

US007214784B2

(12) United States Patent

de la Fuente et al.

(54) PROTECTIVE ANTIGENS FOR THE CONTROL OF *IXODES* SPECIES INFESTATIONS

- (75) Inventors: Jose de Jesús de la Fuente, Stillwater, OK (US); Katherine M. Kocan, Perkins, OK (US); Consuelo García-Almazán, Stillwater, OK (US); Jose Carlos García-García, Stillwater, OK (US); Edmour F. Blouin, Perkins, OK (US)
- (73) Assignee: The Board of Regents for Oklahoma State University, Stillwater, OK (US)
- (*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 490 days.

This patent is subject to a terminal disclaimer.

- (21) Appl. No.: 10/425,563
- (22) Filed: Apr. 29, 2003
- (65) Prior Publication Data

US 2004/0022795 A1 Feb. 5, 2004

Related U.S. Application Data

- (60) Provisional application No. 60/376,251, filed on Apr. 29, 2002.
- (51) Int. Cl. *C07H 21/04* (2006.01) *A01N 25/00* (2006.01)
- (52) U.S. Cl. 536/23.5; 424/405
- (58) Field of Classification Search 536/23.5; 424/405

See application file for complete search history.

(10) Patent No.: US 7,214,784 B2

(45) **Date of Patent:** *May 8, 2007

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Primary Examiner—Jon Weber

Assistant Examiner-Rosanne Kosson

(74) Attorney, Agent, or Firm—Fellers, Snider, Blankenship, Bailey & Tippens

(57) **ABSTRACT**

Protective antigens against infestations with *Ixodes* spp. ticks, gene sequences and encoded proteins for such antigens, related vaccines and methods useful to induce an immune response, which are protective to interfere with infestations by *Ixodes* spp. ticks are presented.

5 Claims, 4 Drawing Sheets



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CDNA POOLS



Inhibition of tick infestation (%)

FIG. 2A



Inhibition of tick infestation (%)

FIG. 2B



FIG. 3

PROTECTIVE ANTIGENS FOR THE CONTROL OF *IXODES* SPECIES INFESTATIONS

CROSS REFERENCE TO RELATED APPLICATION

This application claims the benefit of copending U.S. Provisional Patent Application Ser. No. 60/376,251 filed Apr. 29, 2002.

BACKGROUND OF THE INVENTION

1. Technical Field

The present invention relates to the identification of 15 protective antigens against infestations with *Ixodes* spp. ticks, gene sequences and encoded proteins for such antigens, related vaccines and methods useful to induce an immune response, which are protective to interfere with infestations by *Ixodes* spp. ticks. 20

2. Background

Ticks parasitize wild, domesticated animals and humans and transmit pathogens including fungi, bacteria, viruses and protozoon. Currently, ticks are considered to be second in the world to mosquitoes as vectors of human diseases, but 25 they are considered to be the most important vector of pathogens in North America (Parola and Raoult, 2001). *Ixodes* spp. are distributed worldwide and act as vectors of human diseases caused by Borrelia burgdorferi (Lyme disease), Anaplasma phagocytophila (human granulocytic ehr- 30 lichiosis), Coxiella burnetti (Q fever), Francisella tularensis (tularemia), B. afzelii, B. lusitaniae, B. valaisiana and B. garinii, Rickettsia helvetica, R. japonica and R. australis, Babesia divergens and tick-borne encephalitis (TBE) and Omsk Hemorrhagic fever viruses (Estrada-Peña and Jonge- 35 jan, 1999; Parola and Raoult, 2001). Throughout eastern and southeastern United States and Canada, I. scapularis (the black legged tick) is the main vector of B. burgdorferi sensu stricto and A. phagocytophila (Estrada-Peña and Jongejan, 1999; Parola and Raoult, 2001). 40

Control of tick infestations is difficult and often impractical for multi-host ticks such as Ixodes spp. Presently, tick control is effected by integrated pest management in which different control methods are adapted to one area or against one tick species with due consideration to their environmen- 45 tal effects. Recently, development of vaccines against onehost Boophilus spp. has provided new possibilities for the identification of protective antigens for immunization against tick infestations (Willadsen, 1997; Willadsen and Jongejan, 1999; de la Fuente et al., 1999; 2000; de Vos et al., 50 2001). The recombinant B. microplus BM86 gut antigen included in commercial vaccine formulations TickGARD (Hoechst Animal Health, Australia) and Gavac (Heber Biotec S. A., Havana, Cuba) also confers partial protection against phylogenetically related Hyalomma and Rhipiceph- 55 alus tick genera (de la Fuente et al., 2000; de Vos et al., 2001). However, immunization with BM86 failed to protect against the more phylogenetically distant Amblyomma spp. (de Vos et al., 2001). These results suggest that using Bm86 or a closely related gene for the production of vaccines 60 against Ixodes spp. or other tick genera phylogenetically distant from Boophilus spp. (Black and Piesman, 1994) could be impractical. Therefore, the screening for novel protective antigens is necessary to identify vaccine candidates against infestations with these tick species of medical 65 and veterinary importance. Control of ticks by vaccination would avoid environmental contamination and selection of

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drug resistant ticks that result from repeated acaricide application (de la Fuente et al., 1998; Garcia-Garcia et al., 1999). Anti-tick vaccines also allow for inclusion of multiple antigens in order to target a broad range of tick species and for incorporation of pathogen-blocking antigens.

Vaccination with DNA and cDNA molecules has been used to induce a protective immune response against B. microplus and several pathogens in laboratory animals and livestock (De Rose et al., 1999; Drew et al., 1999; van Drunen Littel-van den Hurk et al., 2001; Kofta and Wedrychowicz, 2001). A new technique, expression library immunization (ELI) in combination with sequence analysis provides an alternative approach for identification of potential vaccine antigens based on rapid screening of the expressed genes without prior knowledge of the antigens encoded by cDNA clones. ELI was first reported for Mycoplasma pulmonis (Barry et al., 1995) and since then has been used for unicellular and multicellular pathogens and viruses (Manoutcharian et al., 1998; Alberti et al., 1998; Brayton et al., 1998; Melby et al., 2000; Smooker et al., 2000; Moore et al., 2002; Singh et al., 2002). However, the identification of individual protective clones has not been reported and it is predicted that identification of protective antigens will be more difficult as the complexity of the genome increases.

Although several reports in the literature have demonstrated by ELI that libraries can offer a degree of protection (Barry et al., 1995; Manoutcharian et al., 1998; Alberti et al., 1998; Brayton et al., 1998; Melby et al., 2000; Smooker et al., 2000; Moore et al., 2002; Singh et al., 2002), none have applied ELI to arthropods and particularly to ticks. Several vaccines have been developed to protect humans against Ixodes-transmitted pathogens including TBE virus and B. burgdorferi. However, it is not clear whether these vaccines will protect against all pathogen strains and genotypes. The inclusion of tick immunogens in pathogen-specific vaccines could enhance their protective effect and increase efficacy (Nuttall, 1999). This transmission-blocking approach is supported by evidence that host resistance to ticks provides some protection against tick-borne transmission of viruses and B. burgdorferi (Wikel et al., 1997). Furthermore, vaccination against B. microplus has been demonstrated to contribute to the control of tick-borne diseases (de la Fuente et al., 1998; 1999).

SUMMARY OF THE INVENTION

The present invention is based upon our identification by ELI and sequence analysis of protective cDNA clones against experimental infestations with I. scapularis. This is the first example of the application of ELI to arthropods and particularly to ticks. The protective antigens are homologous to endopeptidases, nucleotidases, chorion proteins, vitellogenin receptors, peptidoglycan recognition proteins, glutamine-alanine rich proteins, ribosomal proteins, β -adaptin, Beta-amyloid precursor protein, Block of proliferation (Bop1), lectins, chloride channels, RNA polymerases, ATPases and heat-shock proteins. These antigens induce an immune response in vaccinated hosts that either interferes with tick development or results in a pro-feeding activity, which could be due to the expression of cDNAs encoding for tick immunosuppressants, anticoagulants and other proteins with low antigenicity and a pro-feeding activity or they could encode for proteins homologous to host proteins with anti-tick activity, which neutralization results in a tick profeeding activity. These protective antigens, although identified for I. scapularis, may be cross protective between

Ixodes species considering the high degree of conservation of gene sequences and protein function between species of the same genus. A 5'-nucleotidase was identified and characterized in B. microplus by Liyou et al. (1999; 2000) but they did not assay its protection capacity. Although surprising at first glance, the protection capacity of ribosomal and heat shock protein preparations has been previously documented in other organisms (Elad and Segal, 1995; Silva, 1999; Melby et al., 2000; Cassataro et al., 2002) but never in ticks. The effect of cDNA vaccination on I. scapularis experimental infestations of mice was evidenced by the reduction of the number of engorged larvae, the retardation of larval development, the inhibition of molting to nymphal stages and the appearance of visibly damaged larvae with red coloration. These effects were also recorded in vaccination experiments with recombinant BM86 and BM95 against infestations with B. microplus, including the red coloration in some ticks, attributed to blood leakage to the tick haemolymph (Garcia-Garcia et al., 2000).

Thus, in one embodiment of the present invention there is provided cDNA sequences, protein encoding fragments thereof, and derived protein sequences for protective I. scapularis antigens comprising antigens homologous to endopeptidases, nucleotidases, chorion proteins, vitelloge- 25 nin receptors, peptidoglycan recognition proteins, glutamine-alanine rich proteins, ribosomal proteins, β-adaptin, Beta-amyloid precursor protein, Block of proliferation (Bop1), lectins, chloride channels, RNA polymerases, ATPases and heat-shock proteins.

In another embodiment of the present invention there is provided a vaccine composition comprising the I. scapularis protective recombinant proteins and/or modified cDNAs separately or which may optionally be combined with adjuvant to enhance the protection efficacy of vaccine preparations against Ixodes spp., wherein the vaccine composition further comprises a pharmaceutically acceptable carrier or diluent. The vaccine composition also may optionally be combined with tick-borne pathogen components to provide a means to control tick-borne infections, wherein the vaccine composition further comprises a pharmaceutically acceptable carrier or diluent and adjuvant.

In another embodiment of the present invention there is provided a method for inducing an immune response in a 45 mammal to provide immune protection, which reduces or affects infestations by Ixodes spp. ticks and/or transmission of tick-borne pathogens, the method comprising administering to at-risk human population and mammalian reservoir an effective amount of a vaccine composition comprising the I. 50 scapularis protective recombinant proteins and/or modified cDNAs alone or in combination with an adjuvant or tickborne pathogen components to provide a means to control tick infestations and to reduce transmission to humans of tick-borne infections, wherein the vaccine composition fur- 55 ther comprises a pharmaceutically acceptable carrier or diluent.

A better understanding of the present invention and its objects and advantages will become apparent to those skilled in this art from the following detailed description, wherein 60 there is described only the preferred embodiment of the invention, simply by way of illustration of the best mode contemplated for carrying out the invention. As will be realized, the invention is capable of modifications in various obvious respects, all without departing from the scope and 65 spirit of the invention. Accordingly, the description should be regarded as illustrative in nature and not as restrictive.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a summary of the cDNA ELI approach used to identify protective antigens against I. scapularis infestations.

FIG. 2A is a graph depicting the results of a primary screen of cDNA pools (A-H 1-4, A5) by ELI. V, control mice injected with 1 μ g vector DNA alone. * α <0.01, ** α <0.05 (Tukey's post-hoc test for pair comparisons after ANOVA). Number in boxes represent values for inhibition of tick infestation with respect to the control group.

FIG. 2B is a graph depicting the results of a primary screen of cDNA pools (A6-A10, B-H 5-8) by ELI. V, control mice injected with 1 µg vector DNA alone. $\alpha < 0.01$, ** α <0.05 (Tukey's post-hoc test for pair comparisons after ANOVA). Number in boxes represent values for inhibition of tick infestation with respect to the control group.

FIG. 3 is a graph depicting the results of a tertiary screen by ELI of cDNA sub-pools formed according to the pre-20 dicted function of encoded proteins. Only groups with $I \ge 15\%$ are shown (white bars). The number of engorged larvae per mouse is expressed as mean±SD (black bars). Control mice were injected with mitochondrial (MT) cDNAs. *P≦0.05 (Student's t-test).

DETAILED DESCRIPTION OF THE INVENTION

Before explaining the present invention in detail, it is 30 important to understand that the invention is not limited in its application to the details of the construction illustrated and the steps described herein. The invention is capable of other embodiments and of being practiced or carried out in a variety of ways. It is to be understood that the phraseology and terminology employed herein is for the purpose of description and not of limitation.

The present invention derives from the sequences set forth on the Sequence Listing attached hereto and incorporated herein. In particular, there is provided 25 separate and distinct sequences comprising 14 cloned cDNA molecules and 11 deduced amino acid sequences of encoded polypeptides, said sequences having been isolated and identified as possessing the asserted utility in accordance with the following described experimental methodology.

EXAMPLE 1

Construction of an I. scapularis cDNA Library and Screening for Protective Antigens by ELI

Tick Cells

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Monolayers of IDE8 (ATCC CRL 1973) cells, originally derived from embryonic I. scapularis, were maintained at 31° C. in L-15B medium supplemented with 5% foetal bovine serum, tryptose phosphate broth and bovine lipoprotein concentrate after Munderloh et al. (1994). Cells were subcultured at 1:5-1:10 when monolayers reached a density of approximately 107 cells/T-25 flask. Medium was replaced weekly.

Library Construction

A cDNA expression library was constructed in the vector pEXP1 containing the strong cytomegalovirus CMV_{IF} promoter (Clontech). Because we planned to target the early larval stages of I. scapularis, we chose to construct our library from cultured embryonic I. scapularis IDE8 cellsderived poly(A)+ RNA. The cDNA library contained 4.4×

 10^6 independent clones and a titer of approximately 10^{10} cfu/ml with more than 93% of the clones with cDNA inserts. The average cDNA size was 1.7 kb (0.5–4.0 kb).

Primary Screen

The overall schema for identification of protective antigens through ELI, sequential fractionation and sequence analysis is shown in FIG. 1.

Ninety-six LBA (master) plates containing an average of 41 (30–61) cDNA clones per plate were prepared. Replicas were made and clones from each plate were pooled, inoculated in Luria-Bertani with 50 µg/ml ampicillin, grown for 2 hr in a 96 wells plate and plasmid DNA purified from each pool (Wizard SV 96 plasmid DNA purification system, Promega, Madison, Wis., USA). BALB/c female mice, 5-6 weeks of age at the time of first vaccination, were used. Mice were cared for in accordance with standards set in the Guide for Care and Use of Laboratory Animals. Mice were injected with a 1 ml tuberculin syringe and a 27 gauge needle at days 0 and 14. Three mice per group were each immunized IM in 20 the thigh with 1 µg DNA/dose in 50 µl PBS. Two groups of 3 mice each were included as controls. One group was injected with 1 µg vector DNA alone and the second with saline only. Two weeks after the last immunization, mice were infested with 100 I. scapularis larvae per mouse. Ticks 25 were artificially reared at the Oklahoma State University tick rearing facility by feeding larvae on mice, nymphs on rabbits and adults on sheep and using for infestation in our experiments the larvae obtained from the eggs oviposited by a single female. Twelve hours after tick infestation, larvae that 30 did not attach were counted to calculate the number of attached larvae per mouse and mice were transferred to new cages. Replete larvae dropping from each mouse were collected daily and counted during 7 days. The inhibition of tick infestation (I) for each test group was calculated with 35 respect to vector-immunized controls as [1-(<RL>n/ <RL>c×<RL>ic/<RL>in)]×100, where <RL>n is the average number of replete larvae recovered per mouse for each test group, <RL>c is the average number of replete larvae recovered per mouse for control group, <RL>ic is the 40 average number of larvae attached per mouse for control group, and <RL>in is the average number of larvae attached per mouse for each test group.

Pools of 41 (30-61) I. scapularis cDNA clones were screened by ELI. Only 33 cDNA pools and controls were 45 analyzed per experiment. The average tick infestation level was 50 ± 13 and 56 ± 15 and 56 ± 15 and 54 ± 18 larvae/mouse for cDNA immunized and control mice, respectively (P>0.05) (Table 1). The average number of engorged larvae recovered per mouse was 9±3 and 13±4 in the cDNA- 50 immunized mice and 16±4 and 17±3 in the control vectorimmunized group (P<0.05) (Table 1). No reduction was observed in the number of larvae collected from mice that received the vector DNA compared to saline-immunized controls. The maximum number of engorged larvae was 55 collected 3 to 4 days after infestation. However, in mice immunized with cDNA pools B5, A8 and A10 (FIG. 2) a retardation of larval development in 1 to 2 days was recorded. The average inhibition of tick infestation (I) was 49±28% and 30±22% (Table 1). After two experiments 60 covering the analysis of 66 pools (2705 clones), 9 protective pools (351 clones) were selected producing an inhibition of tick infestation $I \ge 60\%$ (FIGS. 2A and 2B and Table 1). When we started these experiments, we planed to screen over 4000 cDNA clones considering the complexity of the 65 tick genome. However, to our surprise 9 protective cDNA pools were identified after screening 66 pools containing

2705 cDNA clones. This result probably reflects the possibility of interfering with tick infestations at many different levels that involve a Pleiades of gene products. Results from vaccination experiments against ticks employing recombinant antigens support this view (reviewed by Mulenga et al., 2000). Because of the complexity of the screening procedure in mice vaccinated and challenged with tick larvae, it was difficult to work with more than 9 protective cDNA pools. Therefore we did not continue screening new cDNA pools and focused our attention on the 9 pools selected after the primary screen.

Secondary Screen

The secondary screen was done to verify the protective capacity of the cDNA pools selected after the primary screen (FIGS. 2A and 2B). After the primary screen of 66 cDNA pools (2705 clones), 9 pools with I≥60% were selected for the secondary screen (re-screening) employing 5 mice per group as described above. Engorged larvae were kept for molting in a 95% humidity atmosphere. Molting of engorged larvae was evaluated by visual examination of tick nymphs under a stereomicroscope 34 days after last larval collection. The inhibition of molting (M) for each test group was calculated with respect to vector-immunized controls as [1-(MLn/MLc×RLc/RLn)]×100, where MLn is the number of nymphs for each test group, MLc is the number of nymphs for the control group, RLc is the number of larvae recovered for the control group, and RLi is the number of larvae recovered for each test group. Control mice were immunized with the negative (I=0%) F2 cDNA pool or saline only. A group was included immunized SC with two doses of 100 µg of total IDE8 tick cell proteins per dose in Freund's incomplete adjuvant.

All 9 protective cDNA pools gave positive results in the secondary screen (data not shown). The tick infestation levels were higher in this experiment (average 85 ± 6 and 84 ± 3 larvae/mouse for cDNA-immunized and control mice, respectively; P>0.05). Nevertheless, the average number of engorged larvae recovered per mouse was 39 ± 7 and 26 ± 6 for control and cDNA-immunized mice, respectively (P<0.05). The group immunized with total IDE8 tick cell proteins was protected with I=33%. Again, no reduction was observed in the number of larvae collected from mice that received the control cDNA (F2 negative pool after the primary screen; FIG. **2**A) compared to saline-immunized controls.

In the secondary screen, molting of engorged larvae was evaluated after 34 days. Molting was affected in all but one test cDNA-immunized group. Inhibition of molting in test cDNA-immunized mice compared to the control cDNAimmunized group varied from 0% to 12% (6±4%). The inhibition of molting was higher than 50% only in the larvae collected from mice immunized with cDNA pools B5 and A10, which showed a retardation of larval development in 1 to 2 days as in the primary screen. No differences were observed between control cDNA and saline-immunized mice. Among the larvae that did not molt to nymph, some were visibly damaged and presented a strong red coloration. The percent of red larvae in cDNA-immunized mice varied between 3% to 18% (7±5%) while in the saline and control cDNA-immunized groups red larvae represented the 6% and 4%, respectively.

Tertiary Screen

For the tertiary screen, 64 clones were grouped in 16 sub-pools each containing 1 to 17 plasmids according to the predicted function of encoded proteins (e.g., all the plasmids that encoded histone proteins were grouped together) and

4∩

used with 4 sub-pools containing 182 clones of unknown function or with sequences without homology to sequence databases to immunize 4 mice per group. Mice were immunized with 0.3 μ g/plasmid/dose in 50 μ l PBS and evaluated as described above. Control mice were immunized with a 5 pool of 20 plasmids containing mitochondrial cDNAs.

Tick infestation levels were similar in all test groups (72±2 larvae/mouse) and in control mice (69±2 larvae/ mouse) (P>0.05). The number of engorged larvae recovered per mouse was also similar between test (16 ± 7) and control 10 (14±6) mice (P>0.05). However, the groups immunized with cDNA sub-pools containing clones with putative endopeptidase, nucleotidase, ribosomal proteins, heat shock proteins, glutamine-alanine-rich proteins and 3 of the sub-pools with unknown function or with sequences without homology to sequence databases had $I \ge 15\%$ (FIG. 3). Furthermore, among them, the groups immunized with sub-pools containing clones with a putative endopeptidase, nucleotidase and two of the cDNA sub-pools with unknown function or with sequences without homology to sequence databases resulted 20 in lower infestation levels compared to control mice $(P \le 0.05)$ and $I \ge 40\%$ (FIG. 3). Clones homologous to chorion proteins, vitellogenin receptors, and peptidoglycan recognition proteins were selected for they potential protection capacity in other stages of tick development.

Statistical Analysis

The number of larvae attached per mouse and the number of engorged larvae recovered per mouse 7 days after infestation were compared by Analysis of Variance (ANOVA) followed by a series of Tukey's post-hoc tests for pair comparisons between cDNA-immunized and control vector DNA-immunized mice (primary screen), and by Student's t-test between mice immunized with positive cDNA pools and the control negative F2 cDNA pool (secondary screen) or between test cDNA sub-pools-immunized and control mice immunized with mitochondrial cDNAs (tertiary ³⁵ screen).

EXAMPLE 2

Sequence Analysis of Protective Clones

All the 351 cDNA clones in the 9 pools that resulted positive in the secondary screen were sequenced. DNA from individual clones in these pools was purified (Wizard SV 96 plasmid DNA purification system, Promega) from the mas-45 ter plate and partially sequenced. In most cases a sequence larger than 700 nucleotides was obtained. Nucleotide sequences were analyzed using the program AlignX (Vector NTI Suite V 5.5, InforMax, North Bethesda, Md., USA). BLAST (Altschul et al., 1990) was used to search the NCBI 50 databases to identify previously cloned sequences that may have homology to those that we sequenced. Sequence analysis allowed grouping the clones according to sequence identity to DNA databases and predicted protein function. The protective clones selected after the tertiary screen were 55 fully sequenced.

Comparison to sequence databases permitted to identify sequence identity to previously reported genes with known function in 152 (43%) of the clones (Table 2). Fifty seven percent of the sequences were homologous to genes with unknown function or had no significant identity to previously reported sequences (Table 2). Of the clones with sequence identity to genes with known function, 85% were homologous to arthropod sequences. Ninety-three clones (61%) contained sequences homologous to *Drosophila melanogaster*, 5 (3%) to other insects and 32 (21%) to Ixodid 65 tick species. Thirty percent of the clones were eliminated from further analysis based on their sequence identity,

including those containing similar sequences (Table 2). The protective clones included antigens homologous to endopeptidases, nucleotidases, chorion proteins, vitellogenin receptors, peptidoglycan recognition proteins, glutamine-alanine rich proteins, ribosomal proteins, and heat-shock proteins.

SUMMARY OF RESULTS

The results obtained with the various protective clones identified in the Sequence Listing, along with certain selected expressed proteins, are summarized in Table 4.

SEQ ID NO:1 denotes the clone designated 4E6, wherein the relevant protein encoding fragment has been identified as comprising residues 1-117, which encodes the polypeptide shown in SEQ ID NO: 2.

SEQ ID NO:3 denotes the clone designated 4D8, wherein the relevant protein encoding fragment has been identified as comprising residues 80–575, which encodes the polypeptide shown in SEQ ID NO: 4.

SEQ ID NO:5 denotes the clone designated 4F8, wherein the relevant protein encoding fragment has been identified as comprising residues 1–951, which encodes the polypeptide shown in SEQ ID NO: 6.

SEQ ID NO:7 denotes the clone designated 4G11, wherein the relevant protein encoding fragment has been identified as comprising residues 1–697, which encodes the polypeptide shown in SEQ ID NO: 8.

SEQ ID NO:9 denotes the clone designated 4D6, wherein the relevant protein encoding fragment has been identified as comprising residues 198–1025, which encodes the polypeptide shown in SEQ ID NO: 10.

SEQ ID NO:11 denotes the clone designated 3E1, wherein the relevant protein encoding fragment has been identified as comprising residues 3–578, which encodes the polypeptide shown in SEQ ID NO: 12.

SEQ ID NO:13 denotes the clone designated 1C10, wherein the relevant protein encoding fragment has been identified as comprising residues 1–1119, which encodes the polypeptide shown in SEQ ID NO: 14.

SEQ ID NO:15 denotes the clone designated 3E10, wherein the relevant protein encoding fragment has been identified as comprising residues 51-1544, which encodes the polypeptide shown in SEQ ID NO: 16.

SEQ ID NO:17 denotes the clone designated 4F11, wherein the relevant protein encoding fragment has been identified as comprising residues 31–2295, which encodes the polypeptide shown in SEQ ID NO: 18.

SEQ ID NO:19 denotes the clone designated 3C12, wherein the relevant protein encoding fragment has been identified as comprising residues 6–332, which encodes the polypeptide shown in SEQ ID NO: 20.

SEQ ID NO:21 denotes the clone designated 2C12, wherein the relevant protein encoding fragment has been identified as comprising residues 3–137, which encodes the polypeptide shown in SEQ ID NO: 22.

SEQ ID NOS: 22, 23 AND 24, denote, respectively, clones 1A9, 1B2 and 4A4, each comprising a partial sequence with no associated polypeptide.

* * * * *

As noted above, the present invention relates to the sequences identified in the Sequence Listing. More generally, the invention concerns the given cDNA sequences and any nucleotide sequence coding for a protein which is capable of eliciting an antibody or other immune response (e.g., T-cell response of the immune system) which recognizes an epitope(s) of the amino acid sequences depicted in the Sequence Listing, including less than the full cDNA sequences and mutants thereof. Hence the nucleotide

sequence may encode a protein which is the entire antigen encoded by the variously identified bases, or a fragment or derivative of the antigen or a fusion product of the antigen or fragment and another protein, provided that the protein which is produced from such sequence is capable of eliciting 5 an antibody or other immune response which recognizes an epitope(s) of the given amino acid sequences.

As a result, the invention encompasses DNA sequences which encode for and/or express in appropriate transformed cells, proteins which may be the full length antigen, antigen 10 fragment, antigen derivative or a fusion product of such antigen, antigen fragment or antigen derivative with another protein.

Proteins included within the present invention have an amino acid sequence depicted in the Sequence Listing. ¹⁵ Other included proteins consist of a fragment of said sequence capable of eliciting an antibody or other immune response which recognizes an epitope(s) of the amino acid sequences depicted and a mutuant of said sequence capable of eliciting an antibody or other immune response which recognizes an epitope(s) of such amino acid sequences.²⁰

The nucleotide sequences may be inserted into any of a wide variety of expression vectors by a variety of procedures. Such procedures and others are deemed to be known by those skilled in the art. Suitable vectors include chromo-25 somal, nonchromosomal and synthetic DNA sequences; e.g., derivatives of SV40; bacterial plasmids; phage DNAs; yeast plasmids; vectors derived from combinations of plasmids and phage DNAs, viral DNA such as baculovirus, vaccinia, adenovirus, fowl pox virus, pseudorabies, etc. The appropriate DNA sequence must be operatively linked in the 30 vector to an appropriate expression control sequence(s) (promoter) to direct mRNA synthesis. As representative examples of such promoters, there may be mentioned LTR or SV40 promoter, the E. coli lac or trp, the phage lambda PL promoter and other promoters known to control expres- 35 sion of genes in prokaryotic and eukaryotic cells or their viruses. The expression vector also includes a non-coding sequence for a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression. 40

The vector containing the appropriate cDNA sequence as hereinabove described, as well as an appropriate promoter or control sequence, may be employed to transform an appropriate host to permit the host to express the protein. Examples of host organisms and cells include bacterial 45 strains (e.g., *E. coli, Pseudomonas, Bacillus, Salmonella*, etc.), fungi (e.g., yeasts and other fungi), animal or plant hosts (e.g., mouse, swine or animal and human tissue cells). The selection of the host is deemed to be within the scope of those skilled in the art.

It is also understood that the appropriate cDNA sequence present in the vector when introduced into a host may express part or only a portion of the protein which is encoded within the noted terminology, it being sufficient that the expressed protein be capable of eliciting an antibody or other immune response which recognizes an epitope(s) of the listed amino acid sequences.

The isolated cDNAs and/or polypeptide expressed by the host transformed by the vector may be harvested by methods which will occur to those skilled in the art and used in a vaccine for protection of a mammal, such as a bovine, swine, ⁶⁰ human, etc., against infestations of *Lxodes* species. Such protective recombinant proteins and/or modified cDNAs are used in an amount effective to induce an immune response against *Lxodes* species ticks and their associated pathogens and may be used in combination with a suitable physiologi- ⁶⁵ cally acceptable carrier. The term "inducing an immune response" when used with respect to the vaccine described

herein means that the vaccine prevents disease associated with a particular tick species or reduces the severity of the disease.

The carrier employed in conjunction with vaccine may be any one of a wide variety of carriers. As representative examples of suitable carriers, there may be mentioned mineral oil, synthetic polymers, etc. Carriers for vaccines are well known in the art and the selection of a suitable carrier is deemed to be within the scope of those skilled in the art. The selection of a suitable carrier is also dependent upon the manner in which the vaccine is to be administered.

The present invention provides a method of immunizing a susceptible mammal, against infestations and disease caused by *Ixodes* species with the vaccine described above. For purposes of this invention, the vaccine is administered in an effective amount. The vaccine may be administered by any of the methods well known to those skilled in the art, for example, by intramuscular, subcutaneous, intraperitoneal or intravenous injection. Alternatively, the vaccine may be administered intranasally or orally. It is also to be understood that the vaccine may include active components, such as tick-borne pathogen components or adjuvants in addition to the antigen(s) or fragments hereinabove described.

The host expressing the antigen may itself be used to deliver antigen to non-human animals, by introducing killed or viable host cells that are capable of propagating in the animal. Direct incorporation of the cDNA sequences into host cells may also be used to introduce the sequences into animal cells for expression of antigen in vivo.

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TABLE 1

	Primary scree	en of the I. scapular	is cDNA library	by ELI in mice.	
Experimental group ^a	Number of pools screened (Number of clones)	Average ± SD number of larvae attached per mouse ^b	Average ± SD number of engorged larvae per mouse ^c	Average ± SD inhibition of tick infestation (I) ^d	Number of pools selected for the secondary screen
Experiment 1	33 (1383)	50 ± 13 (33–80)	9 ± 3 (2–42)	$39 \pm 55\%$ (-183-87%)	6 (I > 75%)
Vector DNA- immunized controls for experiment 1	—	56 ± 13 (45–67)	16 ± 4 (5–27)	_	_
Experiment 2	33 (1322)	56 ± 15 (29–79)	13 ± 4 (1–27)	27 ± 28% (-53- 89%)	3 (I > 60%)

TABLE 1-continued

Experimental group ^a	Primary scree Number of pools screened (Number of clones)	n of the <i>I. scapular</i> Average ± SD number of larvae attached per mouse ^b	ris cDNA library Average ± SD number of engorged larvae per mouse ^c	by ELI in mice. Average \pm SD inhibition of tick infestation (I) ^d	Number of pools selected for the secondary screen
Vector DNA- immunized controls for experiment 2	—	54 ± 18 (36–73)	17 ± 3 (6–28)		

^aNinety six LBA plates containing an average of 41 cDNA clones per plate were prepared. Replicas were made and clones from each plate were pooled, inoculated, grown for 2 hr in a 96 wells plate and plasmid DNA purified from each pool for ELI. Three mice per group were each immunized IM twice with 1 µg DNA/dose in 50 µl PBS two weeks apart. Two groups of 3 mice each were included as controls. One group was injected with vector DNA and the second with saline only. ^bFifteen days after the last immunization, mice were infested with 100 *I. scapularis* larvae per

mouse. Twelve hrs later, larvae that did not attach were counted to calculate the number of "Engorged larvae per mouse and mice were transferred to new cages. "Engorged larvae dropping from each mouse were collected daily and counted after 7 days.

^dThe inhibition of tick infestation (I) for each test group was calculated with respect to vectorimmunized controls as [1-(RLn/RLc × RLic/RLin)] × 100, where RLn is the average number of replete larvae recovered per mouse for each test group, RLc is the average number of replete larvae recovered per mouse for control group, RLic is the average number of larvae attached per mouse for control group, and RLin is the average number of larvae attached per mouse for eachtest group.

TABLE 2

	Putative protein Function	Number of clones		Sub-pool (No. of clones)	Clone	Pool
_	Dicounthatica	2	35	* * /	3G0_3G10	E3
	Catabolism	2			4D11 4D12 4E7 4E7	EJ E1
	Cell adhesion	2		Membrane protein (7)	1D8 1D11 1E10	D1
	Cell cycle ^a	2		Wiembrane protein (7)	2B12	A10
	Cytoskeletala	28			2812	E8
	Defense	2			309	B4
	DNA structure or replication ^a	3	40		3G11	E3
	Extracellular matrix	3		ATPase (6)	1A9, 1B2, 1C9	A5
	Endocvtosis	2		(-)	2C9	A10
	Energy metabolism	10			4A4	C3
	Homeostasis	2			4G12	F1
	Morphogenetic	9		Cell channel/Transporter (5)	1F4	D1
	Mitochondriala	34	45	1 ()	2H11	E8
	Protein synthesis or processing ^{a,b}	34			4A12	C3
	RNA synthesis or processing ^a	7			4G10, 4G11	F1
	Heat-shock proteins	4		Early development-specific (4)	1C8	A5
	Signal transduction	16			3F4	E3
	Transport	8			4C7	C3
	Unknown	199	50		4G9	F1
				G protein-coupled receptor (4)	2B7, 2C12	A10
	Total	351			2F12	E8
			-		4C9	C3
^a El	iminated from further screening of protect	tive antigens. Other clones		Growth factor receptor (3)	2E8	B5
we	re eliminated for containing similar seque	nces.		1 ()	3B8, 3C8	B4
°Ez	scept for ribosomal proteins.		55	Lectin (3)	3E10	E3
				<>		

60

EGF-like (2)

Adaptin (1)

Secreted protein (2)

65 Glutamine-Alanine rich (2)

30

TA	RI	E.	3
- X X X	பா	~	~

Grouping of the clones according to the predicted function of encoded proteins in sub-pools for the tertiary screen.				
Sub-pool (No. of clones)	Clone	Pool ^a		
Ribosomal (17)	1A2,1A10,1C11	A5		
	1F6	D1		
	2B8	A10		
	2F8, 2F10	E8		
	3A10, 2C3, 3D2, 3D10	B4		

	(D11, (D12, (D7, 11)	
Membrane protein (7)	1D8, 1D11, 1E10	D1
	2B12	A1
	2H5	E8
	3C9	B4
	3G11	E3
ATPase (6)	1A9, 1B2, 1C9	A5
	2C9	A1
	4A4	C3
	4G12	F1
Cell channel/Transporter (5)	1F4	D1
	2H11	E8
	4A12	C3
	4G10, 4G11	F1
Early development-specific (4)	1C8	A5
	3F4	E3
	4C7	C3
	4G9	F1
G protein-coupled receptor (4)	2B7, 2C12	A1
	2F12	E8
	4C9	C3
Growth factor receptor (3)	2E8	B5
	3B8, 3C8	B4
Lectin (3)	3E10	E3
	4B8, 4C8	C3
Vitellogenin (3)	1F12	D1
5 ()	4A6	C3
	462	F1
Heat shock (3)	1C10	A5
	1F10	D1
	11 10	

3F6

2H4

4C10

2F9

3C12

3E1

4D6, 4E6

E3

E8

C3

E8

Β4

F1

E3

20

25

30

TABLE 3-continued

Grouping of the clones encoded proteins in	Grouping of the clones according to the predicted function of encoded proteins in sub-pools for the tertiary screen.		
Sub-pool (No. of clones)	Clone	Poolª	
Endopeptidase (1)	4D8	F1	
Nucleotidase (1)	4F8	F1	

^acDNA pools refer to positive pools after primary and secondary screens (FIG. 2Å and 2B).

TABLE	4
	/ T

cDNA clone	Predicted Protein	Inhibition of tick infestation I (%)	Inhibition of molting M %	Efficacy E (%)
4D8	Endopeptidase	40*/54**	7*/8**	44*/58**
4F8	Nucleotidase	50*/64**	17*/-9**	58*/61**
1C10	HSP70	17*	ND	ND
4D6	Glu-Ala-rich	61*	11	66*
4E6	Glu-Ala-rich	20*/46**	16**	55**
3E1	β-adaptin (appendage region)	27*	5*	31*
2C12	Beta-amyloid precursor protein (APP)	-8***	ND	ND
4F11	Block of proliferation Bop1	-39***	ND	ND
3E10	Mannose binding lectin	-48*/-10***	ND	ND
4G11	Chloride channel	38***	30	57

16

TABLE 4-continued

	_	Summary of results with	h I. scapularis	cDNA clone	<u>s.</u>
5	cDNA clone	Predicted Protein	Inhibition of tick infestation I (%)	Inhibition of molting M %	Efficacy E (%)
10	3C12 1A9, 1B2, 4A4	RNA polymerase III ATPase	-104*** -57***	ND ND	ND ND

Mice were immunized with cDNA-containing expression plasmid DNA as described above (*) or with 100 µg/dose of recombinant protein expressed in E. coli (**). I, M and E were calculated as described above. ND, not

15 determined. ***Resulted in a pro-feeding activity. This effect could be due to the expression of cDNAs encoding for tick immunosuppressants, anticoagulants and other proteins with low antigenicity and a pro-feeding activity. Alternatively, they could encode for proteins homologous to host proteins with anti-tick activity, which neutralization results in a tick pro-feeding activity.

In view of the above, it will be seen that the several objectives of the invention are achieved and other advantageous results attained. As various changes could be made in the above DNA molecules, proteins, etc. without departing from the scope of the invention, it is intended that all matter contained in the above description or shown in the accompanying drawings shall be interpreted as illustrative and not in a limiting sense. While the invention has been described with a certain degree of particularity, it is understood that the invention is not limited to the embodiment(s) set for herein for purposes of exemplification, but is to be limited only by the scope of the attached claim or claims, including the full range of equivalency to which each element thereof is entitled.

<160> NUMBER OF SEQ ID NOS: 25 <210> SEQ ID NO 1 <211> LENGTH: 349 <212> TYPE: DNA <213> ORGANISM: Ixodes scapularis <400> SEQUENCE: 1 atggaaatat ctgtgaaacc aaggcccaca aaaaggaaaa gaaaggccat catcatcatg 60 gcaagaatga gaacagcatt ccccaccaga agtgggaaca gcttctcaag gacttgaaca 120 gttaatgatg tgttgtgcaa ttcgaatgtg gctgcaacct cgctagagaa catagtcgac 180 cagctgtagt gctctagtat taaccaagaa gcagtattct gccgtcatat gtacaggcag 240 atttgttacg gcattttcag cttttttta tacaaaatgt agttcttgtt taaaaaaaac 300 349 <210> SEQ ID NO 2 <211> LENGTH: 38

SEQUENCE LISTING

<212> TYPE: PRT <213> ORGANISM: Ixodes scapularis

<400> SEQUENCE: 2

Met Glu Ile Ser Val Lys Pro Arg Pro Thr Lys Arg Lys Ala 5 10 15

-continued

Ile	Ile	Ile	Met 20	Ala	Arg	Met .	Arg	Thr 25	Ala F	he	Pro Thr	Arc 30	g Ser Gly	
Asn	Ser	Phe 35	Ser	Arg	Thr									
<210 <211 <212 <213 <220 <221 <222 <223 <220 <221 <222 <223 <222 <223	 > SE > LE > TY > OF > NA > CI > FE > NA > LC > OT > FE > NA > SE 	Q II NGTH PE: GANJ ATUF ME/H CATJ HER ATUF ME/H CATJ HER	O NO H: 26 DNA ISM: (EY: : INFO RE: (EY: : INFO INFO	3 93 Ixod (685 RMAT misc (196 RMAT 3	les a fea ion: fea ion: _ion:	scapu (685) : n i: ture .(196) : n i:	lari s a, 2) s a,	c,	g, or g, or	t				
aatt	tato	ict o	actta	aaaa	na to	ccacc	aaaa	acc	tocaa	ICC	acaaaaa	agt	tcatcatccg	60
qaqq	taad	rcc f	tatco	cago	na to	aactt	acac	aac	attaa	iaq	cgaacac	acq	attgggatcc	120
qctq	cata	igt d	ccaaa	.cqqa	ia qa	atcqc	ccaa	aco	acqqa	ıqa	tgtatgc	ctt	tgtcggtcac	180
acaa	gcac	icd 9	actcc	ccca	ia ca	aaqqq	caca	cca	aatca	lac	ccttcac	cct	tcqqtqaaqt	240
gcca	looga	iaa 1	ttaac	ttca	ig a	ggaga	tagc	ggc	caaca	itt	cgggagg	aaa	tgcgacgtct	300
gcag	cggo	:gc a	aagca	gcto	t go	cttct	cgtc	tcc	cctgg	ıag	tcgggct	ccc	cgtcggcgac	360
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Gln Lys Met Asp Pro Le 50	eu Thr Asn Leu Gln Val 55	Ala Ile Lys Asn Asn 60
Val Asp Val Phe Tyr P 65 70	he Ser Cys Leu Val Pro 0 75	Met His Val Leu Ser 80
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What is claimed is:

⁴⁵ 1. An isolated cDNA molecule which encodes an *Ixodes* ⁴⁵ associated antigenic polypeptide, said molecule having a nucleoxide sequence comprising at least residues 80-575 of SEQ ID NO: 3.

2. An expression vector comprising the isolated cDNA molecule of claim 1. 50

3. An isolated cell transformed by the expression vector of claim **2**.

4. The isolated cDNA molecule of claim **1**, wherein said cDNA molecule encodes a polypeptide represented by SEQ ID NO: 4.

5. The isolated cDNA molecule of claim **1**, wherein said cDNA molecule encodes a polypeptide that induces antibodies specific for an amino acid sequence represented by SEQ ID NO: 4.

* * * * *