff (S. Henikoff and J. G. Henikoff, *Proc. Natl. Acad. Sci. USA* 89, 10915-10919, 1992). In addition, the deletion of a small number (e.g. approximately 1-10) amino acids from the amino or carboxy terminus or within the sequence, or the substitution of various modified or non-natural amino acids, addition of histidine tags, etc. may also be tolerated without compromising the ability of the immunodominant region or epitope to function in the practice of the present invention. Such modifications may be carried out for any of a variety of purposes, including but not limited to increase or decrease solubility of the polypeptide, to prevent or aid digestion by proteases, to facilitate isolation and purification of the protein (polypeptide), etc. Further, various labels may be included to facilitate tracking of the protein (e.g. introduction of a tryptophan residue, or introduction of a chemical label). In general, polypeptides that are encompassed by the invention will have at least from about 60 to 70% identity with the sequences presented herein, and preferably about 70 to about 80% identity, and more preferably about 80 to 90% identity, and most preferably about 90-95 or even about 95-100% identity, e.g. to R2, or to mLKTA individually. All such sequence variants are intended to be encompassed by the invention, so long as the resultant peptide sequence retains the activity of the parent peptide sequence. Those of skill in the art are well acquainted with methods by which one can test and compare the immunogenicity of different peptides/polypeptides and their effectiveness in eliciting an immune response, and their ability to provide protection to challenge with a disease-causing entity. Exemplary methods are fully described in the Examples section of this application.

In some embodiments, the chimeric protein contains multiple copies of the immunodominant epitopes. In one embodiment, the chimera contains two copies each of R2 and mLKTA. However, this need not be the case. Higher numbers of copies are also contemplated (e.g. 3 or even many more). Any number of copies may be utilized, so long as the construct that encodes the chimera and the chimera itself are able to be successfully manipulated and processed in the laboratory and during vaccine preparation. Further, the number of copies of each moiety (R2 and mLKTA) need not be equal, i.e. a chimera may contain one copy of R2 and two or more copies of mLKTA, or vice versa. In addition, one or more immunodominant epitopes from other species or strains can also be incorporated into the chimera, e.g. from other *M. haemolytica*.

In addition, the arrangement (i.e. order or position) of the immunodominant regions within the chimeric protein may vary. For example, the protein may contain two R2 regions in tandem, followed by two mLKTA regions in tandem. Alternatively, a protein may contain alternate regions, e.g. R2-mLKTA-R2-mLKTA, etc. Any such combination is considered to be within the scope of the invention, so long as the resulting chimeric protein elicits a suitable and useful immune response to *M. haemolytica*, e.g. a sufficient response to prevent or attenuate symptoms of disease that

would likely occur in the vaccinated mammal, if the mammal had not been vaccinated by a preparation containing the chimeric protein.

Likewise, while exemplary DNA acid sequences that encoding chimeric proteins are presented herein, those of skill in the art will recognize that the nucleic acids of the invention are not limited to those specific sequences, or even to DNA. RNA encoding the chimeric proteins is also within the scope of the invention, as are modifications to the encoding sequences that can be tolerated without vitiating the efficacy of the polypeptide that is produced. For example, due to the redundancy of the genetic code, many sequences other than those that are presented may be used in the practice of the invention. Other variants of the nucleic acid sequences may be introduced, for example, in order to stabilize the nucleic acid, to facilitate isolation or tracking of the DNA (e.g. various labels), etc. In general, nucleic acid sequences that are included in the present invention will be about 60 to 70% homologous to the sequences presented herein, and preferably about 70 to about 80% homologous, and more preferably about 80 to 90% homologous, and most preferably about 90-95 or 95-100% homologous, e.g. to R2, or to mLKTA encoding sequences.

In addition to the administration of a proteinaceous vaccine preparation, the invention also contemplates administration of a nucleic acid encoding the antigenic protein. In this embodiment, the protein (e.g. a chimeric protein) is translated within the vaccinated host. Those of skill in the art are aware of the many systems for delivery of nucleic acid (e.g. DNA) vaccines, including liposomal preparations, various viral vectors, (e.g. adenovirus, hepatitis, etc.), and the like.

In view of the above, it will be seen that the several objectives of the invention are achieved and other advantageous results attained. As various changes could be made in the above sequences, antigens, etc. without departing from the scope of the invention, it is intended that all matter contained in the above description or shown in the accompanying drawings shall be interpreted as illustrative and not in a limiting sense.

While the invention has been described with a certain degree of particularity, it is understood that the invention is not limited to the embodiment(s) set forth herein for purposes of exemplification, but is to be limited only by the scope of the attached claim or claims, including the full range of equivalency to which each element thereof is entitled.

SEQUENCE LISTING

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Val Pro Gln Ala Gln Asn Ala Ser Gln Ala Gln Asn Ala Pro Gln Ala
35          40          45
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Gln Ala Gln Asn Ala Pro Gln Val Glu Asn Ala Pro Gln Ala Glu Val
65          70          75          80
Thr Pro Pro Val Pro Gln Pro Gln Ser Gln Lys Ile Asp Gly Ser Phe
85          90          95
Asp Lys Ile Gly Ser Val Lys Leu Asn Lys Glu Ala Gln Thr Leu Glu
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Gly Lys Asn Leu Val Asp Ala Ala Asp Lys Phe Ser Gln Tyr Phe Val
180         185         190
Val Tyr Asp Glu Lys Arg Val Asn Asp Asn Ile Ser Asp Lys Leu Thr
195         200         205
Ala Thr Tyr Arg Lys Lys Glu Gly Phe Val Tyr Gly Ser Asn Pro His
210         215         220
Thr Lys Glu Phe Ala Ala Arg Ile Ser Lys Leu Gly Asp Val Glu Ile
225         230         235         240
Lys Phe Glu Asn Gly Gln Ala Gln Gly Ser Ile Lys Asp Glu Lys Asp
245         250         255
Gly Asn Ala Glu Ile Phe Thr Ile Lys Gly Asp Thr Lys Gln Leu Glu
260         265         270
Ile Thr Pro Thr Glu Ser Asn Arg Ile Ile Ile Ala Ile Leu Asp Gln
275         280         285
Asn Gln Lys Ser Tyr Thr Pro Gly Met Glu Lys Ala Ile Met Glu Thr
290         295         300
Lys Phe Ile Asp Ser Lys Ala Gly Asn Ser Asp Gln Lys Tyr Leu Ile
305         310         315         320
Gly Glu Ala Lys Ser Asp Asn Trp Gln Ala Ile Met Val Ser Glu Lys
325         330         335
Lys

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Lys

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<210> SEQ ID NO 3
<211> LENGTH: 24
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide primer

<400> SEQUENCE: 3

cttgatccc aagcacaaaa tggt 24

<210> SEQ ID NO 4
<211> LENGTH: 24
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide primer

<400> SEQUENCE: 4

cctgatccc aagcagaggt tact 24

<210> SEQ ID NO 5
<211> LENGTH: 24
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide primer

<400> SEQUENCE: 5

attgatcca atgctgaaca actc 24

<210> SEQ ID NO 6
<211> LENGTH: 33
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide primer

<400> SEQUENCE: 6

gactgaattc ttattttttc tcgctaacca tta 33

<210> SEQ ID NO 7
<211> LENGTH: 27
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide primer

<400> SEQUENCE: 7

gataagcttt taccgtgagg caaatc 27

<210> SEQ ID NO 8
<211> LENGTH: 27
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide primer

<400> SEQUENCE: 8

aaaaagcttt tatttaattt ctacatc 27

<210> SEQ ID NO 9
<211> LENGTH: 27
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:

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<223> OTHER INFORMATION: oligonucleotide primer

<400> SEQUENCE: 9

ttaaagcttt tatatacttc cttgagc

27

<210> SEQ ID NO 10

<211> LENGTH: 28

<212> TYPE: DNA

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: oligonucleotide primer

<400> SEQUENCE: 10

gtcaggatcc tgcggaggaa gcggtagc

28

<210> SEQ ID NO 11

<211> LENGTH: 16

<212> TYPE: PRT

<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 11

Pro	Asn	His	Pro	Lys	Pro	Val	Leu	Val	Pro	Lys	Thr	Gln	Asn	Asn	Leu
1				5					10					15	

<210> SEQ ID NO 12

<211> LENGTH: 43

<212> TYPE: PRT

<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 12

Gln	Asn	Ala	Ser	Gln	Ala	Gln	Asn	Ala	Pro	Gln	Ala	Gln	Asn	Ala	Pro
1			5						10					15	

Gln	Ala	Gln	Asn	Ala	Pro	Gln	Val	Glu	Asn	Ala	Pro	Gln	Ala	Gln	Asn
		20					25						30		

Ala	Pro	Gln	Val	Glu	Asn	Ala	Pro	Gln	Ala	Glu
		35				40				

<210> SEQ ID NO 13

<211> LENGTH: 13

<212> TYPE: PRT

<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 13

Gly	Ser	Phe	Asp	Lys	Ile	Gly	Ser	Val	Lys	Leu	Asn	Lys
1				5					10			

<210> SEQ ID NO 14

<211> LENGTH: 19

<212> TYPE: PRT

<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 14

Lys	Leu	Gly	Thr	Pro	Pro	Lys	Phe	Asp	Lys	Val	Ser	Gly	Lys	Lys	Ile
1				5					10					15	

Ile Glu Glu

<210> SEQ ID NO 15

<211> LENGTH: 13

<212> TYPE: PRT

<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 15

Leu Ile Arg Arg Ser Asp Asp Leu Phe Tyr Gly Tyr Tyr

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1           5           10

<210> SEQ ID NO 16
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 16
Ala Asp Lys Phe Ser Gln Tyr Phe Val Val Tyr Asp Glu
1           5           10

<210> SEQ ID NO 17
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 17
Asn Ile Ser Asp Lys Leu Thr Ala Thr Tyr Arg Lys Lys
1           5           10

<210> SEQ ID NO 18
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 18
Pro His Thr Lys Glu Phe Ala Ala Arg Ile Ser Lys Leu
1           5           10

<210> SEQ ID NO 19
<211> LENGTH: 55
<212> TYPE: PRT
<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 19
Thr Pro Asn His Pro Lys Pro Val Leu Val Pro Lys Thr Gln Asn Asn
1           5           10           15
Leu Gln Ala Gln Asn Val Pro Gln Ala Gln Asn Ala Ser Gln Ala Gln
           20           25           30
Asn Ala Pro Gln Ala Gln Asn Ala Pro Gln Ala Gln Asn Ala Pro Gln
           35           40           45
Val Glu Asn Ala Pro Gln Ala
           50           55

<210> SEQ ID NO 20
<211> LENGTH: 165
<212> TYPE: DNA
<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 20
acaccgaatc accccaaacc agtactagta ccaaaaacac aaaataatct tcaagcaciaa      60
aatgttcctc aggacaaaaa tgctctcag gcacaaaatg cccctcaggc acaaaatgct      120
cctcaggcac aaaatgctcc tcaggtggaa aatgctcctc aggca                        165

<210> SEQ ID NO 21
<211> LENGTH: 131
<212> TYPE: PRT
<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 21
Ser Asp Ser Asn Leu Lys Asp Leu Thr Phe Glu Lys Val Lys His Asn
1           5           10           15

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Met	Leu	Lys	Met	Phe	Glu	Asp	Arg	Leu	Cys	His	Lys	Thr	Tyr	Leu	Asn
130						135					140				
Gly	Asp	His	Val	Thr	His	Pro	Asp	Phe	Met	Leu	Tyr	Asp	Ala	Leu	Asp
145					150					155					160
Val	Val	Leu	Tyr	Met	Asp	Pro	Met	Cys	Leu	Asp	Ala	Phe	Pro	Lys	Leu
				165					170					175	
Val	Cys	Phe	Lys	Lys	Arg	Ile	Glu	Ala	Ile	Pro	Gln	Ile	Asp	Lys	Tyr
			180					185					190		
Leu	Lys	Ser	Ser	Lys	Tyr	Ile	Ala	Trp	Pro	Leu	Gln	Gly	Trp	Gln	Ala
		195					200					205			
Thr	Phe	Gly	Gly	Gly	Asp	His	Pro	Pro	Lys	Ser	Asp	Gly	Ser	Thr	Ser
	210					215					220				
Gly	Ser	Gly	His	His	His	His	His	His	Ser	Ala	Gly	Leu	Val	Pro	Arg
225					230					235					240
Gly	Ser	Thr	Ala	Ile	Gly	Met	Lys	Glu	Thr	Ala	Ala	Ala	Lys	Phe	Glu
				245					250					255	
Arg	Gln	His	Met	Asp	Ser	Pro	Asp	Leu	Gly	Thr	Gly	Gly	Gly	Ser	Gly
			260					265					270		
Asp	Asp	Asp	Asp	Lys	Ser	Pro	Met	Gly	Tyr	Arg	Gly	Ser	Thr	Pro	Asn
		275					280					285			
His	Pro	Lys	Pro	Val	Leu	Val	Pro	Lys	Thr	Gln	Asn	Asn	Leu	Gln	Ala
		290					295				300				
Gln	Asn	Val	Pro	Gln	Ala	Gln	Asn	Ala	Ser	Gln	Ala	Gln	Asn	Ala	Pro
305					310					315					320
Gln	Ala	Gln	Asn	Ala	Pro	Gln	Ala	Gln	Asn	Ala	Pro	Gln	Val	Glu	Asn
				325					330					335	
Ala	Pro	Gln	Ala	Arg	Ser	Ser	Asp	Ser	Asn	Leu	Lys	Asp	Leu	Thr	Phe
			340					345					350		
Glu	Lys	Val	Lys	His	Asn	Leu	Val	Ile	Thr	Asn	Ser	Lys	Lys	Glu	Lys
		355					360					365			
Val	Thr	Ile	Gln	Asn	Trp	Phe	Arg	Glu	Ala	Asp	Phe	Ala	Lys	Glu	Val
	370					375					380				
Pro	Asn	Tyr	Lys	Ala	Thr	Lys	Asp	Glu	Lys	Ile	Glu	Glu	Ile	Ile	Gly
385					390					395					400
Gln	Asn	Gly	Glu	Arg	Ile	Thr	Ser	Lys	Gln	Val	Asp	Asp	Leu	Ile	Ala
				405					410					415	
Lys	Gly	Asn	Gly	Lys	Ile	Thr	Gln	Asp	Glu	Leu	Ser	Lys	Val	Val	Asp
			420					425					430		
Asn	Tyr	Glu	Leu	Leu	Lys	His	Ser	Lys	Asn	Val	Thr	Asn	Ser	Leu	Asp
		435					440					445			
Lys	Leu	Ile	Ser	Ser	Val	Ser	Ala	Phe	Thr	Ser	Ser	Asn	Asp	Ser	Arg
	450					455					460				
Asn	Val	Leu	Val	Ala	Pro	Thr	Ser	Met	Arg	Ser	Glu	Phe	Glu	Leu	Arg
465					470					475					480
Arg	Gln	Ala	Leu	Ile	Asn										
				485											

<210> SEQ ID NO 24

<211> LENGTH: 1459

<212> TYPE: DNA

<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 24

tgtcccttat actagggttat tggaaaatta agggccttgt gcaaccact cgacttcttt

60

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Met	Leu	Lys	Met	Phe	Glu	Asp	Arg	Leu	Cys	His	Lys	Thr	Tyr	Leu	Asn
130						135					140				
Gly	Asp	His	Val	Thr	His	Pro	Asp	Phe	Met	Leu	Tyr	Asp	Ala	Leu	Asp
145					150					155					160
Val	Val	Leu	Tyr	Met	Asp	Pro	Met	Cys	Leu	Asp	Ala	Phe	Pro	Lys	Leu
				165					170					175	
Val	Cys	Phe	Lys	Lys	Arg	Ile	Glu	Ala	Ile	Pro	Gln	Ile	Asp	Lys	Tyr
			180					185					190		
Leu	Lys	Ser	Ser	Lys	Tyr	Ile	Ala	Trp	Pro	Leu	Gln	Gly	Trp	Gln	Ala
		195					200					205			
Thr	Phe	Gly	Gly	Gly	Asp	His	Pro	Pro	Lys	Ser	Asp	Gly	Ser	Thr	Ser
	210					215					220				
Gly	Ser	Gly	His	His	His	His	His	His	Ser	Ala	Gly	Leu	Val	Pro	Arg
225				230						235					240
Gly	Ser	Thr	Ala	Ile	Gly	Met	Lys	Glu	Thr	Ala	Ala	Ala	Lys	Phe	Glu
				245					250					255	
Arg	Gln	His	Met	Asp	Ser	Pro	Asp	Leu	Gly	Thr	Gly	Gly	Gly	Ser	Gly
			260					265						270	
Asp	Asp	Asp	Asp	Lys	Ser	Pro	Met	Gly	Tyr	Arg	Gly	Ser	Thr	Pro	Asn
	275						280					285			
His	Pro	Lys	Pro	Val	Leu	Val	Pro	Lys	Thr	Gln	Asn	Asn	Leu	Gln	Ala
	290					295					300				
Gln	Asn	Val	Pro	Gln	Ala	Gln	Asn	Ala	Ser	Gln	Ala	Gln	Asn	Ala	Pro
305					310					315					320
Gln	Ala	Gln	Asn	Ala	Pro	Gln	Ala	Gln	Asn	Ala	Pro	Gln	Val	Glu	Asn
				325					330					335	
Ala	Pro	Gln	Ala	Arg	Ser	Ser	Asp	Ser	Asn	Leu	Lys	Asp	Leu	Thr	Phe
			340					345					350		
Glu	Lys	Val	Lys	His	Asn	Leu	Val	Ile	Thr	Asn	Ser	Lys	Lys	Glu	Lys
	355					360						365			
Val	Thr	Ile	Gln	Asn	Trp	Phe	Arg	Glu	Ala	Asp	Phe	Ala	Lys	Glu	Val
	370					375					380				
Pro	Asn	Tyr	Lys	Ala	Thr	Lys	Asp	Glu	Lys	Ile	Glu	Glu	Ile	Ile	Gly
385					390					395					400
Gln	Asn	Gly	Glu	Arg	Ile	Thr	Ser	Lys	Gln	Val	Asp	Asp	Leu	Ile	Ala
				405					410					415	
Lys	Gly	Asn	Gly	Lys	Ile	Thr	Gln	Asp	Glu	Leu	Ser	Lys	Val	Val	Asp
		420						425					430		
Asn	Tyr	Glu	Leu	Leu	Lys	His	Ser	Lys	Asn	Val	Thr	Asn	Ser	Leu	Asp
	435						440					445			
Lys	Leu	Ile	Ser	Ser	Val	Ser	Ala	Phe	Thr	Ser	Ser	Asn	Asp	Ser	Arg
	450					455					460				
Asn	Val	Leu	Val	Ala	Pro	Thr	Ser	Met	Arg	Ser	Thr	Pro	Asn	His	Pro
465					470					475					480
Lys	Pro	Val	Leu	Val	Pro	Lys	Thr	Gln	Asn	Asn	Leu	Gln	Ala	Gln	Asn
				485					490					495	
Val	Pro	Gln	Ala	Gln	Asn	Ala	Ser	Gln	Ala	Gln	Asn	Ala	Pro	Gln	Ala
			500					505					510		
Gln	Asn	Ala	Pro	Gln	Ala	Gln	Asn	Ala	Pro	Gln	Val	Glu	Asn	Ala	Pro
	515						520					525			
Gln	Ala	Arg	Ser	Ser	Asp	Ser	Asn	Leu	Lys	Asp	Leu	Thr	Phe	Glu	Lys
	530					535					540				

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Val	Lys	His	Asn	Leu	Val	Ile	Thr	Asn	Ser	Lys	Lys	Glu	Lys	Val	Thr
545					550					555					560
Ile	Gln	Asn	Trp	Phe	Arg	Glu	Ala	Asp	Phe	Ala	Lys	Glu	Val	Pro	Asn
				565					570						575
Tyr	Lys	Ala	Thr	Lys	Asp	Glu	Lys	Ile	Glu	Glu	Ile	Ile	Gly	Gln	Asn
			580					585						590	
Gly	Glu	Arg	Ile	Thr	Ser	Lys	Gln	Val	Asp	Asp	Leu	Ile	Ala	Lys	Gly
		595					600					605			
Asn	Gly	Lys	Ile	Thr	Gln	Asp	Glu	Leu	Ser	Lys	Val	Val	Asp	Asn	Tyr
	610					615					620				
Glu	Leu	Leu	Lys	His	Ser	Lys	Asn	Val	Thr	Asn	Ser	Leu	Asp	Lys	Leu
	625				630					635					640
Ile	Ser	Ser	Val	Ser	Ala	Phe	Thr	Ser	Ser	Asn	Asp	Ser	Arg	Asn	Val
				645					650					655	
Leu	Val	Ala	Pro	Thr	Ser	Met	Arg	Ser	Glu	Phe	Glu	Leu	Arg	Arg	Gln
			660				665						670		
Ala	Leu	Ile	Asn												
			675												

<210> SEQ ID NO 26

<211> LENGTH: 1459

<212> TYPE: DNA

<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 26

tgtccctat actaggttat tggaaaatta agggccttgt gcaaccact cgacttcttt	60
tggaatatct tgaagaaaaa tatgaagagc atttgatga gcgcatgaa ggtgataaat	120
ggcgaaacaa aaagtgtgaa ttgggtttgg agtttcccaa tcttcttat tatattgatg	180
gtgatgttaa attaacacag tctatggcca tcatacgta tatagctgac aagcacaaca	240
tgttgggtgg ttgtccaaaa gacgtgcag agatttcaat gcttgaagga gcggttttgg	300
atattagata cgggtgttcg agaattgcat atagtaaaga ctttgaaact ctcaaagttg	360
attttcttag caagctacct gaaatgctga aaatgttcga agatcgttta tgcataaaa	420
catattttaa tggatgatcat gtaaccatc ctgacttcat gttgatgac gctcttgatg	480
ttgttttata catggacca atgtgcctgg atgcgttccc aaaattagtt tgttttaaaa	540
aacgtattga agctatccca caaattgata agtacttgaa atccagcaag tatatagcat	600
ggccttttga gggctggcaa gccacgtttg gtggtggcga ccactctcca aaatcggatg	660
gttcaactag tggttctggt catcaccatc accatcactc cgcgggtctg gtgccacgcg	720
gtagtactgc aattggtatg aaagaaaccg ctgctgctaa attcgaacgc cagcacatgg	780
acagcccaga tctgggtacc ggtggtggct ccggtgatga cgacgacaag agtcccatgg	840
gatatcgggg atccacaccg aatcacccca aaccagtact agtaccaaaa acacaaaata	900
atcttcaagc acaaaatggt cctcaggcac aaaatgcctc tcaggcacaa aatgccctc	960
aggcacaaaa tgctcctcag gcacaaaatg ctctcagggt ggaaaatgct cctcaggcaa	1020
gatcctctga ttcgaaacta aaagatttaa catttgaaaa agttaaacat aatcttgtca	1080
tcacgaatag caaaaaagag aaagtgacca ttcaaaactg gttccgagag gctgattttg	1140
ctaaagaagt gcctaattat aaagcaacta aagatgagaa aatcgaagaa atcatcggtc	1200
aaaatggcga gcggtatcac tcaaagcaag ttgatgatct tatcgcaaaa ggtaacggca	1260
aaattaccca agatgagcta tcaaaagttg ttgataacta tgaattgctc aaacatagca	1320

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aaaatgtgac aaacagctta gataagttaa tctcatctgt aagtgcattt acctcgtcta 1380
atgattcgag aaatgtatta gtggctccaa cttcaatgag atccgaattc gagctccgtc 1440
gacaagcttt aattaatta 1459

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<210> SEQ ID NO 27
<211> LENGTH: 235
<212> TYPE: PRT
<213> ORGANISM: Mannheimia haemolytica

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<400> SEQUENCE: 27

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Met Gly Ser Ser His His His His His Ser Ser Gly Leu Val Pro
1          5          10          15
Arg Gly Ser His Met Ala Ser Met Thr Gly Gly Gln Gln Met Gly Arg
20        25        30
Gly Ser Thr Pro Asn His Pro Lys Pro Val Leu Val Pro Lys Thr Gln
35        40        45
Asn Asn Leu Gln Ala Gln Asn Val Pro Gln Ala Gln Asn Ala Ser Gln
50        55        60
Ala Gln Asn Ala Pro Gln Ala Gln Asn Ala Pro Gln Ala Gln Asn Ala
65        70        75        80
Pro Gln Val Glu Asn Ala Pro Gln Ala Arg Ser Ser Asp Ser Asn Leu
85        90        95
Lys Asp Leu Thr Phe Glu Lys Val Lys His Asn Leu Val Ile Thr Asn
100       105       110
Ser Lys Lys Glu Lys Val Thr Ile Gln Asn Trp Phe Arg Glu Ala Asp
115       120       125
Phe Ala Lys Glu Val Pro Asn Tyr Lys Ala Thr Lys Asp Glu Lys Ile
130       135       140
Glu Glu Ile Ile Gly Gln Asn Gly Glu Arg Ile Thr Ser Lys Gln Val
145       150       155       160
Asp Asp Leu Ile Ala Lys Gly Asn Gly Lys Ile Thr Gln Asp Glu Leu
165       170       175
Ser Lys Val Val Asp Asn Tyr Glu Leu Leu Lys His Ser Lys Asn Val
180       185       190
Thr Asn Ser Leu Asp Lys Leu Ile Ser Ser Val Ser Ala Phe Thr Ser
195       200       205
Ser Asn Asp Ser Arg Asn Val Leu Val Ala Pro Thr Ser Met Arg Ser
210       215       220
Glu Phe Glu Leu Arg Arg Gln Ala Leu Ile Asn
225       230       235

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<210> SEQ ID NO 28
<211> LENGTH: 1278
<212> TYPE: DNA
<213> ORGANISM: Mannheimia haemolytica

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<400> SEQUENCE: 28

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atgggcagca gccatcatca tcatcatcac agcagcggcc tgggtgccgcg cggcagccat 60
atggctagca tgactggtgg acagcaaatg ggctcgccgat ccacaccgaa tcaccccaaa 120
ccagtactag taccaaaaac acaaaataat cttcaagcac aaaatgttcc tcaggcacia 180
aatgcctctc aggcacaaaa tgcccctcag gcacaaaatg ctctcaggc acaaaatgct 240
cctcaggtgg aaaatgtctc tcaggcaaga tcctctgatt cgaacttaaa agatttaaca 300
tttgaaaaag ttaacataa tcttgtcatc acgaatagca aaaaagagaa agtgaccatt 360

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caaaactggt tccgagaggc tgattttgct aaagaagtgc ctaattataa agcaactaaa	420
gatgagaaaa tcgaagaaat catcggtcaa aatggcgagc ggatcacctc aaagcaagtt	480
gatgatctta tcgcaaaagg taacggcaaa attacccaag atgagctatc aaaagttggt	540
gataactatg aattgctcaa acatagcaaa aatgtgacaa acagcttaga taagttaatc	600
tcactgttaa gtgcatttac ctctgctaatt gattcgagaa atgtattagt ggctccaact	660
tcaatgagat ccacaccgaa tcaccccaaa ccagtactag taccaaaaac acaaaataat	720
cttcaagcac aaaatgttcc tcaggcacia aatgcctctc aggcacaaaa tgcccctcag	780
gcacaaaatg ctctcaggc acaaaatgct cctcagggtg aaaatgctcc tcaggcaaga	840
tcctctgatt cgaacttaaa agatttaaca ttgaaaaag ttaaacataa tcttgctatc	900
acgaatagca aaaaagagaa agtgaccatt caaaactggt tccgagaggc tgattttgct	960
aaagaagtgc ctaattataa agcaactaaa gatgagaaaa tcgaagaaat catcggtcaa	1020
aatggcgagc ggatcacctc aaagcaagtt gatgatctta tcgcaaaagg taacggcaaa	1080
attacccaag atgagctatc aaaagttggt gataactatg aattgctcaa acatagcaaa	1140
aatgtgacaa acagcttaga taagttaatc tcactgttaa gtgcatttac ctctgctaatt	1200
gattcgagaa atgtattagt ggctccaact tcaatgagat ccgaattcga gctccgtcga	1260
caagctttaa ttaattaa	1278

<210> SEQ ID NO 29

<211> LENGTH: 425

<212> TYPE: PRT

<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 29

Met Gly Ser Ser His His His His Ser Ser Gly Leu Val Pro	
1 5 10 15	
Arg Gly Ser His Met Ala Ser Met Thr Gly Gly Gln Gln Met Gly Arg	
20 25 30	
Gly Ser Thr Pro Asn His Pro Lys Pro Val Leu Val Pro Lys Thr Gln	
35 40 45	
Asn Asn Leu Gln Ala Gln Asn Val Pro Gln Ala Gln Asn Ala Ser Gln	
50 55 60	
Ala Gln Asn Ala Pro Gln Ala Gln Asn Ala Pro Gln Ala Gln Asn Ala	
65 70 75 80	
Pro Gln Val Glu Asn Ala Pro Gln Ala Arg Ser Ser Asp Ser Asn Leu	
85 90 95	
Lys Asp Leu Thr Phe Glu Lys Val Lys His Asn Leu Val Ile Thr Asn	
100 105 110	
Ser Lys Lys Glu Lys Val Thr Ile Gln Asn Trp Phe Arg Glu Ala Asp	
115 120 125	
Phe Ala Lys Glu Val Pro Asn Tyr Lys Ala Thr Lys Asp Glu Lys Ile	
130 135 140	
Glu Glu Ile Ile Gly Gln Asn Gly Glu Arg Ile Thr Ser Lys Gln Val	
145 150 155 160	
Asp Asp Leu Ile Ala Lys Gly Asn Gly Lys Ile Thr Gln Asp Glu Leu	
165 170 175	
Ser Lys Val Val Asp Asn Tyr Glu Leu Leu Lys His Ser Lys Asn Val	
180 185 190	
Thr Asn Ser Leu Asp Lys Leu Ile Ser Ser Val Ser Ala Phe Thr Ser	
195 200 205	

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Ser	Asn	Asp	Ser	Arg	Asn	Val	Leu	Val	Ala	Pro	Thr	Ser	Met	Arg	Ser
210						215					220				
Thr	Pro	Asn	His	Pro	Lys	Pro	Val	Leu	Val	Pro	Lys	Thr	Gln	Asn	Asn
225					230					235					240
Leu	Gln	Ala	Gln	Asn	Val	Pro	Gln	Ala	Gln	Asn	Ala	Ser	Gln	Ala	Gln
				245					250					255	
Asn	Ala	Pro	Gln	Ala	Gln	Asn	Ala	Pro	Gln	Ala	Gln	Asn	Ala	Pro	Gln
			260					265					270		
Val	Glu	Asn	Ala	Pro	Gln	Ala	Arg	Ser	Ser	Asp	Ser	Asn	Leu	Lys	Asp
	275						280					285			
Leu	Thr	Phe	Glu	Lys	Val	Lys	His	Asn	Leu	Val	Ile	Thr	Asn	Ser	Lys
	290					295					300				
Lys	Glu	Lys	Val	Thr	Ile	Gln	Asn	Trp	Phe	Arg	Glu	Ala	Asp	Phe	Ala
305					310					315					320
Lys	Glu	Val	Pro	Asn	Tyr	Lys	Ala	Thr	Lys	Asp	Glu	Lys	Ile	Glu	Glu
				325					330					335	
Ile	Ile	Gly	Gln	Asn	Gly	Glu	Arg	Ile	Thr	Ser	Lys	Gln	Val	Asp	Asp
		340						345					350		
Leu	Ile	Ala	Lys	Gly	Asn	Gly	Lys	Ile	Thr	Gln	Asp	Glu	Leu	Ser	Lys
	355						360					365			
Val	Val	Asp	Asn	Tyr	Glu	Leu	Leu	Lys	His	Ser	Lys	Asn	Val	Thr	Asn
	370					375					380				
Ser	Leu	Asp	Lys	Leu	Ile	Ser	Ser	Val	Ser	Ala	Phe	Thr	Ser	Ser	Asn
385				390						395					400
Asp	Ser	Arg	Asn	Val	Leu	Val	Ala	Pro	Thr	Ser	Met	Arg	Ser	Glu	Phe
				405					410					415	
Glu	Leu	Arg	Arg	Gln	Ala	Leu	Ile	Asn							
		420					425								

<210> SEQ ID NO 30

<211> LENGTH: 1278

<212> TYPE: DNA

<213> ORGANISM: Mannheimia haemolytica

<400> SEQUENCE: 30

atgggcagca gccatcatca tcatcatcac agcagcggcc tgggtgccgcg cggcagccat	60
atggctagca tgactggtgg acagcaaatg ggtcgcggt ccacaccgaa tcaccccaaa	120
ccagtactag taccaaaaac acaaaataat cttcaagcac aaaatgttcc tcaggcacia	180
aatgcctctc aggcacaaaa tgcccctcag gcacaaaatg ctctcaggc acaaaatgct	240
cctcaggtgg aaaatgtccc tcaggcaaga tcctctgatt cgaacttaaa agatttaaca	300
tttgaaaaag ttaaacataa tcttgtcatc acgaatagca aaaaagagaa agtgaccatt	360
caaaactggt tccgagaggc tgattttgct aaagaagtgc ctaattataa agcaactaaa	420
gatgagaaaa tcgaagaat catcggtcaa aatggcgagc ggatcacctc aaagcaagtt	480
gatgatctta tcgcaaaagg taacggcaaa attacccaag atgagctatc aaaagttggt	540
gataactatg aattgtctca acatagcaaa aatgtgacaa acagcttaga taagttaatc	600
tcactgttaa gtgcatttac ctctgtctaat gattcgagaa atgtattagt ggctccaact	660
tcaatgagat ccacaccgaa tcaccccaaa ccagtactag taccaaaaac acaaaataat	720
cttcaagcac aaaatgttcc tcaggcacia aatgcctctc aggcacaaaa tgcccctcag	780
gcacaaaaatg ctctcaggc acaaaatgct cctcaggtgg aaaatgtccc tcaggcaaga	840
tcctctgatt cgaacttaaa agatttaaca ttgaaaaag ttaaacataa tcttgtcatc	900

-continued

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acgaatagca aaaaagagaa agtgaccatt caaaactggt tccgagaggc tgattttgct    960
aaagaagtgc ctaattataa agcaactaaa gatgagaaaa tcgaagaaat catcggtcaa    1020
aatggcgagc ggatcacctc aaagcaagtt gatgatctta tcgcaaaagg taacggcaaa    1080
attacccaag atgagctatc aaaagttggt gataactatg aattgctcaa acatagcaaa    1140
aatgtgacaa acagcttaga taagttaatc tcattctgtaa gtgcatttac ctctgtctaat    1200
gattcgagaa atgtattagt ggctccaact tcaatgagat ccgaattcga gctccgtcga    1260
caagctttaa ttaattaa                                         1278

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<210> SEQ ID NO 31
<211> LENGTH: 242
<212> TYPE: PRT
<213> ORGANISM: Mannheimia haemolytica

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<400> SEQUENCE: 31

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Met Gly Ser Ser His His His His His Ser Ser Gly Leu Val Pro
 1              5              10              15

Arg Gly Ser His Met Ala Ser Met Thr Gly Gly Gln Gln Met Gly Arg
      20              25              30

Gly Ser Gly Gly Gly Gly Ser Gln Ala Gln Asn Ala Ser Gln Ala Gln
      35              40              45

Asn Ala Pro Gln Ala Gln Asn Ala Pro Gln Ala Gln Asn Ala Pro Gln
      50              55              60

Val Glu Asn Ala Pro Gln Ala Gln Asn Ala Pro Gln Val Glu Asn Ala
      65              70              75              80

Pro Gln Gly Gly Gly Gly Ser Phe Arg Glu Ala Glu Phe Ala Lys Thr
      85              90              95

Ile Gln Asn Tyr Val Ala Thr Arg Asp Asp Lys Ile Glu Glu Ile Ile
      100             105             110

Gly Gln Asn Gly Glu Arg Ile Gly Gly Gly Gly Ser Arg Ser Gly Gly
      115             120             125

Gly Gly Ser Gln Ala Gln Asn Ala Ser Gln Ala Gln Asn Ala Pro Gln
      130             135             140

Ala Gln Asn Ala Pro Gln Ala Gln Asn Ala Pro Gln Val Glu Asn Ala
      145             150             155             160

Pro Gln Ala Gln Asn Ala Pro Gln Val Glu Asn Ala Pro Gln Gly Gly
      165             170             175

Gly Gly Ser Phe Arg Glu Ala Glu Phe Ala Lys Thr Ile Gln Asn Tyr
      180             185             190

Val Ala Thr Arg Asp Asp Lys Ile Glu Glu Ile Ile Gly Gln Asn Gly
      195             200             205

Glu Arg Ile Gly Gly Gly Gly Ser Arg Ser Glu Phe Glu Leu Arg Arg
      210             215             220

Gln Ala Cys Gly Arg Thr Arg Ala Pro Pro Pro Pro Pro Leu Arg Ser
      225             230             235             240

Gly Cys

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<210> SEQ ID NO 32
<211> LENGTH: 727
<212> TYPE: DNA
<213> ORGANISM: Mannheimia haemolytica

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<400> SEQUENCE: 32

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tgggcagcag ccatcatcat catcatcaca gcagcggcct ggtgccgcgc ggcagccata    60

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tggctagcat gactggtgga cagcaaatgg gtcgcggatc cggtggcggc ggatctcagg 120
cacaaaatgc ctctcaggca caaaatgccc ctcaggcaca aaatgctcct caggcacaaa 180
atgctcctca ggtggaaaat gctcctcagg cacaaaatgc tcctcaggta gaaaatgctc 240
ctcaagggtgg cggtggtctcg ttccgtgaag cagagtttgc aaaaacaatt caaaattatg 300
ttgcaacaag agacgataaa attgaagaga ttatcggtca aaatggtgaa cggattggcg 360
gtggtgggtc gagatccggg ggcggcggat ctcaggcaca aaatgcctct caggcacaaa 420
atgcccccca ggcacaaaat gctcctcagg cacaaaatgc tcctcagggt gaaaatgctc 480
ctcaggcaca aaatgctcct caggtagaaa atgctcctca aggtggcggg ggctcgttcc 540
gtgaagcaga gtttgcaaaa acaattcaaa attatgttgc aacaagagac gataaaattg 600
aagagattat cggtcacaaa ggtgaacgga ttggcgggtg tgggtcgaga tccgaattcg 660
agctccgtcg acaagcttgc ggccgcactc gagcaccacc accaccacca ctgagatccg 720
gctgcta 727

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<210> SEQ ID NO 33
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: Mannheimia haemolytica

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<400> SEQUENCE: 33

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Gln Asn Ala Pro Gln
1 5

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<210> SEQ ID NO 34
<211> LENGTH: 4
<212> TYPE: PRT
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: spacer peptide

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<400> SEQUENCE: 34

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Gly Gly Gly Ser
1

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<210> SEQ ID NO 35
<211> LENGTH: 5
<212> TYPE: PRT
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic linker/spacer sequence

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<400> SEQUENCE: 35

```

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Gly Gly Gly Gly Ser
1 5

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We claim:

1. A purified chimeric protein comprising one or more copies of an isolated N-terminal immunodominant epitope of recombinant *Mannheimia haemolytica* S1 outer membrane lipoprotein (rPlpE) and one or more copies of an isolated immunodominant epitope of recombinant *Mannheimia haemolytica* leukotoxin (LKT), wherein the immunodominant epitope of the LKT is mLKT A having the amino acid sequence as set forth in SEQ ID No: 21.

2. The chimeric protein of claim 1, wherein said immunodominant epitope of said rPlpE is R2 epitope having the amino acid sequence as set forth in SEQ ID No: 19.

3. The chimeric protein of claim 1, wherein said chimeric protein further comprises a leader sequence.

4. The chimeric protein of claim 3, wherein said leader sequence is glutathione-S-transferase leader sequence.

5. The chimeric protein of claim 1, wherein said chimeric protein further comprises one or more spacer peptides.

6. The chimeric protein of claim 1, wherein said chimeric protein comprises two copies of said rPlpE and two copies of said LKT.

7. A vaccine preparation comprising at least one purified chimeric protein comprising one or more copies of an isolated N-terminal immunodominant epitope of recombinant *Mannheimia haemolytica* S1 outer membrane lipoprotein

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(rPlpE) and one or more copies of an isolated immunodominant epitope of recombinant *Mannheimia haemolytica* leukotoxin (LKT), wherein the immunodominant epitope of the LKT is mLKT A having the amino acid sequence as set forth in SEQ ID NO: 21, and a physiologically compatible carrier.

8. The vaccine preparation of claim 7, wherein said immunodominant epitope of said rPlpE is R2 epitope having the amino acid sequence as set forth in SEQ ID NO: 19.

9. The vaccine preparation of claim 7, wherein said at least one chimeric protein further comprises a leader sequence.

10. The vaccine preparation of claim 9, wherein said leader sequence is glutathione-S-transferase leader sequence.

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11. The vaccine preparation of claim 7, wherein said at least one chimeric protein further comprises one or more spacer peptides.

12. The vaccine preparation of claim 7, wherein said at least one chimeric protein comprises two copies of said rPlpE and two copies of said LKT.

13. The vaccine preparation of claim 7 further comprising an adjuvant.

* * * * *