

**COMPARTMENT-BASED WETLAND ECOSYSTEM
CONTAMINANT TRANSFER MODEL FOR
ASSESSING ECOSYSTEM RISK**

By

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Submitted to the Faculty of the
Graduate College of
Oklahoma State University
in partial fulfillment of
the requirements for
the Degree of
MASTER OF SCIENCE
May, 2000

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ACKNOWLEDGEMENTS

I first wish to give thanks and glory to my Lord and Savior Jesus Christ, without whom my life and any personal accomplishments would be meaningless and hollow. I also thank my major advisor, Dr. William McTernan, for his guidance, support, encouragement, and unwavering belief in my abilities. Dr. McTernan's constant challenges resulted in me a level of intellectual and professional maturation which I did not expect to gain from graduate school.

I thank my family for their support. I could not have realized many of my ambitions if not for the love, support, and personal sacrifices of my parents. My wife, Lisa, also deserves a sincere thank you. Lisa's complete selfless love has been one of the few constants in my life over the past several years. To the countless other family members and friends who have provided their support and encouragement, I thank them all.

Lastly, I would like to thank my employer, Fibercast Company, who provided much of the financial support for my graduate education.

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that many environmental contaminants may exist in several different forms once released into the environment, either through natural attenuation or by reacting with other substances already present. This only serves to increase the list of

1. INTRODUCTION

Degradation of the natural environment due to human activity has been well documented in American literature by the likes of Aldo Leopold in *Sand County Almanac* (1949) and Rachel Carson's *Silent Spring* (1962). However, it was not until the 1970's when widespread human health was imminently threatened that the U.S. Environmental Protection Agency (EPA) was created by an Executive Order. A series of Federal environmental regulations that became increasingly popular with the American public were subsequently promulgated. The EPA, along with its state analogues and other governmental agencies, is primarily charged with determining to what extent environmental releases of contaminants will result in a minimal negative impact on the general population. In other words, "What level of risk is acceptable to society?" This is not a simple task.

More often than not, it seems that determining what is an acceptable level of risk is based not so much on science, but rather on political agendas, industrial propaganda, and grass roots ideologies. Unfortunately, groups on all sides of an environmental debate can often produce "scientific evidence" to support their particular views. The reason for this is largely due to the complexity of the human physiology coupled with the myriad of chemicals which must be regulated. Adding to the dilemma is a scientific lack of understanding surrounding many of these chemicals. Even more confounding to the problem is

that many environmental contaminants may exist in several different forms once released into the environment, either through natural attenuation or by reacting with elements already present. This only serves to increase the level of uncertainty in the regulatory arena of human health and associated decision-making processes.

The subject of this work is ecological risk assessment. For all of the uncertainties that may exist in determining an acceptable level of risk to allow in human health assessments, broadening the application to the complexity of an ecosystem-level analysis greatly widens the chasm between what is known and what must be derived by mathematical interpolation (i.e., modeling).

Environmental contamination may adversely affect human health and/or ecosystem viability. As environmental contamination often results from routine economic activities, management approaches are needed to define contaminant release levels that afford protection to humans and the receiving ecosystems. Often these levels can be adequately described and attained by a specific numeric standard. More frequently however, complexities associated with either propagation of disease in humans or in environmental degradation within ecosystems serves to minimize the effectiveness of a standard. Risk assessment approaches have been developed to address more complex situations.

In a broad classification, risk assessments have traditionally focused on human health considerations (*environmental risk assessment*) or ecosystem concerns (*ecological risk assessment*). Ecological risk assessment requires an

effective method of assigning and attaining regulatory and societal goals at a reasonable cost and within an acceptable timeframe. Due to the massive variability and complexity inherent in ecosystems, this is only possible through a logical breakdown of responsibilities performed by a multidisciplinary team.

The term *ecological risk assessment* is defined below. Several simulation models will be reviewed as well as some of the techniques which attempt to deal with the uncertainty surrounding model inputs and outputs. The focus here is the application of a risk assessment model to a wetland ecosystem. The term wetland will be defined and the importance of wetland ecosystems will be discussed. Lastly, recent status and trends of wetlands in North America will be described.

1.1 Ecological Risk Assessment - Defined

An ecological risk assessment can be defined as the process of scientifically evaluating the probability (or likelihood) of compromising ecological integrity due to exposure to stressors (or contaminants) related to human activities (Norton *et al.* 1992). Furthermore, the assessment process should also give some measure of the magnitude of the compromising effects (Matlock *et al.* 1994). Norton *et al.* (1992) defined stressors as any chemical, physical, or biological entity that can elicit harmful ecological effects. Thus, the ecological risk assessment process must be broad enough to encompass the variety found in any given ecosystem. Due to the inherent complexities and lack of understanding of many ecological processes, no ecological risk assessment will

ever be perfect or totally complete (Cairns and McCormick 1992). However, there are guidelines in place that, if followed, should give as complete an assessment as possible.

The purpose of this work is not to provide a treatise on ecological risk assessment, but rather to develop and describe a mathematical model which may be used as a tool by risk assessors along with a discussion of the methods for understanding and managing inherent uncertainties. Modeling ecological risk is only part of the overall risk assessment process. Several protocols have been developed for completing an ecological risk assessment. The particular protocol chosen may depend upon several factors including: (1) statutory requirements, (2) site-, contaminant-, or species-specific applicability, (3) time limits/requirements, and (4) ultimate goals. Norton *et al.* (1992), Cairns and McCormick (1992), and Hope (1995), provide in-depth discussions of three ecological risk paradigms. A brief description of these paradigms is provided below. Although not intended to be inclusive of the whole of ecological risk assessment philosophies, these three methodologies illustrate the variety of the field.

The EPA's ecological risk assessment paradigm, as described by Norton *et al.* (1992), is a *tri-phase* process consisting of (1) Problem Formulation, (2) Analysis, and (3) Risk Characterization. In the formulation phase, the specific stressor of concern is identified and a conceptual model created or chosen to explain the ecological interactions. The analysis phase uses the site-specific or

site-appropriate model to evaluate collected information and stressor-related ecological impacts. Risk characterization estimates the potential ecological risks and evaluates potential remedial technologies.

Hope (1995) described an ecological risk paradigm similar to that of the EPA's but broke the process down into seven phases in an attempt to better define goals, optimize time, increase overall quality, and more efficiently allocate costs. Each of Hope's (1995) phases, which he called *work breakdown structures* (WBS's), were further divided into several sub-phases. Hope (1995) stated that it is extremely important to maintain an orderly and structured approach to ecological risk assessment in order to keep projects within budget and on schedule.

Cairns and McCormick (1992) offer a slightly different philosophy towards ecological risk assessment than those described by Norton *et al.* (1992) and Hope (1995). Rather than emphasizing laboratory experimentation and conceptual modeling, Cairns and McCormick (1992) suggest ongoing biological monitoring of carefully selected indicators of ecological integrity. As Cairns and McCormick (1992) point out, since "everything is an indicator of something, but nothing is an indicator of everything" selection of the appropriate indicator species is paramount to the successful discharge of their protocol.

1.2 Modeling Ecological Risk

Both Norton *et al.* (1992) and Hope (1995) state the need for a conceptual model that relates stressor-receptor interactions. Even Cairns and McCormick

(1992), who warn against the shortfalls and many uncertainties associated with describing complex ecological processes in the relatively simple format of simulation models, concede their importance. Ideally, ecological risk assessment models should be generic enough to predict the likelihood of many different types of stressors causing adverse effects in various regions to many different types of species (Hanratty and Stay 1994). These models aid the assessment team by estimating stressor exposure to receptors and stressor concentrations in particular media (MacIntosh *et al.* 1993). The models available to risk assessors range from simple bioconcentration models to much more complex food web models which require numerous inputs and extensive laboratory and field investigations. The key is understanding that simulation models are merely tools and that the risk assessor is charged with identifying the correct tool needed to achieve identified goals and objectives.

Aquatic ecological risk assessments have historically relied on steady-state and equilibrium partitioning models (Landrum *et al.* 1992). However, as Landrum *et al.* (1992) argue, simulation of non-steady-state and non-equilibrium phenomena in proper temporal and spatial context requires the use of kinetic models. Kinetic models are better able to simulate stressor concentrations in receptor tissues due to absorption, distribution, metabolism, and elimination (Landrum *et al.* 1992). Landrum *et al.* (1992) and MacIntosh *et al.* (1993) provide an overview of many of the previously mentioned models. A few of those are examined here.

1.2.1 Steady-State Models

Receptors are said to be at steady-state when stressor exposure and uptake and loss factors remain unchanged. This equilibrium state may be assumed for resident receptors with relatively long life-cycles (MacIntosh *et al.* 1993). For aquatic stressor exposures under these conditions, a tissue bioconcentration factor (BCF) in milliliters per gram can be expressed as:

$$BCF = \frac{C_a}{C_w} \quad (1)$$

where:

C_a = stressor concentration in the receptor ($\mu\text{g/g}$)

C_w = stressor concentration in the water ($\mu\text{g/mL}$)

When assessing risk to benthic receptors the above equation can be modified to focus on stressor uptake from sediment or food source to yield a bioaccumulation factor (BAF) in grams of sediment or food source per gram of tissue:

$$BAF = \frac{C_a}{C_s} \quad (2)$$

where:

C_s = stressor concentration in the sediment or food source ($\mu\text{g/g}$)

The last steady-state model given by Landrum *et al.* (1992) again describes stressor uptake from sediment or food source. However, it also takes into account the lipid content in the stressor and organic carbon content in the

foodstuff. This accumulation factor (AF) in grams of organic carbon per grams of lipid is:

$$AF = \frac{C_a(l)}{C_s(c)} \quad (3)$$

where:

$C_a(l)$ = receptor stressor concentration per gram lipid ($\mu\text{g/g}$)

$C_s(c)$ = source stressor concentration per gram organic carbon ($\mu\text{g/g}$)

1.2.2 Kinetic Models

Landrum *et al.* (1992) state that kinetic models can be placed into one of two categories: (1) compartment-based or (2) physiology-based. Compartment-based models describe stressor movement between the biotic components, or compartments (e.g., species, population, trophic level), and the abiotic components (i.e., water, soil, sediment) of an ecosystem system. Physiology-based pharmacokinetic (PBPK) models simulate stressor accumulation and distribution within receptor tissues.

1.2.3 Compartment Models

Landrum *et al.* (1992) detail many of the assumptions associated with compartment models: (1) homogeneous mixture of stressor within each compartment, (2) no stressor biotransformation, (3) rate constants do not change over time, and (4) first-order transfer between compartments.

Using first-order transfer functions, with the stressor concentration in the water compartment (C_w) held static, Landrum *et al.* (1992) provide the following

model for calculation of the stressor concentration in the receptor (C_a):

$$C_a = \left(\frac{k_u \cdot C_w}{k_e} \right) (1 - e^{-k_e t}) \quad (4)$$

where:

k_u = conditional uptake clearance (mg/g/hour)

k_e = conditional elimination rate constant (hour⁻¹)

t = time (hours)

The term clearance is defined as the mass of a compartment relieved of stressor per mass of receptor per time (Landrum *et al.* 1992).

Growth compensation can be applied to the above model by adding a first-order growth rate constant in grams per gram per hour (g) to the elimination rate constant (k_e):

$$C_a = \left(\frac{k_u \cdot C_w}{k_e + g} \right) (1 - e^{-(k_e + g)t}) \quad (5)$$

Decay could similarly be incorporated into the model by applying the term as a negative value.

MacIntosh *et al.* (1993) described a compartment-based model similar to equations 3 and 4 which gave a bioaccumulation factor for the stressor from contaminated food:

$$BAF = (BCF + BMF)(1 - e^{-k_e t}) \quad (6)$$

where:

BCF = bioconcentration factor: stressor partitioning between compartments

(mL/g)

BMF = biomagnification factor: stressor uptake from contaminated food
(unitless)

k_{el} = conditional elimination rate constant (day^{-1})

t = average receptor life span (days)

To calculate stressor uptake (BMF) from food and drinking water, MacIntosh *et al.* (1993) employed the following equation:

$$\text{BMF} = M \cdot \sum f \cdot \text{BAF} + I_w \cdot A_{\text{eff}} \cdot k_{\text{el}}^{-1} \quad (7)$$

with,

$$M = I_p \cdot A_{\text{eff}} \cdot k_{\text{el}}^{-1} \quad (8)$$

where:

M = magnification term (unitless)

f = fraction of predator diet consisting of prey (unitless)

I_p = amount of food ingested (g/g/day)

I_w = amount of water ingested (g/g/day)

BAF = bioaccumulation factor for each prey (unitless)

A_{eff} = fraction of ingested stressor that is absorbed across the gut lining of the receptor (unitless)

In small static systems stressor concentration in the water compartment decreases as it is accumulated in the receptor. By incorporating the mass balance of stressor in each compartment, Landrum *et al.* (1992) state that the concentration of the stressor in the receptor (Q_a) can be expressed as:

$$Q_a = \frac{(k_{um} \cdot A)(1 - e^{-(k_{um} + k_e)t})}{k_{um} + k_e} \quad (9)$$

where:

A = stressor in the system ($Q_w + Q_a$) in μg

k_{um} = uptake rate constant (hour^{-1})

1.2.4 Physiology-Based Pharmacokinetic Models

Physiology-based pharmacokinetic (PBPK) models compartmentalize the receptor into related tissue types (Landrum *et al.* 1992). PBPK models are attractive due to the fact that they can be used on practically any species given the availability of that species' physiological information. However, these models also require much more prior information for proper implementation than do the aforementioned compartment models (Landrum *et al.* 1992).

1.3 Model Selection

The assessor should take great care in choosing the proper model which will be used to describe a study site. If the initial phases of the risk assessment are performed properly (i.e., within a logical framework and with clear goals in mind), choosing the correct simulation model will be a much easier task. Once the proper model or suite of models is chosen, the risk assessor must then make a decision on how to deal with the uncertainty associated with his choice.

1.4 Uncertainty Analysis

Simulation models used in ecological risk assessment are often plagued with a large amount of uncertainty surrounding their input and output values. These

uncertainties can come from a variety of sources, such as: limited understanding of ecological processes, limited access or ability to measure actual ranges of model inputs and parameters, sampling error, ecological variability, and mathematical oversimplification of ecological processes (Dakins *et al.* 1995). If ignored, these uncertainties can compromise the utility of a model and thus lead to poor or improper decision making. A widely used method to combat the uncertainty found in parameter selection for simulation modeling is Monte Carlo analysis.

1.4.1 Monte Carlo Analysis

Monte Carlo analysis samples statistical frequency distributions (a.k.a. probability distribution functions) for uncertain model parameters. These probability distribution functions (pdfs) are calculated from value ranges obtained from published or field data. Simulations are run numerous times, randomly accessing values for each iteration from the user-defined distributions (Dilks *et al.* 1992). Monte Carlo analysis is attractive to modelers due to its applicability to many different types of models as well as to many types of frequency distributions (i.e., normal, log-normal, triangular, etc.) (Slob 1994). Figure 1 clearly shows the difference between a typical deterministic (point) estimate and a Monte Carlo (probabilistic or stochastic) estimate using distribution functions rather than single average values for each input parameter.

The point estimate shown at the top of Figure 1 depicts four values (A, B, C, and D) which are used to calculate the result (E). In a traditional analysis, the

four values would likely represent averages of several actual field or laboratory measurements. The result (E) then represents a rather sterile average answer which does not enlighten the researcher to the variability common in natural systems. The graphic in the lower half of Figure 1 depicts the same four input variables, however, rather than only using single average values, the Monte Carlo approach utilizes density functions developed from measurements. In this way, the analyst can define probabilities of occurrence for the outputs.

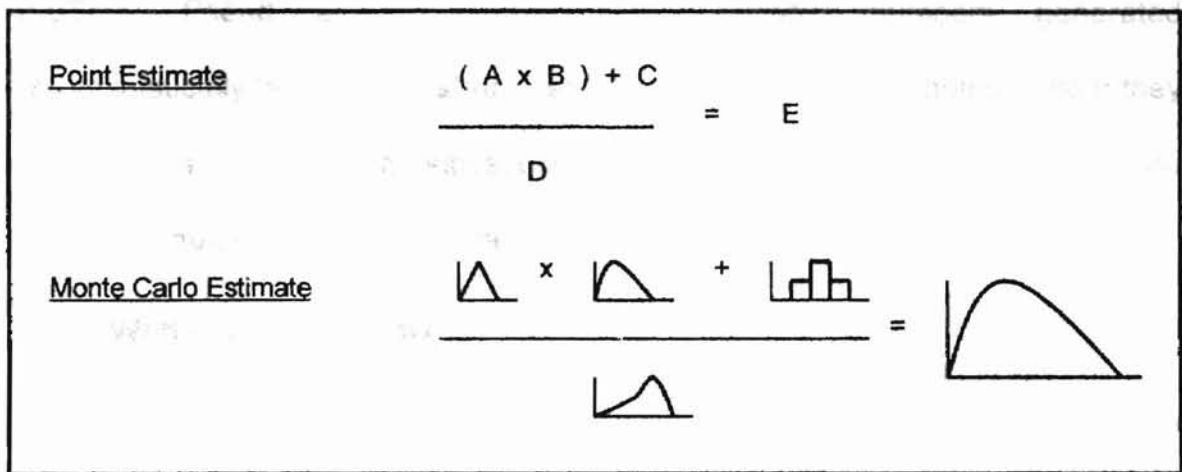


Figure 1. Comparison of a point estimate and a Monte Carlo estimate.

1.4.2 Latin Hypercube

Latin Hypercube is a stratification of the pdfs which "allows the output distribution to be characterized with a smaller number of replications than simple random sampling" (Dakins *et al.* 1995, pg. 69). Once the pdf is stratified, or separated into equal intervals, samples are randomly taken from each interval (Palisade 1996). Each stratification is sampled only once during an analysis. In this manner, Latin Hypercube analysis reportedly more accurately represents the

input pdf with fewer iterations than with a traditional Monte Carlo analysis (Palisade 1996).

1.4.3 Statistical Quality of Monte Carlo Analysis

The statistical quality of a Monte Carlo analysis is dependent upon the quality of the uniform random number generator employed by the particular software used. Unbeknownst to many analysts is the fact that computers use “mechanical, wholly deterministic methods” to generate random numbers (Barry 1995). Pseudo-random numbers (i.e., random numbers generated deterministically by computers) are defined by Barry (1995) as being good if they exhibit statistical uniformity, statistical independence, reproducibility, and can be generated quickly and economically.

1.5 Wetland Ecosystems

Wetlands are ecosystems where saturation with water is the dominant factor determining the types of plant and animal communities living in the soil or on its surface (i.e., habitat which is transitional between terrestrial and aquatic systems in which the water table usually lies at or near the surface.) (Cowardian *et al.* 1979). Wetlands provide critical feeding, resting, and breeding habitat for many waterfowl, shorebirds, and Neotropical migrants as well as many other animals (Hobbs and Barksdale 1993). Wetlands are also very important to humans by reducing flooding problems through temporary storage of flood waters. Wetlands aid water quality by filtering out pollutants and sediments, and also provide recreation (e.g., hunting, bird watching) (USDI 1988).

1.5.1 Status of Wetlands in North America

Of the approximately 87 million-ha of wetlands that existed in North America at the time of the continent's settlement, only about 41 million-ha remained in the 1980's (Frayer 1991). These figures equate to an approximate 53% net loss of the nation's wetlands.

patterns of population growth: (1) exponential growth, typical of small populations that are increasing rapidly in an under-utilized habitat; (2) logarithmic growth, typical of relatively larger populations in which growth is slowed due to

2. METHODOLOGY

The model chosen for this wetland ecosystem risk assessment is an adaptation of the classic *carrying capacity* equation which has seen widespread use among wildlife management professionals. This model was chosen because it incorporates many of the characteristics of the equations previously described from Landrum *et al.* (1992) and MacIntosh *et al.* (1993) such as first-order kinetics and steady-state equilibrium partitioning. Further, the model lends itself to a compartment-based application. The basic carrying capacity equation, as taken from Anderson (1991), is as follows:

$$\frac{dN}{dt} = rN \left(\frac{K-N}{K} \right) \quad (10)$$

where:

- r = constant, *intrinsic rate of increase*, equal to the birth rate minus the death rate
- N = number of individuals in the population at a given moment
- K = *carrying capacity*, the maximum number of individuals a habitat can support
- t = time

The carrying capacity model provides wildlife professionals with a useful tool for managing animals in a population context. The equation simulates three basic

phenomena of population growth: (1) exponential growth, typical of small populations increasing rapidly in an under-utilized habitat; (2) logarithmic growth, typical of relatively larger populations in which growth is slowed due to decreasing availability of resources; and (3) population stabilization, in which the population growth is in equilibrium with the environment (see Figure 2).

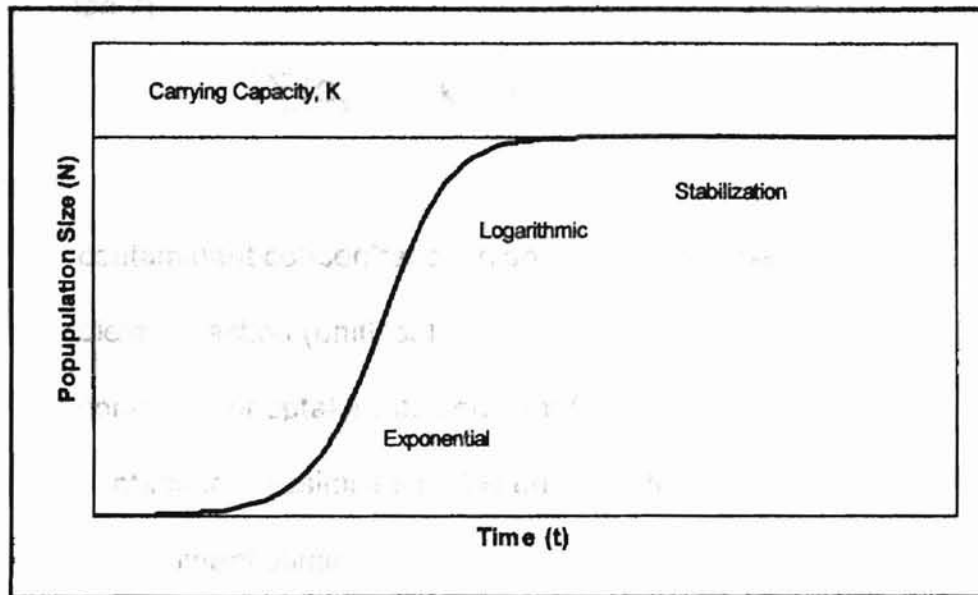


Figure 2. Visual representation of the carrying capacity equation.

2.1 Compartment Model for Assessing Ecological Risk in a Wetland

The model utilized for this work has the same basic form as the carrying capacity equation.

$$\frac{dC}{dt} = rC \left(\frac{C_e - C}{C_e} \right) \quad (11)$$

where:

r = net contaminant uptake rate (time^{-1})

C = compartmental contaminant concentration (mg/kg or L) (14)

C_e = endpoint contaminant concentration (mg/kg)

t = time

Further, net contaminant uptake rate, r , is represented by the following expression and is similar to the BMF term employed by MacIntosh *et al.* (1993) (see equation 7):

$$\left[\sum (C_s \cdot df) \cdot (k_u \cdot \text{eff}) \right] - k_e \quad (12)$$

where:

C_s = contaminant concentration in source compartment (mg/kg or L)

df = dietary fraction (unitless)

k_u = contaminant uptake rate constant (kg or L/mg/time)

eff = contaminant assimilation efficiency (unitless)

k_e = contaminant elimination rate constant (time^{-1})

Therefore, the compartmental contaminant concentration can be calculated for C_t as follows:

$$C_t = \left(\left[\left(\sum (C_s \cdot df) \cdot (k_u \cdot \text{eff}) \right) - k_e \right] \cdot C_{t-1} \right) \cdot \left[\frac{C_e - C_{t-1}}{C_e} \right] + C_{t-1} \quad (13)$$

The model above provides an estimate of risk, the *Risk Factor* (RF), based on the ratio of the contaminant concentration within a compartment and the user-defined endpoint concentration (C_e) for that compartment. The risk present at a specific time interval or at the maximum contaminant concentration level for a compartment is calculated as follows:

$$RF_t = \frac{C_t}{C_e} \quad (14)$$

and

$$RF_{max} = \frac{C_{max}}{C_e} \quad (15)$$

where:

RF_t = risk factor at time t (unitless)

RF_{max} = risk factor at maximum contaminant concentration level (unitless)

C_t = contaminant concentration at time t (mg/kg)

C_{max} = maximum contaminant concentration (mg/kg)

C_e = endpoint contaminant concentration (mg/kg)

The risk factor is expressed as a unitless number between zero and one. The greater the RF, the greater the risk.

2.2 Explanation of Model Terms

The model requires the user to input several variables. Based on these user-defined inputs, the model will calculate output data. The nomenclature used within the model to identify these variables is described below.

Contaminant Concentration (C). Represents the stressor concentration within a compartment of interest at a particular time. Dependent upon the compartment, the contaminant concentration contains units of mg/kg or mg/L. This term can be expressed in several different forms: C_o , initial contaminant concentration; C_t , contaminant concentration at a specific time interval; C_{max} , maximum contaminant concentration over a user-defined time period; C_e ,

endpoint concentration; C_s , contaminant concentration in the source compartment. This is the fraction of the

Endpoint Concentration (C_e). Mathematically, this term sets the asymptotic maximum value which the model will return for a particular compartmental contaminant concentration. For risk assessment purposes, this term is used to identify the concentration at which individuals or components within a compartment of interest may be expected to show signs of overexposure (e.g., infertility, birth defects, mutations, death, etc.). C_e may also be used to represent the concentration at which humans may exhibit ill-effects via exposure to the individuals or components within the contaminated compartments.

Contaminant Concentration in the Source Compartment (C_s). In order for the model to calculate a compartmental contaminant concentration, all source compartments must be identified and "loaded" with an initial contaminant concentration (C_0).

Dietary Fraction (df). Once all relevant inter-compartmental relationships were identified, the fraction of each source compartment utilized by the receiving compartment was estimated. For example, a herbivorous invertebrate compartment could be identified to potentially receive contaminant from terrestrial plants, soil, and water compartments. In this scenario, the model would require the user to define what fraction of each of these compartments the herbivorous invertebrates utilize in their diet. These unitless values must be input as decimals and their sum should equal 1.00.

Contaminant Uptake Rate Constant (k_u). This is the gross contaminant

uptake rate for a compartment expressed as either kg/mg/time or L/mg/time, and

Contaminant Assimilation Efficiency (eff). This is the fraction of the contaminant which is actually absorbed into the tissues of the individuals comprising a compartment.

Contaminant Elimination Rate Constant (k_e). This is the rate per time at which contaminant is purged from the tissues of the individuals comprising a compartment.

Net Contaminant Uptake Rate (r). Calculated from equation 12, the net contaminant uptake rate represents the relative per time increase, or decrease, of contaminant which persists within each compartment.

Risk Factor (RF). The amount of risk present within each compartment is calculated from equations 14 and 15. RF_i (equation 14) allows the user to estimate the incremental risk present within a given compartment at any point between C_o and C_{max} . RF_{max} (equation 15), the maximum risk factor, calculates the amount of risk present within a compartment at the maximum contaminant concentration encountered over the user-defined time interval. The risk factor is a unitless value between 0 and 1.00, with risk increasing as the RF approaches unity.

2.3 Generalized Wetland Ecosystem Food Web Matrix

The previously described model was applied to a wetland food web matrix consisting of fifteen compartments, twelve biotic and three abiotic. Each of the biotic compartments represents an entire trophic level (e.g., top predator, waterfowl, aquatic plants) rather than populations (e.g., Canada geese, river

elms) or individuals. The three abiotic compartments include soil, sediment, and water. The contaminant transfer relationships between the fifteen compartments are provided below as Figure 3.

- | | |
|--|---|
| 1. Top Carnivore (TC) | 7. Terrestrial Plants (TP) |
| 2. Waterfowl (WF) | 8. Carnivorous Fish (CF) |
| 3. Carnivorous Mammals (CM) | 9. Herbivorous Fish (HF) |
| 4. Herbivorous Mammals (HM) | 10. Aquatic Carnivorous Invertebrates (ACI) |
| 5. Terrestrial Carnivorous Invertebrates (TCI) | 11. Aquatic Herbivorous Invertebrates (AHI) |
| 6. Terrestrial Herbivorous Invertebrates (THI) | 12. Aquatic Plants (AP) |

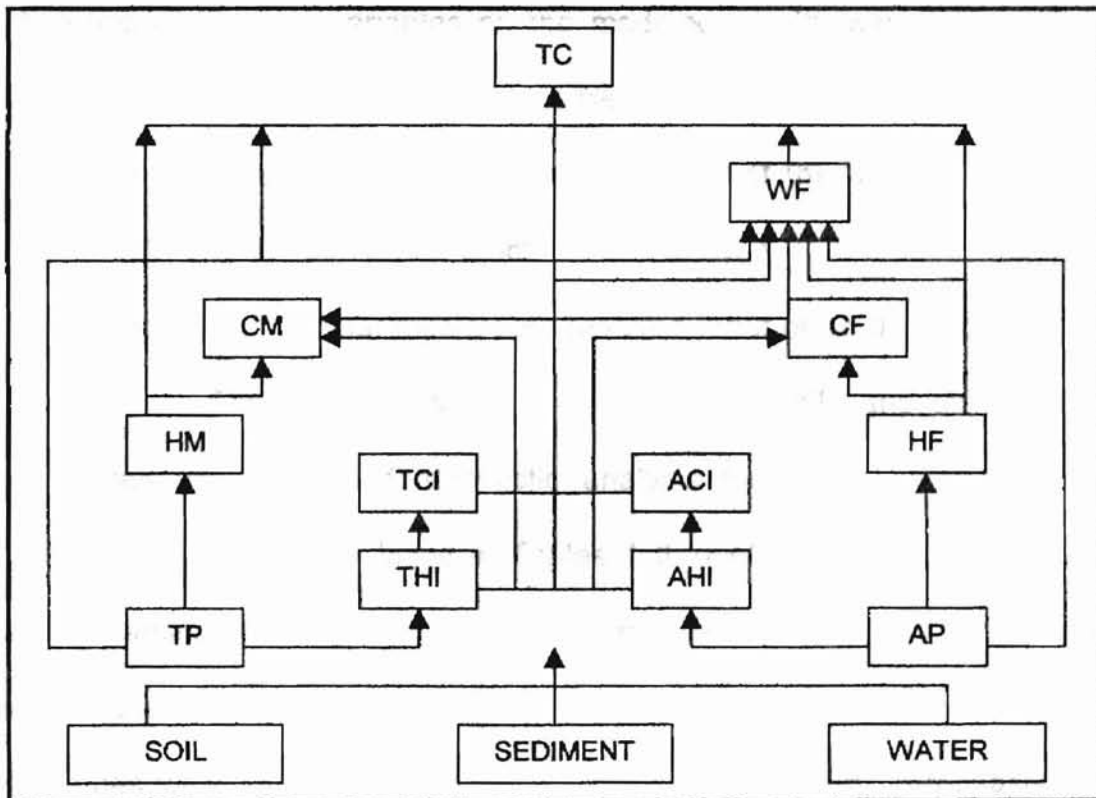


Figure 3. Generalized wetland ecosystem food web matrix.

2.4 Model Input Data

Initial stressor concentration values were adapted from selenium data from Kesterson Reservoir, California (CH2M Hill 1986 and USBR 1990). Site-specific and contaminant-specific data for the remaining input variables (i.e., endpoint concentrations, uptake and elimination rates, assimilation efficiencies, and dietary fractions) were not included in the original reports. Therefore, statistical distributions describing uptake and elimination rates for contaminants other than selenium were adapted from Stehly *et al.* (1990), similar to the approaches of Connolly and Tonelli (1985) and MacIntosh *et al.* (1993). The remaining input variables were estimated from available sources. Subsequently, a sensitivity analysis was performed to determine the applicability of this effort where the relative impact of input variables on the modeled results was evaluated. As discussed later, these uncertain variables were shown to have a negligible effect on the final model output in the sensitivity portion of the stochastic analysis, thereby reinforcing the utility of the technique employed.

The three abiotic compartments only required input of an initial contaminant concentration. The model was run both deterministically and stochastically. Input values used in the deterministic analysis for each compartment and compartment variables are listed in Tables 1 through 14. Table 1 shows the initial contaminant concentration, C_0 , deterministic input values for variables common to all biotic compartments. Tables 3 through 14 show the deterministic input values for variables specific to each biotic compartment. Stochastic input value ranges are provided in Tables 15 through 31.

Table 1. Deterministic Input Values for Variables
Specific to Top Carnivore Compartment

**Table 1. Initial Se Contaminant Concentration, C_0 , Deterministic
Input Values for Abiotic Compartments**

COMPARTMENT NAME	UNITS	INPUT VALUE ¹
Soil	mg/kg	7.00
Sediment	mg/kg	7.00
Water	mg/L	0.23

¹ CH2M Hill (1986)

**Table 2. Deterministic Input Values for Variables
Common to all Biotic Compartments**

COMPARTMENT NAME	INPUT VALUE				
	C_0	C_0 ⁴	k_u ³	k_e ³	eff ⁴
	mg/kg	mg/kg	(kg or L) / mg/time	time ⁻¹	unitless
Top Carnivore, TC	9.40 ¹	100.00	0.04	0.02	0.40
Carnivorous Mammals, CM	9.40 ¹	100.00	0.04	0.02	0.35
Herbivorous Mammals, HM	9.40 ¹	100.00	0.05	0.02	0.30
Terrestrial Carnivorous Invertebrates, TCI	10.20 ²	100.00	0.02	0.01	0.20
Terrestrial Herbivorous Invertebrates, THI	7.80 ²	100.00	0.05	0.02	0.20
Terrestrial Plants, TP	5.00 ¹	100.00	0.08	0.02	0.40
Waterfowl, WF	4.80 ¹	100.00	0.04	0.02	0.35
Carnivorous Fish, CF	135.00 ¹	250.00	0.04	0.02	0.30
Herbivorous Fish, HF	135.00 ¹	250.00	0.08	0.02	0.30
Aquatic Carnivorous Invertebrates, ACI	114.50 ²	250.00	0.04	0.02	0.60
Aquatic Herbivorous Invertebrates, AHI	114.50 ²	350.00	0.08	0.02	0.20
Aquatic Plants, AP	5.00 ¹	100.00	0.10	0.02	0.40

¹ CH2M Hill (1986)

² USBR (1990)

³ Adapted from Stehly (1990)

⁴ Estimated

Table 3. Deterministic Input Values for Variables Specific to Top Carnivore Compartment

TOP CARNIVORE SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.10
Sediment	0.05
Water	0.05
Carnivorous Mammals, CM	0.05
Herbivorous Mammals, HM	.030
Terrestrial Carnivorous Invertebrates, TCI	0.10
Terrestrial Herbivorous Invertebrates, THI	0.10
Waterfowl, WF	0.05
Carnivorous Fish, CF	0.05
Herbivorous Fish, HF	0.05
Aquatic Carnivorous Invertebrates, ACI	0.05
Aquatic Herbivorous Invertebrates, AHI	0.05

¹ Estimated

Table 4. Deterministic Input Values for Variables Specific to Carnivorous Mammals Compartment

CARNIVOROUS MAMMALS SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.05
Sediment	0.05
Water	0.10
Herbivorous Mammals, HM	0.35
Terrestrial Carnivorous Invertebrates, TCI	0.10
Terrestrial Herbivorous Invertebrates, THI	0.10
Waterfowl, WF	0.05
Carnivorous Fish, CF	0.05
Herbivorous Fish, HF	0.05
Aquatic Carnivorous Invertebrates, ACI	0.05
Aquatic Herbivorous Invertebrates, AHI	0.05

¹ Estimated

Table 5. Deterministic Input Values for Variables Specific to Terrestrial Plants Compartment

Table 5. Deterministic Input Values for Variables Specific to Herbivorous Mammals Compartment

HERBIVOROUS MAMMALS SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.05
Sediment	0.05
Water	0.10
Terrestrial Plants, TP	0.70
Aquatic Plants, AP	0.10

¹ Estimated

Table 6. Deterministic Input Values for Variables Specific to Terrestrial Carnivorous Invertebrates Compartment

TERRESTRIAL CARNIVOROUS INVERTEBRATES SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.20
Sediment	0.05
Water	0.05
Terrestrial Herbivorous Invertebrates, THI	0.60
Aquatic Carnivorous Invertebrates, ACI	0.05
Aquatic Herbivorous Invertebrates, AHI	0.05

¹ Estimated

Table 7. Deterministic Input Values for Variables Specific to Terrestrial Herbivorous Invertebrates Compartment

TERRESTRIAL HERBIVOROUS INVERTEBRATES SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.10
Sediment	0.10
Water	0.05
Terrestrial Plants, TP	0.70
Aquatic Plants, AP	0.05

¹ Estimated

Table 8. Deterministic Input Values for Variables Specific to Terrestrial Plants Compartment

TERRESTRIAL PLANTS SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.75
Sediment	0.05
Water	0.20

¹ Estimated

Table 9. Deterministic Input Values for Variables Specific to Waterfowl Compartment

WATERFOWL SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.05
Sediment	0.05
Water	0.20
Terrestrial Carnivorous Invertebrates, TCI	0.05
Terrestrial Herbivorous Invertebrates, THI	0.05
Terrestrial Plants, TP	0.10
Carnivorous Fish, CF	0.10
Herbivorous Fish, HF	0.10
Aquatic Carnivorous Invertebrates, ACI	0.10
Aquatic Herbivorous Invertebrates, AHI	0.10
Aquatic Plants, AP	0.10

¹ Estimated

Table 10. Deterministic Input Values for Variables Specific to Carnivorous Fish Compartment

CARNIVOROUS FISH SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.05
Sediment	0.05
Water	0.20
Terrestrial Carnivorous Invertebrates, TCI	0.05
Terrestrial Herbivorous Invertebrates, THI	0.05
Herbivorous Fish, HF	0.50
Aquatic Carnivorous Invertebrates, ACI	0.05
Aquatic Herbivorous Invertebrates, AHI	0.05

¹ Estimated

Table 14. Deterministic Input Values for Variables Specific to Aquatic Plants Compartment

Table 11. Deterministic Input Values for Variables Specific to Herbivorous Fish Compartment

HERBIVOROUS FISH SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.05
Sediment	0.05
Water	0.20
Aquatic Plants, AP	0.70

¹ Estimated

Table 12. Deterministic Input Values for Variables Specific to Aquatic Carnivorous Invertebrates Compartment

AQUATIC CARNIVOROUS INVERTEBRATES SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.05
Sediment	0.05
Water	0.20
Terrestrial Carnivorous Invertebrates, TCI	0.05
Terrestrial Herbivorous Invertebrates, THI	0.05
Aquatic Herbivorous Invertebrates, AHI	0.60

¹ Estimated

Table 13. Deterministic Input Values for Variables Specific to Aquatic Herbivorous Invertebrates Compartment

AQUATIC HERBIVOROUS INVERTEBRATES SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.05
Sediment	0.05
Water	0.20
Aquatic Plants, AP	0.70

¹ Estimated

Table 14. Deterministic Input Values for Variables Specific to Aquatic Plants Compartment

AQUATIC PLANTS SOURCE COMPARTMENTS	INPUT VALUE ¹
	df
	Unitless
Soil	0.05
Sediment	0.75
Water	0.20

¹ Estimated

The stochastic input ranges for each model variable are provided below in Tables 15 through 31. The ranges shown in Table 15 for the initial contaminant concentration, C_0 , are again taken from published values for selenium contamination at Kesterson Reservoir, California (CH2M Hill 1986 and USBR 1990). All published contaminant concentration ranges, with the exception of the water selenium contaminant concentration, were reported as a minimum, maximum, and mean and therefore, a triangular distribution was assigned to each of these variables in the model. The water selenium contaminant concentration reported by CH2M Hill (1986) consisted of seventy-three data points. These data were analyzed with a probability distribution fitting software package and assigned a normal distribution. The results of the curve fitting analysis is provided as Appendix A.

Tables 16, 17, 18, and 19 list input ranges for endpoint contaminant concentrations, elimination rate constants, and assimilation efficiencies respectively for all biotic compartments. Tables 20 through 31 list the dietary fraction input ranges specific to each of the twelve biotic compartments. As previously mentioned, site- and contaminant-specific values for these variables

were not found in the literature. In order to randomize these variables in the stochastic analysis, triangular ranges were created for each value by setting the maximum to 1.5 times the deterministic values and the minimum to 0.5 times the deterministic values. The deterministic value was used as the mean.

**Table 15. Initial Se Contaminant Concentration, C_0 ,
Stochastic Input Values for All Compartments**

COMPARTMENT NAME	INPUT RANGE (mg / [kg or L])			DISTRIBUTION TYPE
	MIN	MAX	MEAN	
Soil ¹	1.000	85.000	7.000	Triangular
Sediment ¹	1.000	85.000	7.000	Triangular
Water ¹	standard deviation = 0.129		0.228	Normal
Top Carnivore, TC ¹	0.100	125.000	9.400	Triangular
Carnivorous Mammals, CM ¹	0.100	125.000	9.400	Triangular
Herbivorous Mammals, HM ¹	0.100	125.000	9.400	Triangular
Terrestrial Carnivorous Invertebrates, TCI ²	2.000	84.000	10.200	Triangular
Terrestrial Herbivorous Invertebrates, THI ²	0.400	42.000	7.800	Triangular
Terrestrial Plants, TP ¹	0.100	82.000	5.000	Triangular
Waterfowl, WF ¹	1.600	31.000	4.600	Triangular
Carnivorous Fish, CF ¹	2.000	200.000	135.000	Triangular
Herbivorous Fish, HF ¹	2.000	200.000	135.000	Triangular
Aquatic Carnivorous Invertebrates, ACI ²	20.000	218.000	114.500	Triangular
Aquatic Herbivorous Invertebrates, AHI ²	20.000	218.000	114.500	Triangular
Aquatic Plants, AP ¹	0.100	82.000	5.000	Triangular

¹ CH2M Hill (1986)

² USBR (1990)

Table 16. Endpoint Se Contaminant Concentration, C_e, Stochastic Input Values for Biotic Compartments

COMPARTMENT NAME	INPUT RANGE (mg/kg) ¹			DISTRIBUTION TYPE
	MIN	MAX	MEAN	
Top Carnivore, TC	50.000	150.000	100.000	Triangular
Carnivorous Mammals, CM	50.000	150.000	100.000	Triangular
Herbivorous Mammals, HM	50.000	150.000	100.000	Triangular
Terrestrial Carnivorous Invertebrates, TCI	50.000	150.000	100.000	Triangular
Terrestrial Herbivorous Invertebrates, THI	50.000	150.000	100.000	Triangular
Terrestrial Plants, TP	50.000	150.000	100.000	Triangular
Waterfowl, WF	50.000	150.000	100.000	Triangular
Carnivorous Fish, CF	125.000	375.000	250.000	Triangular
Herbivorous Fish, HF	125.000	375.000	250.000	Triangular
Aquatic Carnivorous Invertebrates, ACI	125.000	375.000	250.000	Triangular
Aquatic Herbivorous Invertebrates, ACI	125.000	375.000	250.000	Triangular
Aquatic Plants, AP	50.000	150.000	100.000	Triangular

¹ Estimated

Table 17. Se Uptake Rate Constants, k_u, Stochastic Input Values for Biotic Compartments

COMPARTMENT NAME	INPUT RANGE ([kg or L] / mg/time) ¹			DISTRIBUTION TYPE
	MIN	MAX	MEAN	
Top Carnivore, TC	0.020	0.060	0.040	Triangular
Carnivorous Mammals, CM	0.020	0.060	0.040	Triangular
Herbivorous Mammals, HM	0.025	0.075	0.050	Triangular
Terrestrial Carnivorous Invertebrates, TCI	0.010	0.030	0.020	Triangular
Terrestrial Herbivorous Invertebrates, THI	0.025	0.075	0.050	Triangular
Terrestrial Plants, TP	0.030	0.090	0.060	Triangular
Waterfowl, WF	0.005	0.015	0.010	Triangular
Carnivorous Fish, CF	0.020	0.060	0.040	Triangular
Herbivorous Fish, HF	0.040	0.120	0.080	Triangular
Aquatic Carnivorous Invertebrates, ACI	0.020	0.060	0.040	Triangular
Aquatic Herbivorous Invertebrates, ACI	0.040	0.120	0.080	Triangular
Aquatic Plants, AP	0.050	0.150	0.100	Triangular

¹ Adapted from Stehly (1990)

Table 18. Se Elimination Rate Constants, k_e , Stochastic Input Values for Biotic Compartments

COMPARTMENT NAME	INPUT RANGE (time ⁻¹) ¹			DISTRIBUTION TYPE
	MIN	MAX	MEAN	
Top Carnivore, TC	0.001	0.003	0.002	Triangular
Carnivorous Mammals, CM	0.010	0.030	0.020	Triangular
Herbivorous Mammals, HM	0.010	0.030	0.020	Triangular
Terrestrial Carnivorous Invertebrates, TCI	0.005	0.015	0.010	Triangular
Terrestrial Herbivorous Invertebrates, THI	0.010	0.030	0.020	Triangular
Terrestrial Plants, TP	0.010	0.030	0.020	Triangular
Waterfowl, WF	0.010	0.030	0.020	Triangular
Carnivorous Fish, CF	0.010	0.030	0.020	Triangular
Herbivorous Fish, HF	0.010	0.030	0.020	Triangular
Aquatic Carnivorous Invertebrates, ACI	0.010	0.030	0.020	Triangular
Aquatic Herbivorous Invertebrates, ACI	0.010	0.030	0.020	Triangular
Aquatic Plants, AP	0.010	0.030	0.020	Triangular

¹ Adapted from Stehly (1990)

Table 19. Se Contaminant Assimilation Efficiency, eff, Stochastic Input Values for Biotic Compartments

COMPARTMENT NAME	INPUT RANGE (unitless) ¹			DISTRIBUTION TYPE
	MIN	MAX	MEAN	
Top Carnivore, TC	0.200	0.600	0.400	Triangular
Carnivorous Mammals, CM	0.175	0.525	0.350	Triangular
Herbivorous Mammals, HM	0.100	0.300	0.200	Triangular
Terrestrial Carnivorous Invertebrates, TCI	0.100	0.300	0.200	Triangular
Terrestrial Herbivorous Invertebrates, THI	0.100	0.300	0.200	Triangular
Terrestrial Plants, TP	0.200	0.600	0.400	Triangular
Waterfowl, WF	0.035	0.105	0.070	Triangular
Carnivorous Fish, CF	0.100	0.300	0.200	Triangular
Herbivorous Fish, HF	0.100	0.300	0.200	Triangular
Aquatic Carnivorous Invertebrates, ACI	0.100	0.300	0.200	Triangular
Aquatic Herbivorous Invertebrates, ACI	0.100	0.300	0.200	Triangular
Aquatic Plants, AP	0.200	0.600	0.400	Triangular

¹ Estimated

Table 20. Stochastic Input Values for Variables Specific to Top Carnivore Compartment

TOP CARNIVORE SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.025	0.075	0.050	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.050	0.150	0.100	Triangular
Carnivorous Mammals, CM	0.025	0.075	0.050	Triangular
Herbivorous Mammals, HM	0.150	0.450	0.300	Triangular
Terrestrial Carnivorous Invertebrates, TCI	0.050	0.150	0.100	Triangular
Terrestrial Herbivorous Invertebrates, THI	0.050	0.150	0.100	Triangular
Waterfowl, WF	0.025	0.075	0.050	Triangular
Carnivorous Fish, CF	0.025	0.075	0.050	Triangular
Herbivorous Fish, HF	0.025	0.075	0.050	Triangular
Aquatic Carnivorous Invertebrates, ACI	0.025	0.075	0.050	Triangular
Aquatic Herbivorous Invertebrates, AHI	0.025	0.075	0.050	Triangular

¹ Estimated

Table 21. Stochastic Input Values for Variables Specific to Carnivorous Mammals Compartment

CARNIVOROUS MAMMALS SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.025	0.075	0.050	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.050	0.150	0.100	Triangular
Herbivorous Mammals, HM	0.175	0.525	0.350	Triangular
Terrestrial Carnivorous Invertebrates, TCI	0.050	0.150	0.100	Triangular
Terrestrial Herbivorous Invertebrates, THI	0.050	0.150	0.100	Triangular
Waterfowl, WF	0.025	0.075	0.050	Triangular
Carnivorous Fish, CF	0.025	0.075	0.050	Triangular
Herbivorous Fish, HF	0.025	0.075	0.050	Triangular
Aquatic Carnivorous Invertebrates, ACI	0.025	0.075	0.050	Triangular
Aquatic Herbivorous Invertebrates, AHI	0.025	0.075	0.050	Triangular

¹ Estimated

Table 22. Stochastic Input Values for Variables Specific to Herbivorous Mammals Compartment

HERBIVOROUS MAMMALS SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.025	0.075	0.050	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.050	0.150	0.100	Triangular
Terrestrial Plants, TP	0.350	1.050	0.700	Triangular
Aquatic Plants, AP	0.050	0.150	0.100	Triangular

¹ Estimated

Table 23. Stochastic Input Values for Variables Specific to Terrestrial Carnivorous Invertebrates Compartment

TERRESTRIAL CARNIVOROUS INVERTEBRATES SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.100	0.300	0.200	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.025	0.075	0.050	Triangular
Terrestrial Herbivorous Invertebrates, THI	0.300	0.900	0.600	Triangular
Aquatic Carnivorous Invertebrates, ACI	0.025	0.075	0.050	Triangular
Aquatic Herbivorous Invertebrates, AHI	0.025	0.075	0.050	Triangular

¹ Estimated

Table 24. Stochastic Input Values for Variables Specific to Terrestrial Herbivorous Invertebrates Compartment

TERRESTRIAL HERBIVOROUS INVERTEBRATES SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.050	0.150	0.100	Triangular
Sediment	0.050	0.150	0.100	Triangular
Water	0.025	0.075	0.050	Triangular
Terrestrial Plants, TP	0.350	1.050	0.700	Triangular
Aquatic Plants, AP	0.025	0.075	0.050	Triangular

¹ Estimated

Table 25. Stochastic Input Values for Variables Specific to Terrestrial Plants Compartment

TERRESTRIAL PLANTS SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.375	1.125	0.750	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.100	0.300	0.200	Triangular

¹ Estimated

Table 26. Stochastic Input Values for Variables Specific to Waterfowl Compartment

WATERFOWL SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.025	0.075	0.050	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.100	0.300	0.200	Triangular
Terrestrial Carnivorous Invertebrates, TCI	0.025	0.075	0.050	Triangular
Terrestrial Herbivorous Invertebrates, THI	0.025	0.075	0.050	Triangular
Terrestrial Plants, TP	0.050	0.150	0.100	Triangular
Carnivorous Fish, CF	0.050	0.150	0.100	Triangular
Herbivorous Fish, HF	0.050	0.150	0.100	Triangular
Aquatic Carnivorous Invertebrates, ACI	0.050	0.150	0.100	Triangular
Aquatic Herbivorous Invertebrates, AHI	0.050	0.150	0.100	Triangular
Aquatic Plants, AP	0.050	0.150	0.100	Triangular

¹ Estimated

Table 27. Stochastic Input Values for Variables Specific to Carnivorous Fish Compartment

CARNIVOROUS FISH SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.025	0.075	0.050	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.100	0.300	0.200	Triangular
Terrestrial Carnivorous Invertebrates, TCI	0.025	0.075	0.050	Triangular
Terrestrial Herbivorous Invertebrates, THI	0.025	0.075	0.050	Triangular
Herbivorous Fish, HF	0.025	0.075	0.050	Triangular
Aquatic Carnivorous Invertebrates, ACI	0.025	0.075	0.050	Triangular
Aquatic Herbivorous Invertebrates, AHI	0.025	0.075	0.050	Triangular

¹ Estimated

Table 28. Stochastic Input Values for Variables Specific to Herbivorous Fish Compartment

HERBIVOROUS FISH SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.025	0.075	0.050	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.100	0.300	0.200	Triangular
Aquatic Plants, AP	0.035	1.050	0.700	Triangular

¹ Estimate

Table 29. Stochastic Input Values for Variables Specific to Aquatic Carnivorous Invertebrates Compartment

AQUATIC CARNIVOROUS INVERTEBRATES SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.025	0.075	0.050	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.100	0.300	0.200	Triangular
Terrestrial Carnivorous Invertebrates, TCI	0.025	0.075	0.050	Triangular
Terrestrial Herbivorous Invertebrates, THI	0.025	0.075	0.050	Triangular
Aquatic Herbivorous Invertebrates, AHI	0.300	0.900	0.600	Triangular

¹ Estimate

Table 30. Stochastic Input Values for Variables Specific to Aquatic Herbivorous Invertebrates Compartment

AQUATIC HERBIVOROUS INVERTEBRATES SOURCE COMPARTMENTS	DIETARY FRACTION, df, (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.025	0.075	0.050	Triangular
Sediment	0.025	0.075	0.050	Triangular
Water	0.100	0.300	0.200	Triangular
Aquatic Plants, AP	0.350	1.050	0.700	Triangular

¹ Estimate

Table 31. Stochastic Input Values for Variables Specific to Aquatic Plants Compartment

AQUATIC PLANTS SOURCE COMPARTMENTS	DIETARY FRACTION, <i>df</i> , (unitless) ¹			DISTRIBUTION TYPE
	INPUT RANGE			
	MIN	MAX	MEAN	
Soil	0.025	0.075	0.050	Triangular
Sediment	0.375	1.125	0.750	Triangular
Water	0.100	0.300	0.200	Triangular

¹ Estimate

2.5 Model Development and Use

The model which has been described here can be incorporated into readily available personal computer spreadsheet software. For this work, Microsoft® Excel Version 7.0 was used. Lotus® 1-2-3, or other similar desktop spreadsheets would be equally acceptable for this type of analysis. The Monte Carlo sampling portion of the modeling employed the use of @RISK, Risk Analysis and Simulation Add-In for Microsoft® Excel or Lotus® 1-2-3 (Palisade 1996). BestFit, Probability Distribution Fitting for Windows®, a companion product to @RISK, was used as well to identify the distribution type for the water concentration contaminant level input range (Palisade 1998).

2.5.1 Environment and Architecture

As viewed from the spreadsheet, the model consists of nine worksheets. A flow diagram of the model is provided as Figure 4. The first worksheet, labeled INTRO, provides a brief description of the other eight worksheets and their contents. The INTRO worksheet also provides instruction on saving input parameters and output results.

The second worksheet, labeled MATRIX, includes a graphical representation

of the simplified wetland ecosystem food web matrix similar to that of Figure 2. The third worksheet, EQUATIONS, provides a description of the mathematical terms on which the model is based.

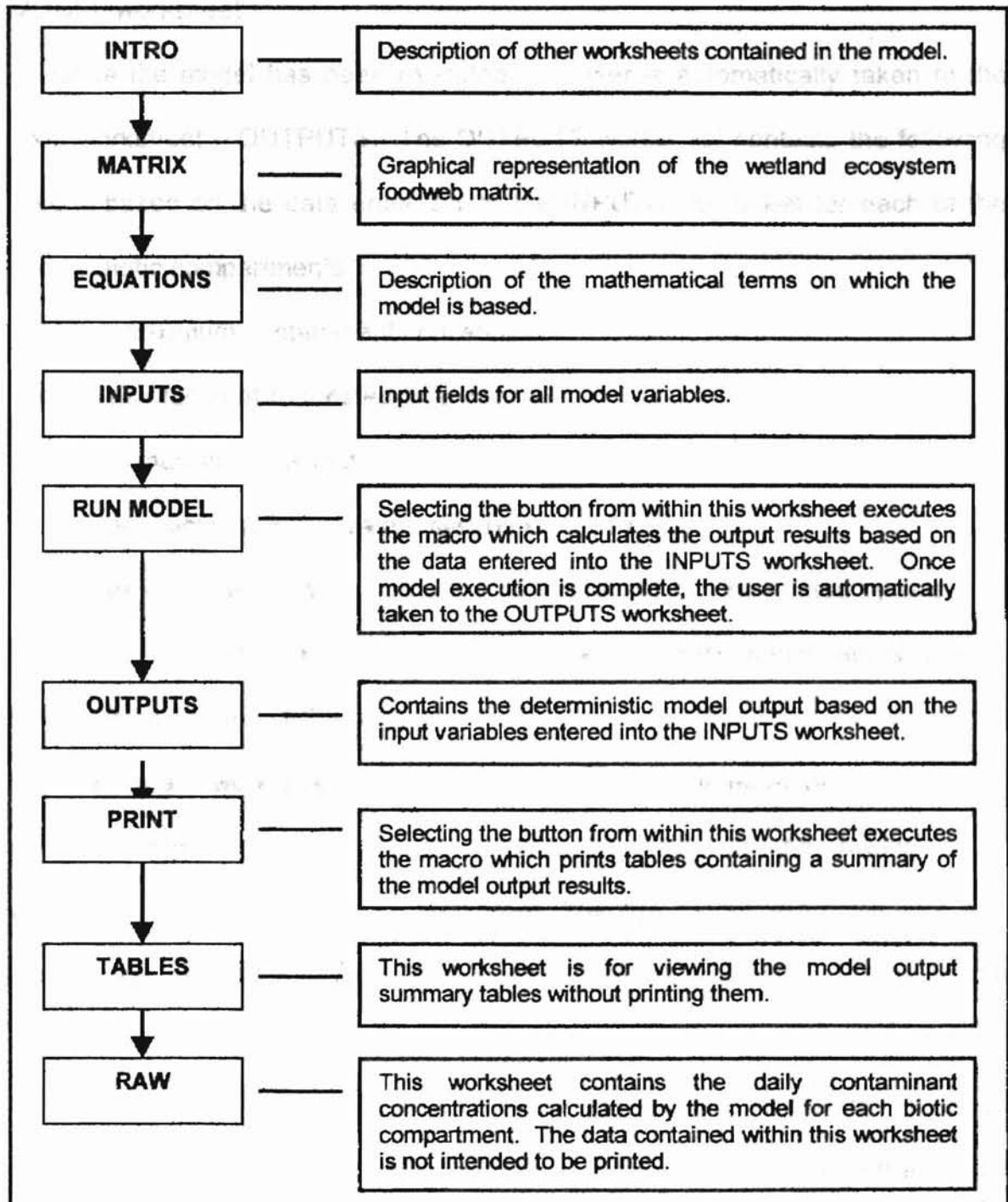


Figure 4. Model flow diagram.

The **INPUTS** worksheet contains input fields for all of the variables associated with the fifteen compartments which comprise the model. Upon entering all required input data, the model can be executed from the **RUN MODEL** worksheet.

Once the model has been executed, the user is automatically taken to the sixth worksheet – **OUTPUTS**. The **OUTPUTS** worksheet contains the following results based on the data entered into the **INPUTS** worksheet for each of the twelve biotic compartments:

C_{max} , maximum contaminant concentration;

RF_t , risk factor at requested time interval;

RF_{max} , maximum risk factor.

In order to calculate RF_t , a value must be entered for C_t .

Tables containing a summary of the model results can be printed by selecting the button located within the **PRINT** worksheet. These same tables can be viewed from the **TABLES** worksheet.

The final worksheet, **RAW**, contains the incremental contaminant concentrations calculated by the model for each of the twelve biotic compartments. It is from this worksheet that the model estimates risk and summarizes results in the **OUTPUTS** and **TABLES** worksheets. Due to the number of data points present in the raw data output, it exists only for the successful execution of the model and generation of desired results. Although the user can print and view these raw data, it could be rather awkward and time consuming and is therefore not recommended.

2.5.2 Saving Input Data and Output Results

Each time a new set of input parameters is entered into the model and the model is executed, the old input data and results are overwritten. Therefore, a companion file to the model, filename DATA.xls, was created that automatically updates the current input parameters and output results when opened simultaneously with the model, filename MODEL.xls. The user can manage the DATA.xls file as with any other Excel workbook. This will allow for efficient archiving and replication of previous data sets as well as to conserve disk space. The user should not corrupt the DATA.xls filename designation, but rather save previous work under another user-defined filename.

2.5.3 Automation

In an effort to increase efficiency and applicability, decrease confusion, and minimize human error, several steps within the model have been greatly simplified or completely automated via macros (see Appendix B for a complete listing of the macro code used for the model). Microsoft Excel employs the Visual Basic programming language for creating macros. The macros within the model allow the user to perform fairly simple tasks such as "point-and-click" printing of data tables and instantaneous archiving of current input variables and output data. A much more complex macro routine, developed for this research, greatly adds to the utility of the model. This macro allows the user to employ a wide range for the average life span variable without the need to repeatedly edit the raw data output range.

The initial portion of the code turns off the normal screen updating feature when the model is executed. This makes the macro more transparent and speeds up the model execution time (Walkenbach 1996).

```
Sub Run_Model()  
    Application.ScreenUpdating = False
```

The next section of the code selects the RAW worksheet and then clears the existing raw data output from the previous model run for each of the twelve biotic compartments. Although only the top carnivore compartment code is shown in this excerpt, the code for the other compartments is similar.

```
Sheets("RAW").Select  
'Clear previous TOP CARNIVORE results  
Range("B9").Select  
Range(ActiveCell, ActiveCell.End(xlDown)).Select  
Selection.Borders(xlLeft).LineStyle = xlNone  
Selection.Borders(xlRight).LineStyle = xlNone  
Selection.Borders(xlTop).LineStyle = xlNone  
Selection.Borders(xlBottom).LineStyle = xlNone  
Selection.BorderAround LineStyle:=xlNone  
Selection.ClearContents  
Range("C9").Select  
Range(ActiveCell, ActiveCell.End(xlDown)).Select  
Selection.Borders(xlLeft).LineStyle = xlNone  
Selection.Borders(xlRight).LineStyle = xlNone  
Selection.Borders(xlTop).LineStyle = xlNone  
Selection.Borders(xlBottom).LineStyle = xlNone  
Selection.BorderAround LineStyle:=xlNone  
Selection.ClearContents
```

The next section of the code calculates the new results for each abiotic compartment based on the data entered into the INPUTS worksheet. Notice the range reference format has changed from absolute (i.e., B8) to relative (i.e., 8,2). This allows the programmer to incorporate mathematical functions into the macro which in turn allows the modeler to manipulate the macro without actually changing the code. In this case, it allows the macro to handle potentially highly

variable average life span input values. Again, only the top carnivore compartment is shown here.

```
'Calculate new results for TOP CARNIVORE
Range(Cells(8, 2), Cells(8, 3)).Select
Selection.Copy
Range(Cells(8 + 1, 2), Cells(8 + (Cells(4, 3).Value - 1), 3)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 3).Value - 1), 2), Cells(8 + (Cells(4, 3).Value - 1), 3)).Select
With Selection.Borders(xlBottom)
    .Weight = xlMedium
    .ColorIndex = xlAutomatic
End With
'Return to cell A1 of RAW DATA OUTPUT worksheet
Cells(1, 1).Select
```

The final portion of the code returns the user to the OUTPUTS worksheet where the model results can be viewed.

```
'Return to cell A1 of OUTPUTS worksheet
Sheets("OUTPUTS").Select
Cells(1, 1).Select
End Sub
```

2.6 Kesterson Reservoir Site Description

As mentioned, the data set used for this model was taken from selenium contamination at Kesterson Reservoir, California. The Kesterson site, as described by the USBR (1990) and Freedman (1995), in addition to twelve shallow ponds (1.0 – 1.5 m deep) totaling approximately 500 ha, is composed of four main habitat types. These are: (1) grasslands dominated by saltgrass (*Distichlis spicata*), (2) soil-filled former wetland areas dominated by annual plant species such as burning bush (*Kochia spp.*) and grasses, (3) open areas sparsely covered with burning bush (*Kochia spp.*), prickly lettuce (*Lactuca serriola*), and clover (*Melilotus spp.*), and (4) intermittent rainwater puddles. Due

to the area's use as habitat by several important wildlife species and for human recreation, it has received much attention from ecologists regarding the high levels of selenium and other contaminant concentrations. The contaminant source was subsurface drainwaters from irrigated agricultural lands that were delivered to the reservoir via the San Luis Drain (CH2M Hill 1986). Delivery of this water to Kesterson was halted in mid-1986 (CH2M Hill 1986).

Effects of selenium overexposure at Kesterson were most noticeable in avian populations and included reproductive failure, reduced fecundity, and adult mortality (CH2M Hill 1986). Brain damage and other developmental abnormalities were highly prevalent among embryos and chicks (Freedman 1995). Several mammal species at Kesterson were shown to exhibit reduced reproductive success as compared to populations located at the Volta Wildlife Area (CH2M Hill 1986). Although no specific effects of overexposure were noticed for fish, invertebrates, and plants, Kesterson populations exhibited selenium levels of approximately 10 to 300 times higher than those of the reference wetland at the Volta Wildlife Area (Freedman 1995). Limited human health studies conducted on persons living adjacent to and working at the Kesterson site found selenium levels to be within normal ranges with no symptoms of overexposure (CH2M Hill 1986). Due to the levels of contamination found and the toxic effects witnessed, especially among bird populations, coupled with its ability to bioaccumulate, selenium was made the focus of risk assessment and mitigation efforts (CH2M Hill 1986).

Three levels of selenium removal were suggested by CH2M Hill (1986) and a

risk assessment was performed for each as well as the original condition. The cleanup alternatives were: (1) a flexible response plan (FRP) consisting of "spot-removal" of contaminated soil and vegetation, (2) removal and on-site storage of 450,000 yd³ (340,050 m³) of soil and vegetation (ONSITE-1), and (3) removal and on-site storage of 1,000,000 yd³ (764,555 m³) of soil and vegetation (ONSITE-2). The ONSITE-1 and ONSITE-2 alternatives would have reportedly reduced the soil selenium concentrations to 3.0 and 1.5 mg/kg respectively. The CH2M Hill (1986) analysis culminated in a risk characterization for the site for each cleanup alternative using a stochastic approach to describe a range and likelihood of possible exposure scenarios under each cleanup alternative. This effort focussed upon the application of an alternative modeling technique to these suggested remediation levels. As with the CH2M Hill (1986) effort, the output results from the model developed for this work were used to compare the compartmental selenium contaminant concentrations under pre-clean-up conditions and the ONSITE-1 and ONSITE-2 mitigation alternatives.

2.7 Analytical Design

Models were prepared which compared several selenium contaminant scenarios for the Kesterson site. Scenarios 1 through 3 assume that the 1986 biotic compartmental selenium concentrations were not in a state of equilibrium (referred to as ASSUMPTION A). The C_o values employed for these scenarios were arbitrarily selected but critical to model performance. Scenarios 1 through 3 are defined as follows:

Scenario 1. Initial conditions present at 1986;

Scenario 2. Reduced soil, sediment, and water concentrations representative of ONSITE-1 cleanup goals with biotic compartment concentrations at 1986 levels; and

Scenario 3. Reduced soil, sediment, and water concentrations representative of ONSITE-2 cleanup goals with biotic compartment concentrations at 1986 levels.

The remaining five scenarios (4 through 6) assume that the biotic compartmental selenium concentrations reported by CH2M Hill (1986) and USBR (1990) represent conditions at equilibrium (referred to as ASSUMPTION B). For these analyses, the 1986 compartmental selenium concentrations were entered as the endpoint concentration (C_e) and the initial concentrations (C_o) were set to 0.01 mg/kg for all compartments. The 0.01 mg/kg value falls within the typical range for selenium in food (ASTDR 1989). Scenarios 4 through 6 are similar to 1 through 3 and are defined as follows:

Scenario 4. Initial conditions present at 1986;

Scenario 5. Reduced soil, sediment, and water concentrations representative of ONSITE-1 cleanup goals with biotic compartment endpoint concentrations (C_e) set at 1986 measured levels; and

Scenario 6. Reduced soil, sediment, and water concentrations representative of ONSITE-2 cleanup goals with biotic compartment endpoint concentrations (C_e) set at 1986 measured levels.

3. RESULTS

The deterministic output data returned the incremental contaminant concentration for each biotic compartment. This incremental selenium concentration was used to compare the time to C_e for each biotic compartment under the initial 1986 conditions (scenarios 1 and 4) and the ONSITE-1 (scenarios 2 and 5) and ONSITE-2 (scenarios 3 and 6) cleanup alternatives proposed by CH2M Hill (1986). The stochastic approach was used to (1) estimate the relative effects each of the input parameters had on the output results via a rank order correlation and (2) to generate and compare contaminant concentration probability distribution functions of each biotic compartment for the six modeled scenarios and two basic equilibrium assumptions.

3.1 Deterministic Output Data

The deterministic, or point, analysis was repeated three times under the two equilibrium state assumption criteria (once for each of the six described scenarios). Figures 5 through 28 compare the selenium concentration for each biotic compartment for scenarios 1 through 3 (ASSUMPTION A) and scenarios 4 through 6 (ASSUMPTION B) up to 100% of the endpoint concentration.

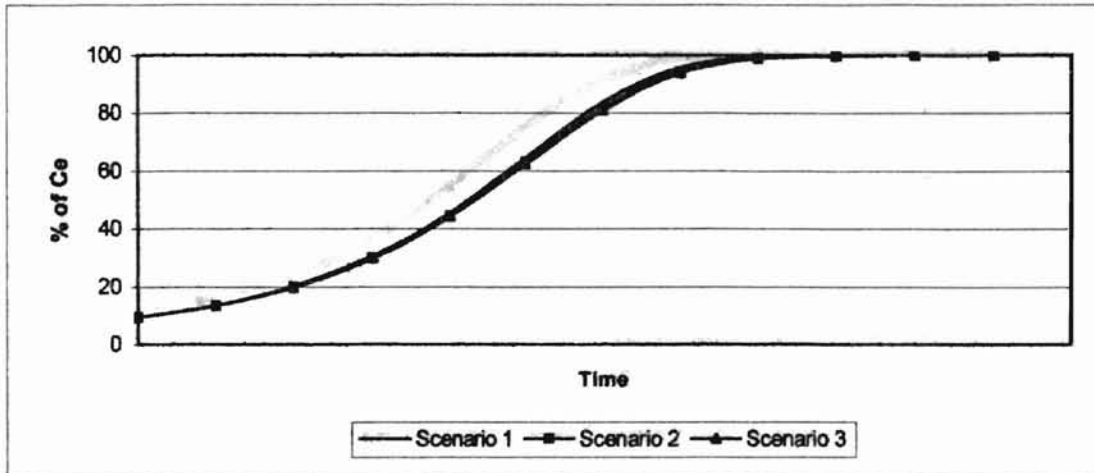


Figure 5. Top carnivore compartment time to 100% of endpoint concentration comparison.

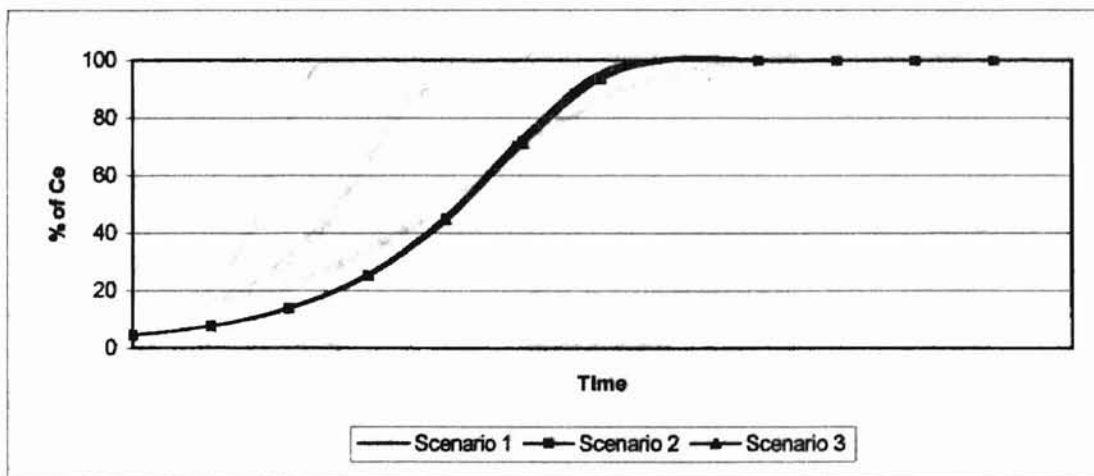


Figure 6. Waterfowl compartment time to 100% of endpoint concentration comparison.

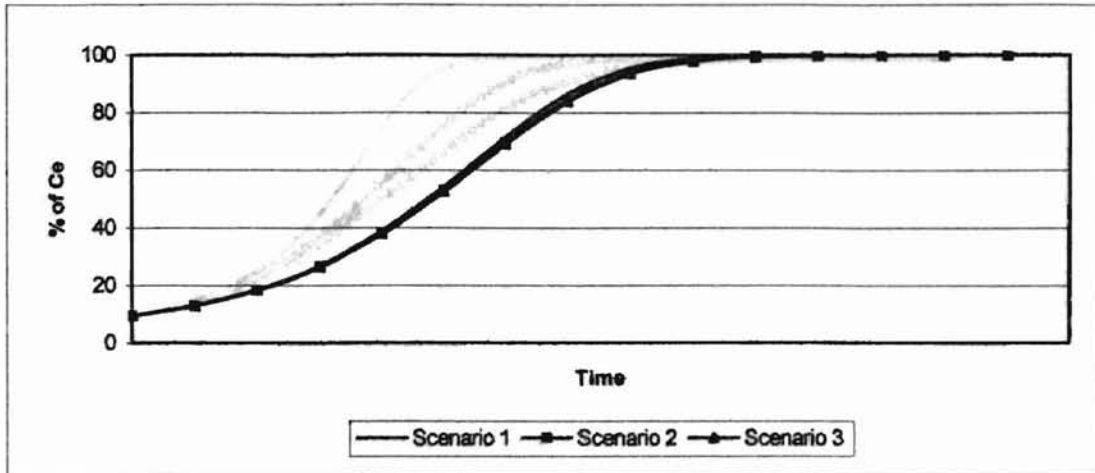


Figure 7. Carnivorous mammals compartment time to 100% of endpoint concentration comparison.

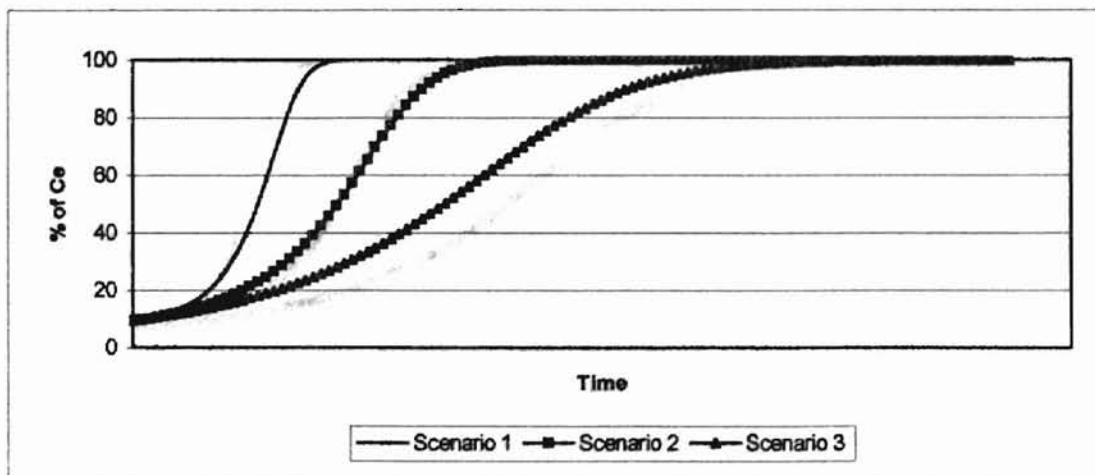


Figure 8. Herbivorous mammals compartment time to 100% of endpoint concentration comparison.

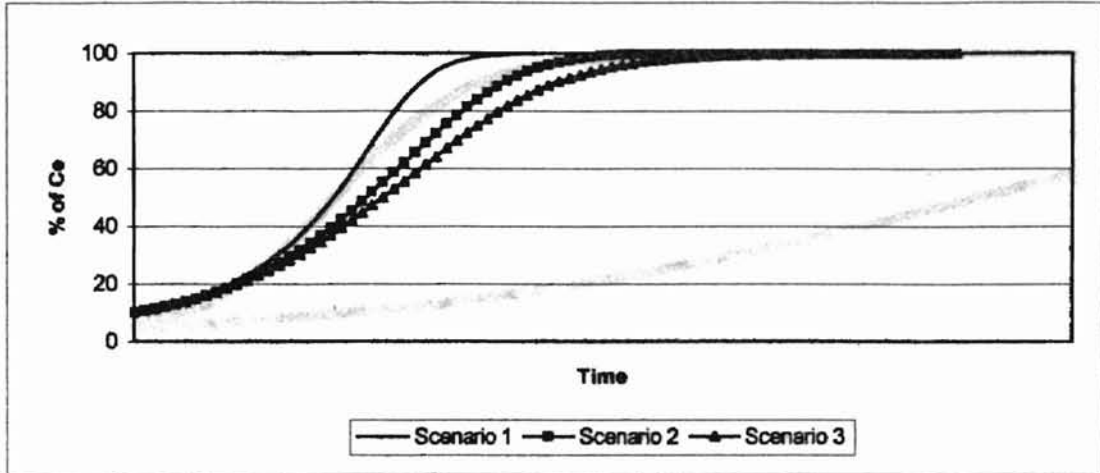


Figure 9. Terrestrial carnivorous invertebrates compartment time to 100% of endpoint concentration comparison.

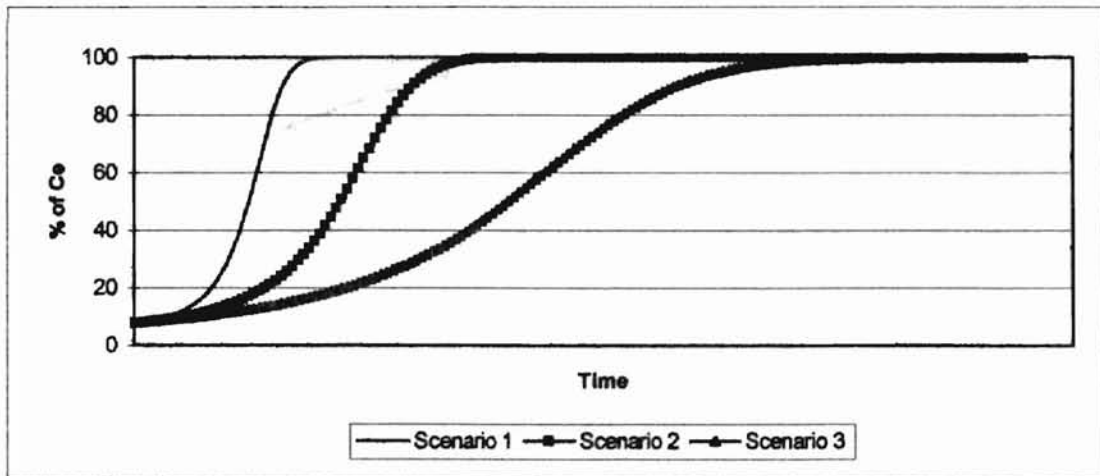


Figure 10. Terrestrial herbivorous invertebrates compartment time to 100% of endpoint concentration comparison.

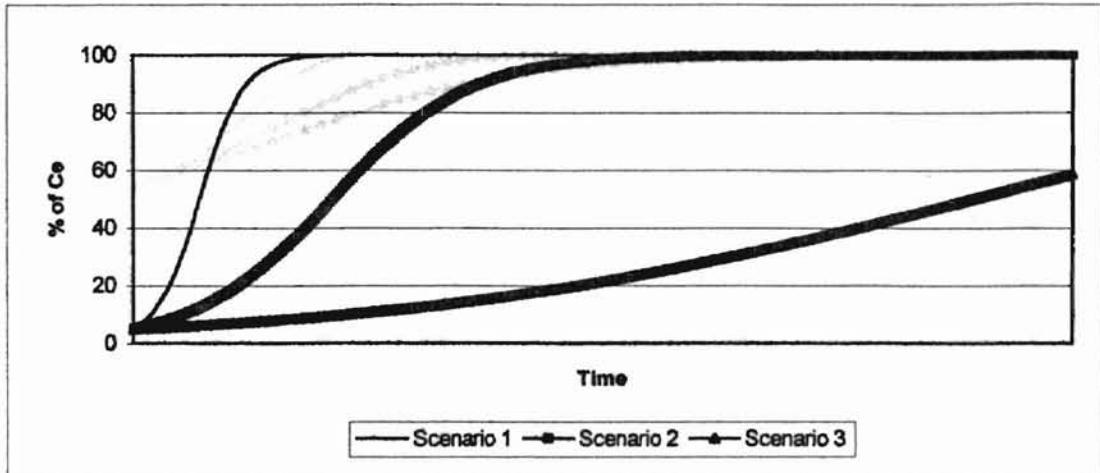


Figure 11. Terrestrial plants compartment time to 100% of endpoint concentration comparison.

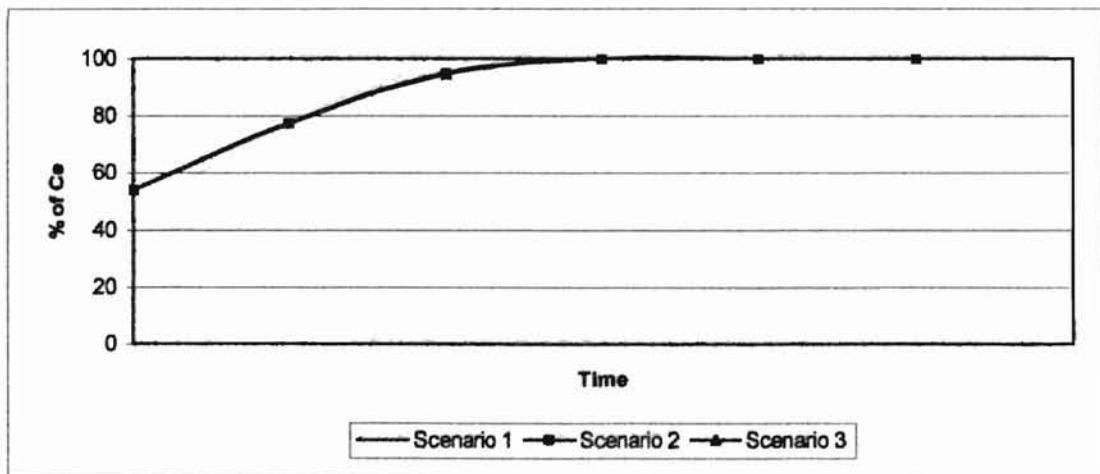


Figure 12. Carnivorous fish compartment time to 100% of endpoint concentration comparison.

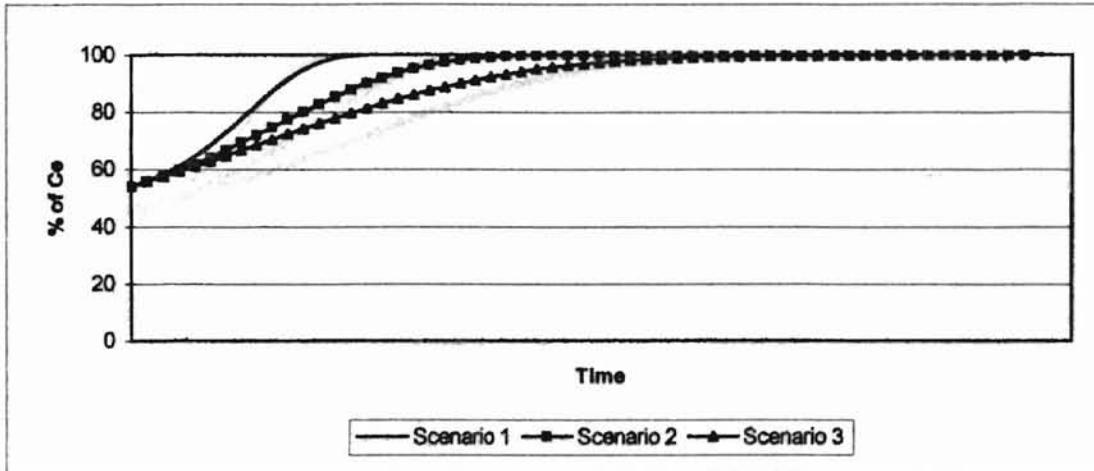


Figure 13. Herbivorous fish compartment time to 100% of endpoint concentration comparison.

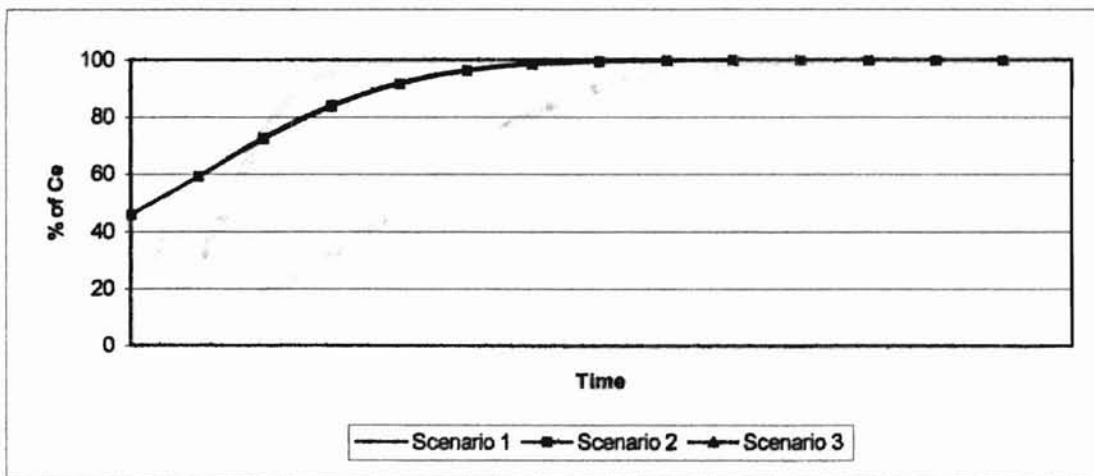


Figure 14. Aquatic carnivorous invertebrates compartment time to 100% of endpoint concentration comparison.

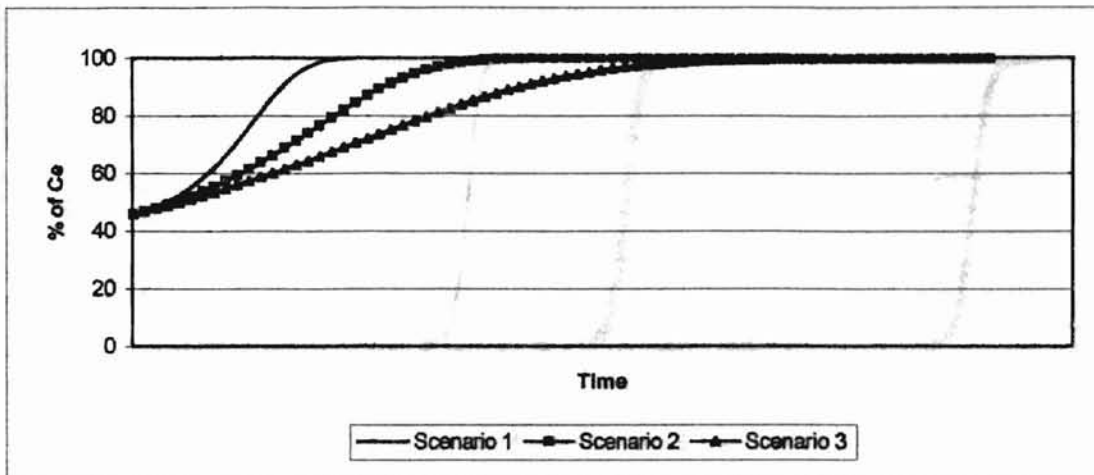


Figure 15. Aquatic herbivorous invertebrates compartment time to 100% of endpoint concentration comparison.

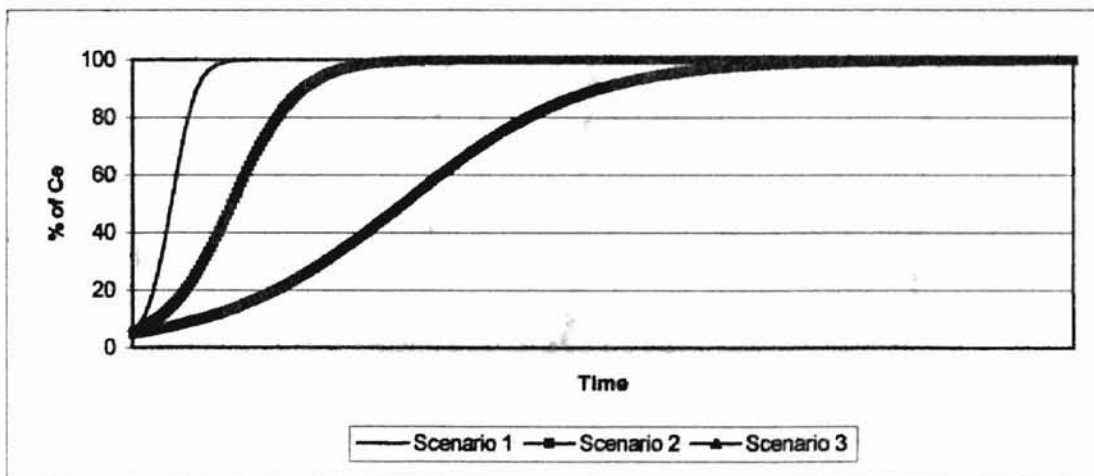
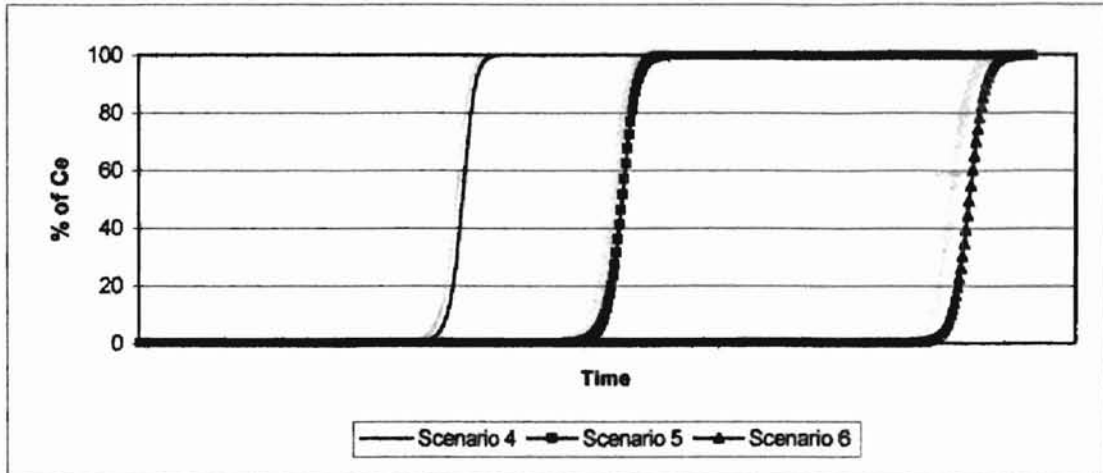


Figure 16. Aquatic plants compartment time to 100% of endpoint concentration comparison.



5 Figure 17. Top carnivore compartment time to 100% of endpoint concentration comparison.

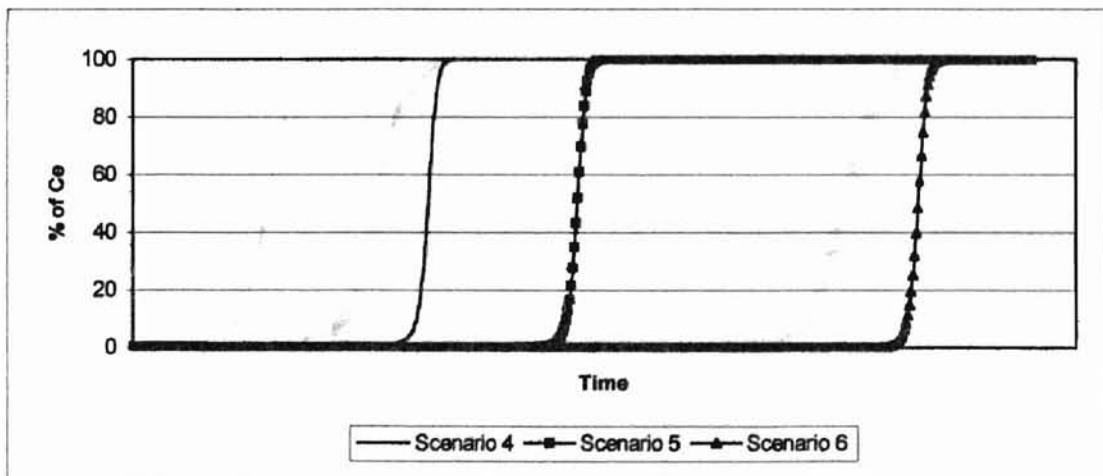


Figure 18. Waterfowl compartment time to 100% of endpoint concentration comparison.

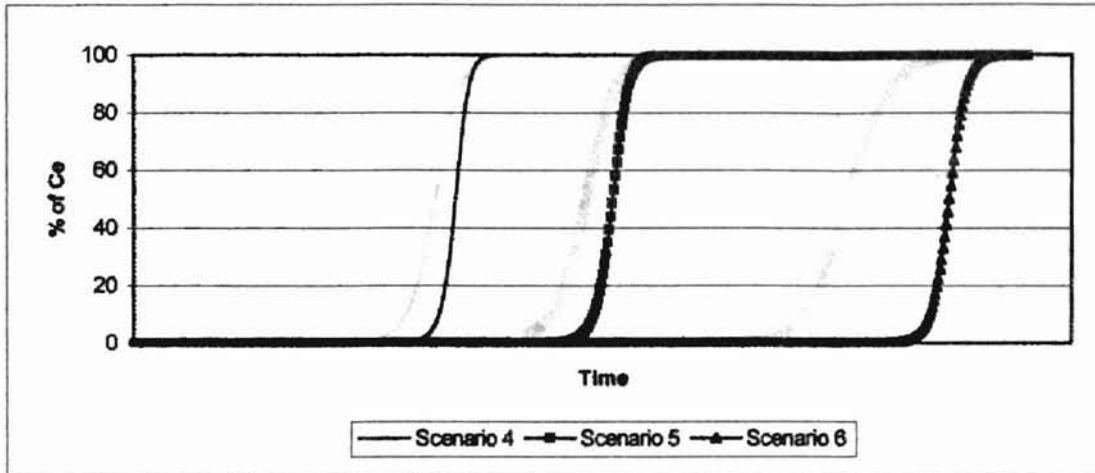


Figure 19. Carnivorous mammals compartment time to 100% of endpoint concentration comparison.

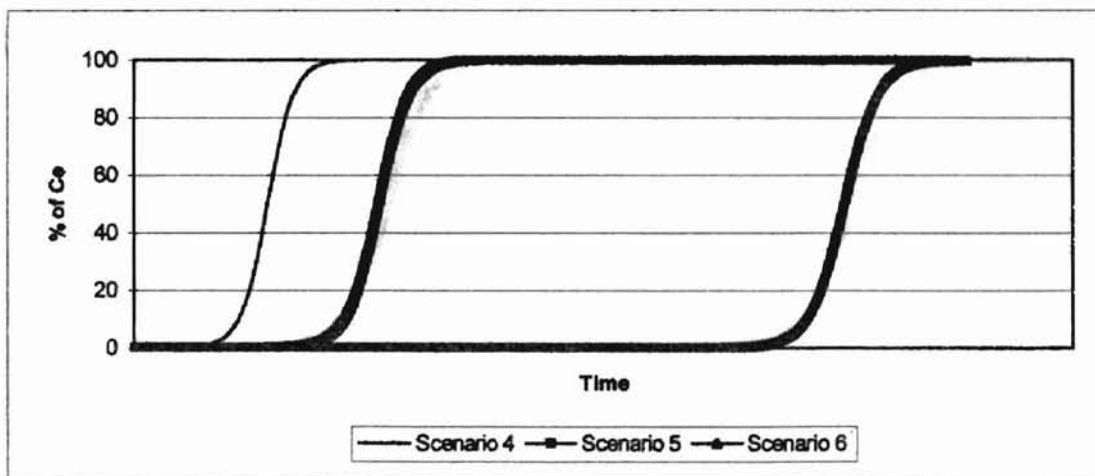


Figure 20. Herbivorous mammals compartment time to 100% of endpoint concentration comparison.

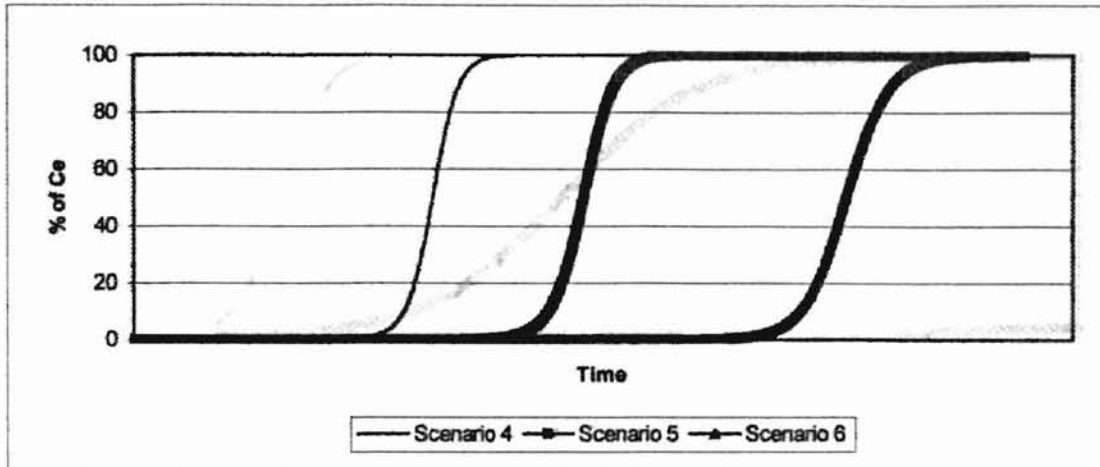


Figure 21. Terrestrial carnivorous invertebrates compartment time to 100% of endpoint concentration comparison.

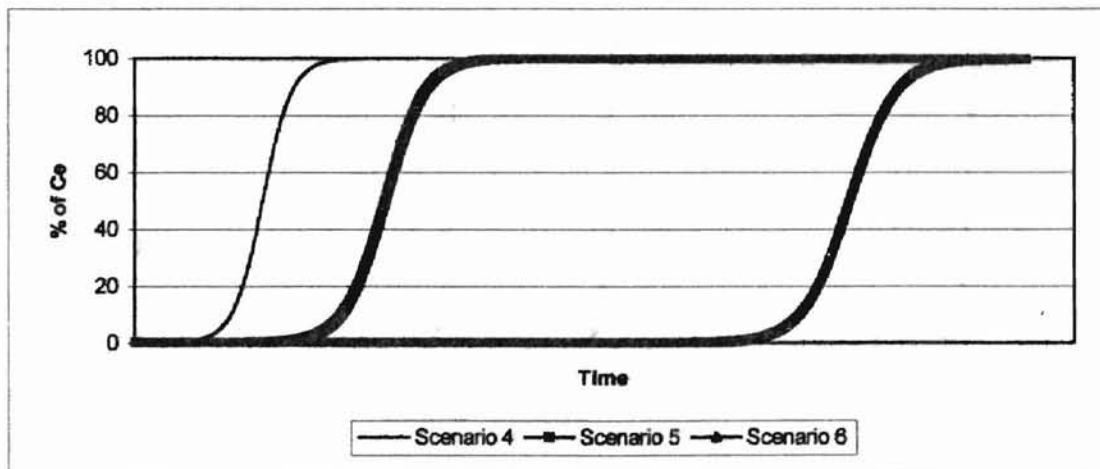


Figure 22. Terrestrial herbivorous invertebrates compartment time to 100% of endpoint concentration comparison.

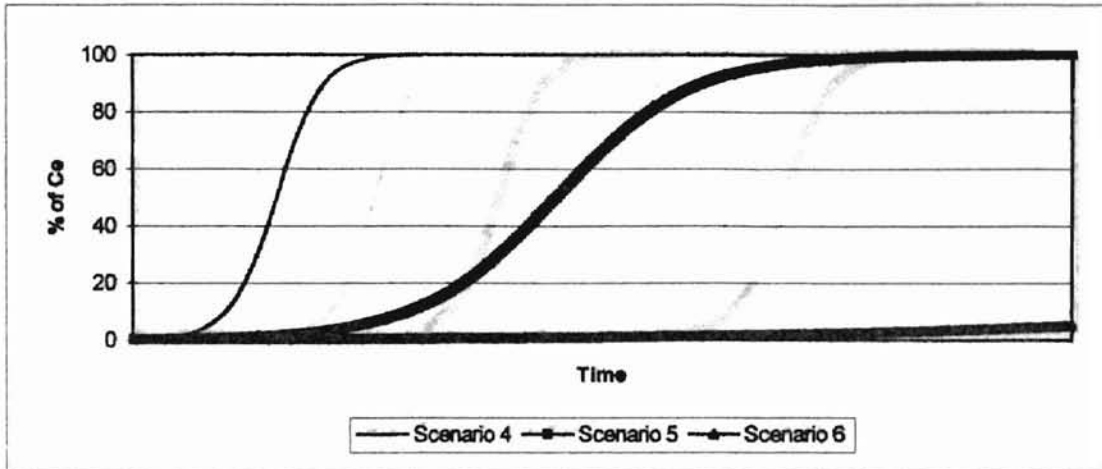


Figure 23. Terrestrial plants compartment time to 100% of endpoint concentration comparison.

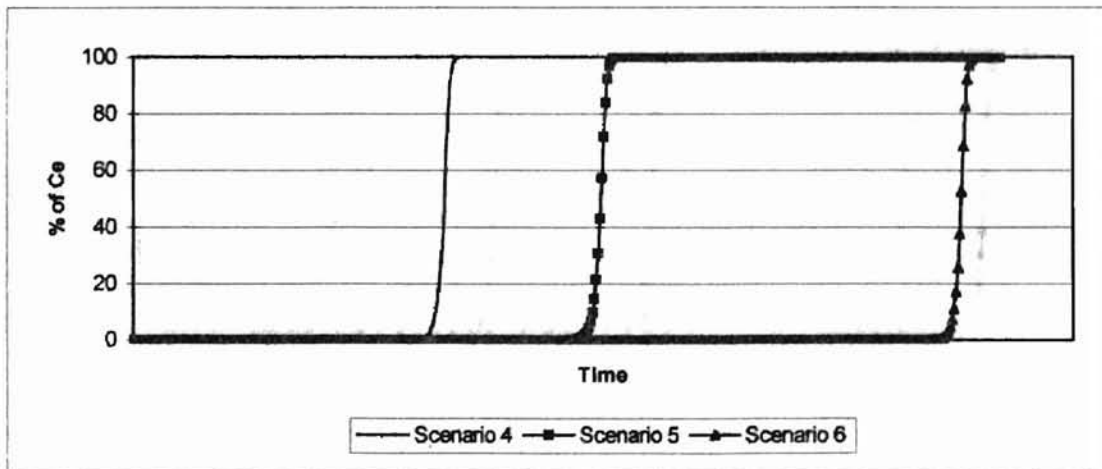


Figure 24. Carnivorous fish compartment time to 100% of endpoint concentration comparison.

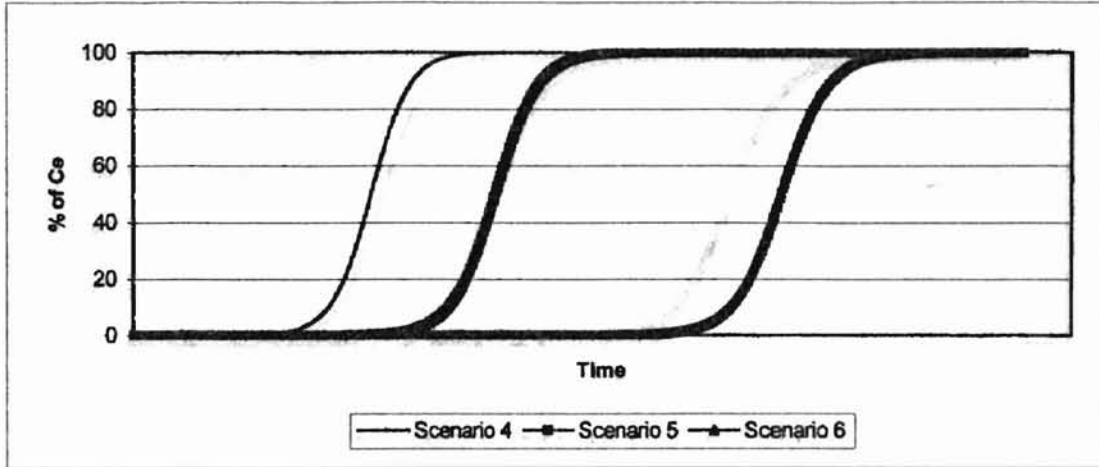


Figure 25. Herbivorous fish compartment time to 100% of endpoint concentration comparison.

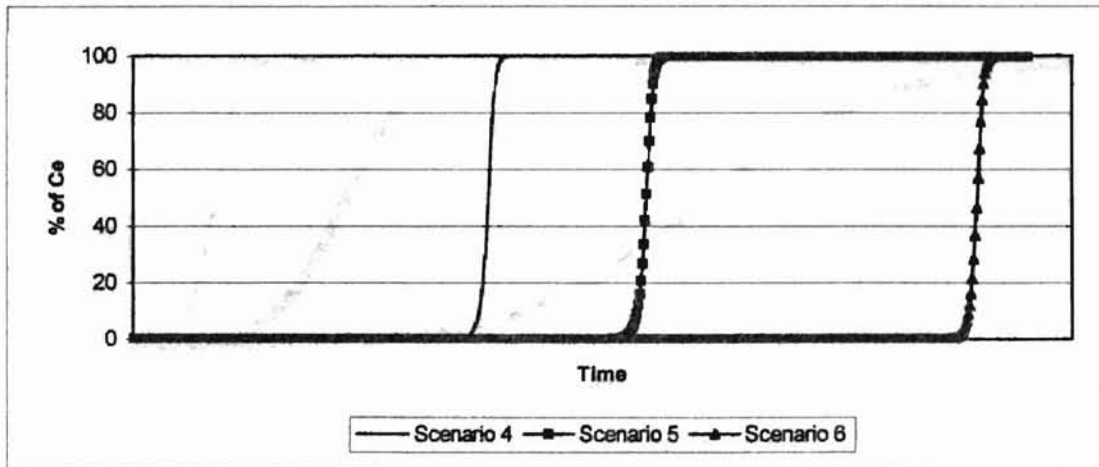


Figure 26. Aquatic carnivorous invertebrates compartment time to 100% of endpoint concentration comparison.

3.2 Stochastic Output Data

The stochastic estimate approach employed a Latin Hypercube sampling technique in order to generate probability distributions for the compartments

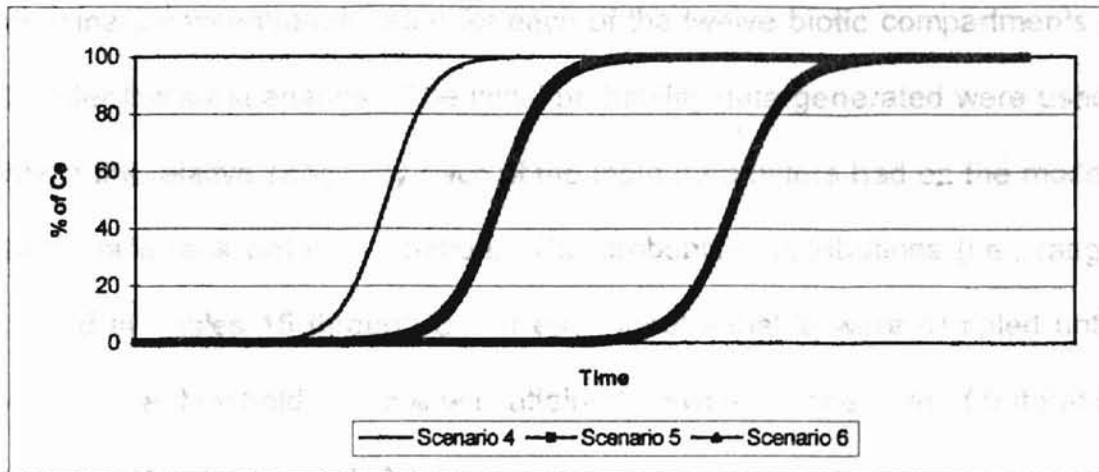


Figure 27. Aquatic herbivorous invertebrates compartment time to 100% of endpoint concentration comparison.

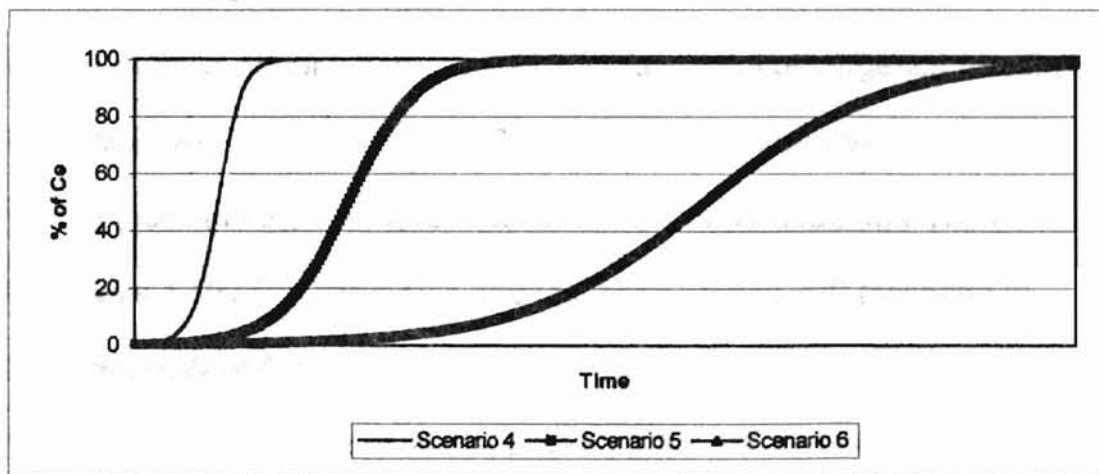


Figure 28. Aquatic plants compartment time to 100% of endpoint concentration comparison.

3.2 Stochastic Output Data

The stochastic estimate approach employed a Latin Hypercube sampling technique in order to generate probability distributions for the compartmental contaminant concentration value for each of the twelve biotic compartments at $t = 1$ under the six scenarios. The initial probability data generated were used to analyze the relative sensitivity each of the input parameters had on the modeled results via a rank order correlation. The probability distributions (i.e., ranges) provided in Tables 15 through 31 for each input variable were sampled until a convergence threshold of 1.5% was attained between successive 100 iterations based on the following statistics:

- percentiles
- mean
- standard deviation

The total number of iterations required to attain this convergence threshold for each of the five scenarios under ASSUMPTION A criteria was 1,400 (scenario 1), 1,300 (scenario 2), and 1,100 (scenario 3). Under ASSUMPTION B criteria the tally was 1,400 (scenario 4), 1,400 (scenario 5), and 700 (scenario 6).

3.2.1 Sensitivity of Outputs to Inputs

A rank order correlation analysis was performed to better understand the relative impact each of the uncertain input range variables had on the modeled results. A rank order correlation was chosen over a multivariate stepwise regression due to the very low R^2 value (i.e., $< 1\%$) returned for all compartments. An R^2 value less than approximately 60% suggests that the

linear multivariate stepwise regression does not sufficiently explain the relationship between the inputs and outputs (Palisade 1996). The results of the sensitivity analysis showed an almost perfect positive correlation between the modeled output and the initial contaminant concentration (C_0) for all compartments. The remaining input parameters showed extremely weak correlations, both positively and negatively. The sensitivity analysis was performed under ASSUMPTION A and B criteria with similar results.

3.2.2 Stochastic Output Result Ranges and Probability Distributions

Based on the input data ranges, the Latin Hypercube sampling analysis produced a range of possible values for each compartmental contaminant concentration ($C_{t=1}$). Probability distributions were also calculated for each compartment. The modeled ranges, means, standard deviations, and 25% and 75% confidence intervals, representing the intraquartile range, for each biotic compartment under each of the six scenarios are given in Tables 32 through 37. The probability distributions for each compartment under the six scenarios are presented as box and whisker diagrams in Figures 29 through 52. The upper and lower horizontal elements of each box represent the 75th and 25th percentiles respectively. The upper and lower vertical lines extend to the maximum and minimum selenium concentrations returned by the model under each scenario. Summary statistics for the stochastic analyses under the six scenarios are provided as Appendices C through H.

Table 32. Compartmental Se Stochastic Output Result Ranges, $C_{t=1}$ (mg/kg) under Scenario 1 – ASSUMPTION A Conditions

	TC	WF	CM	HM	TCI	THI	TP	CF	HF	ACI	AHI	AP
MAX	130.6	61.7	131.0	128.6	62.5	53.8	123.1	353.0	302.6	187.6	267.2	165.4
MIN	2.3	3.2	1.5	1.1	2.8	0.8	0.7	6.8	7.5	0.0	26.3	0.8
MEAN	54.2	20.1	54.5	50.3	31.9	20.9	39.2	157.5	139.3	79.7	136.1	45.4
SD	28.7	10.8	28.6	27.5	18.8	10.8	23.2	51.5	49.8	30.3	42.9	27.2
25%	29.5	11.7	30.3	27.4	15.8	12.0	19.7	126.0	106.9	59.6	106.0	23.3
75%	76.2	28.3	77.2	71.8	45.5	28.4	56.9	193.1	173.8	98.0	166.8	65.5

Table 33. Compartmental Se Stochastic Output Result Ranges, $C_{t=1}$ (mg/kg) under Scenario 2 – ASSUMPTION A Conditions

	TC	WF	CM	HM	TCI	THI	TP	CF	HF	ACI	AHI	AP
MAX	132.6	56.1	131.5	117.8	83.1	47.7	81.0	298.8	311.2	165.3	266.7	81.9
MIN	0.0	2.2	1.2	1.0	2.6	0.6	0.6	10.5	11.6	0.0	30.9	0.5
MEAN	56.6	20.5	53.0	49.5	32.4	19.1	29.2	154.9	136.1	81.1	133.2	29.4
SD	27.9	10.5	28.1	28.0	18.5	10.0	18.6	50.2	48.0	31.4	43.7	19.0
25%	34.5	11.5	29.2	26.2	16.6	11.0	13.3	124.0	105.9	61.3	101.1	13.1
75%	78.9	27.3	74.7	71.5	45.5	25.9	42.5	189.6	167.9	101.5	164.2	42.6

Table 34. Compartmental Se Stochastic Output Result Ranges, $C_{t=1}$ (mg/kg) under Scenario 3 – ASSUMPTION A Conditions

	TC	WF	CM	HM	TCI	THI	TP	CF	HF	ACI	AHI	AP
MAX	148.5	55.6	130.6	117.8	81.2	59.8	79.9	302.9	305.2	176.6	252.3	80.0
MIN	2.5	2.6	1.5	1.2	2.5	1.3	0.4	14.7	8.8	0.0	27.2	0.7
MEAN	55.8	20.1	54.2	50.9	31.5	19.5	29.0	155.3	136.7	80.1	132.4	29.2
SD	28.5	10.6	28.4	50.9	18.5	10.3	18.5	49.8	49.8	30.0	43.9	10.6
25%	33.3	11.6	29.8	25.4	15.6	10.7	13.3	121.4	102.4	60.5	99.9	13.0
75%	76.9	27.7	76.9	72.7	44.2	26.6	42.8	190.3	171.0	97.9	164.5	42.2

Table 35. Compartmental Se Stochastic Output Result Ranges, $C_{t=1}$ (mg/kg) under Scenario 4 – ASSUMPTION B Conditions

	TC	WF	CM	HM	TCI	THI	TP	CF	HF	ACI	AHI	AP
MAX	1.54E-02	1.56E-02	1.56E-02	1.59E-02	1.48E-02	1.68E-02	4.13E-02	1.51E-02	1.65E-02	1.47E-02	1.54E-02	6.94E-02
MIN	5.39E-03	5.30E-03	5.30E-03	5.21E-03	5.23E-03	5.08E-03	6.09E-03	5.19E-03	5.28E-03	5.15E-03	5.12E-03	6.14E-03
MEAN	1.04E-02	1.02E-02	1.02E-02	1.03E-02	9.92E-03	1.04E-02	1.57E-02	1.01E-02	1.05E-02	9.99E-03	1.02E-02	1.96E-02
SD	2.10E-03	2.10E-03	2.13E-03	2.17E-03	2.01E-03	2.20E-03	5.34E-03	2.13E-03	2.24E-03	1.98E-03	2.09E-03	8.31E-03
25%	8.89E-03	8.74E-03	8.75E-03	8.71E-03	8.48E-03	8.82E-03	1.19E-02	8.64E-03	8.84E-03	8.54E-03	8.75E-03	1.38E-02
75%	1.19E-02	1.17E-02	1.17E-02	1.18E-02	1.14E-02	1.20E-02	1.85E-02	1.16E-02	1.21E-02	1.15E-02	1.17E-02	2.37E-02

Table 36. Compartmental Se Stochastic Output Result Ranges, $C_{t=1}$ (mg/kg) under Scenario 5 – ASSUMPTION B Conditions

	TC	WF	CM	HM	TCI	THI	TP	CF	HF	ACI	AHI	AP
MAX	1.46E-02	1.47E-02	1.44E-02	1.46E-02	1.48E-02	1.46E-02	1.54E-02	1.47E-02	1.46E-02	1.48E-02	1.47E-02	1.67E-02
MIN	4.99E-03	4.99E-03	5.09E-03	5.06E-03	5.11E-03	5.11E-03	5.42E-03	5.06E-03	5.05E-03	5.24E-03	5.11E-03	5.28E-03
MEAN	1.00E-02	9.91E-03	9.94E-03	9.85E-03	9.80E-03	9.89E-03	1.03E-02	9.82E-03	9.85E-03	1.01E-02	9.80E-03	1.06E-02
SD	2.06E-03	2.04E-03	2.02E-03	2.03E-03	2.03E-03	2.03E-03	2.09E-03	2.05E-03	1.99E-03	2.06E-03	2.07E-03	2.20E-03
25%	8.54E-03	8.46E-03	8.58E-03	8.29E-03	8.30E-03	8.38E-03	8.82E-03	8.35E-03	8.50E-03	8.61E-03	8.37E-03	9.03E-03
75%	1.15E-02	1.14E-02	1.15E-02	1.13E-02	1.12E-02	1.13E-02	1.18E-02	1.13E-02	1.13E-02	1.16E-02	1.13E-02	1.22E-02

Table 37. Compartmental Se Stochastic Output Result Ranges, $C_{t=1}$ (mg/kg) under Scenario 6 – ASSUMPTION B Conditions

	TC	WF	CM	HM	TCI	THI	TP	CF	HF	ACI	AHI	AP
MAX	1.45E-02	1.45E-02	1.44E-02	1.45E-02	1.48E-02	1.45E-02	1.48E-02	1.45E-02	1.47E-02	1.45E-02	1.45E-02	1.55E-02
MIN	5.06E-03	5.21E-03	5.06E-03	4.98E-03	5.11E-03	5.14E-03	5.21E-03	4.96E-03	5.06E-03	5.08E-03	5.19E-03	5.47E-03
MEAN	9.67E-03	9.89E-03	9.74E-03	9.82E-03	9.98E-03	9.84E-03	1.01E-02	9.80E-03	9.74E-03	1.00E-02	9.92E-03	1.03E-02
SD	2.05E-03	1.93E-03	1.94E-03	1.99E-03	2.02E-03	1.98E-03	1.98E-03	2.01E-03	2.01E-03	1.99E-03	2.00E-03	2.11E-03
25%	8.26E-03	8.53E-03	8.26E-03	8.38E-03	8.56E-03	8.42E-03	8.82E-03	8.38E-03	8.32E-03	8.64E-03	8.42E-03	8.75E-03
75%	1.14E-02	1.13E-02	1.12E-02	1.12E-02	1.15E-02	1.12E-02	1.15E-02	1.12E-02	1.11E-02	1.14E-02	1.14E-02	1.19E-02

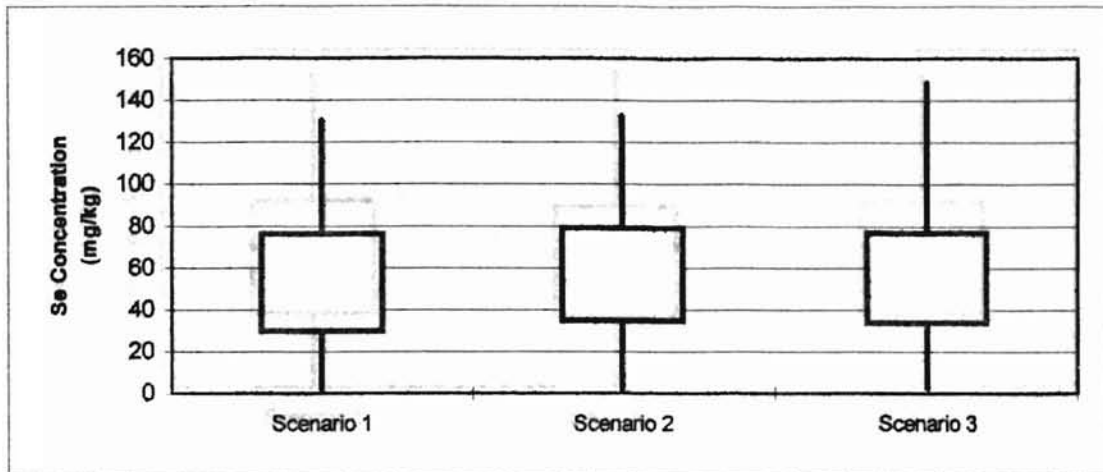


Figure 29. Probability distribution comparison for top carnivore compartment.

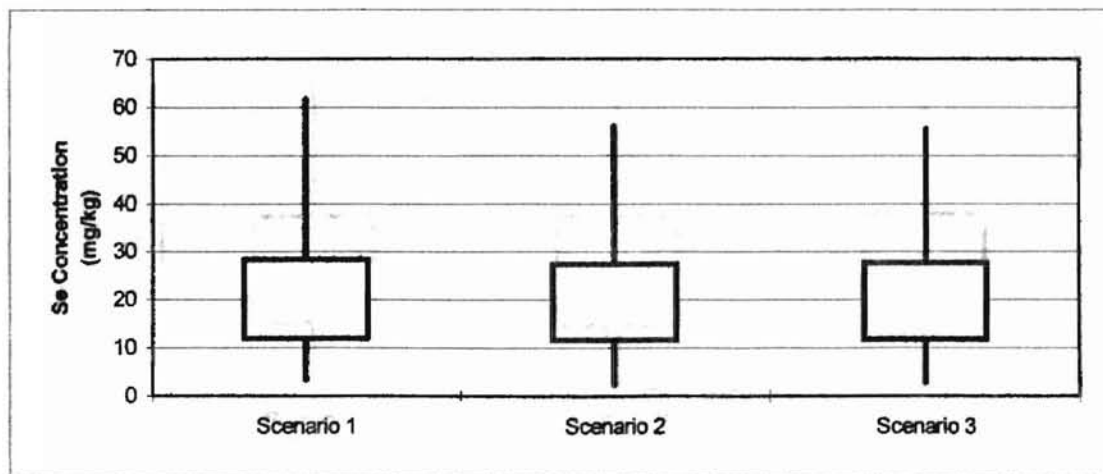


Figure 30. Probability distribution comparison for waterfowl compartment.

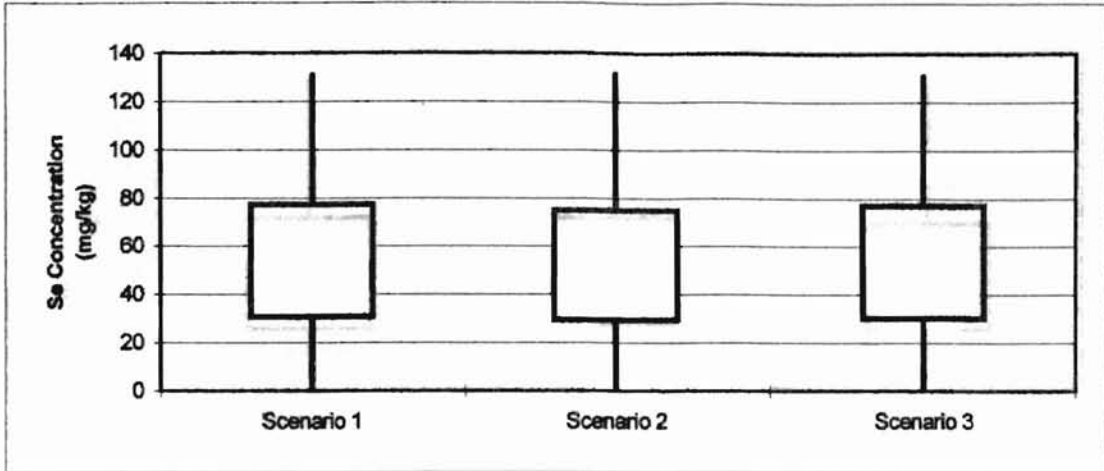


Figure 31. Probability distribution comparison for carnivorous mammals compartment.

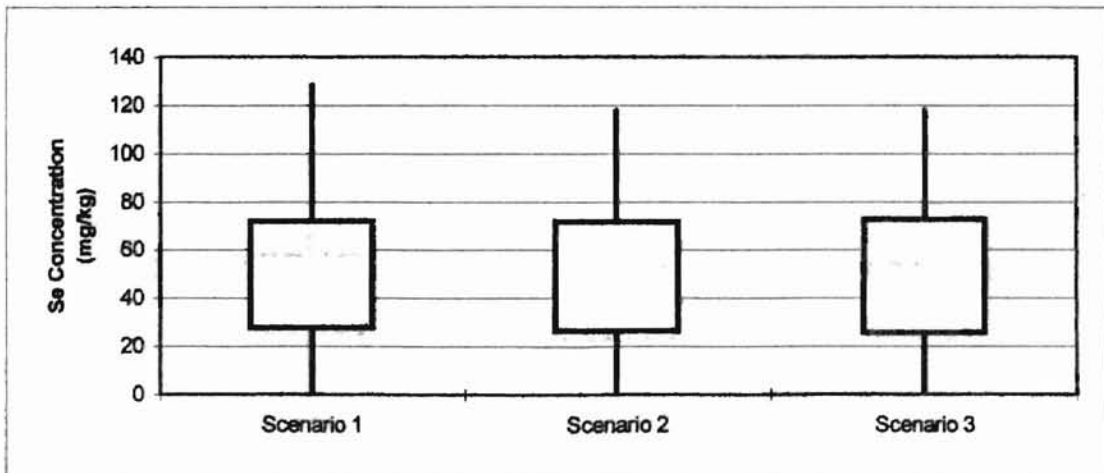


Figure 32. Probability distribution comparison for herbivorous mammals compartment.

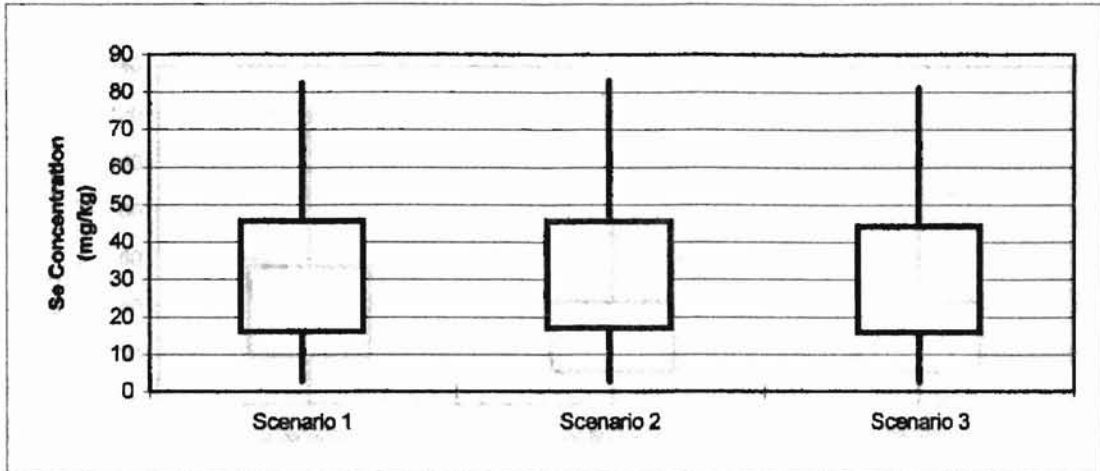


Figure 33. Probability distribution comparison for terrestrial carnivorous invertebrates compartment.

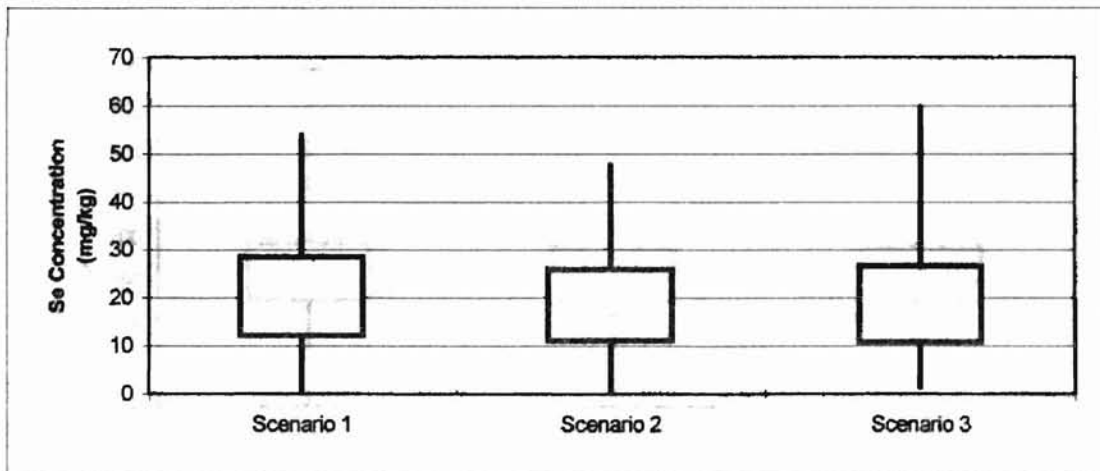


Figure 34. Probability distribution comparison for terrestrial herbivorous invertebrates compartment.

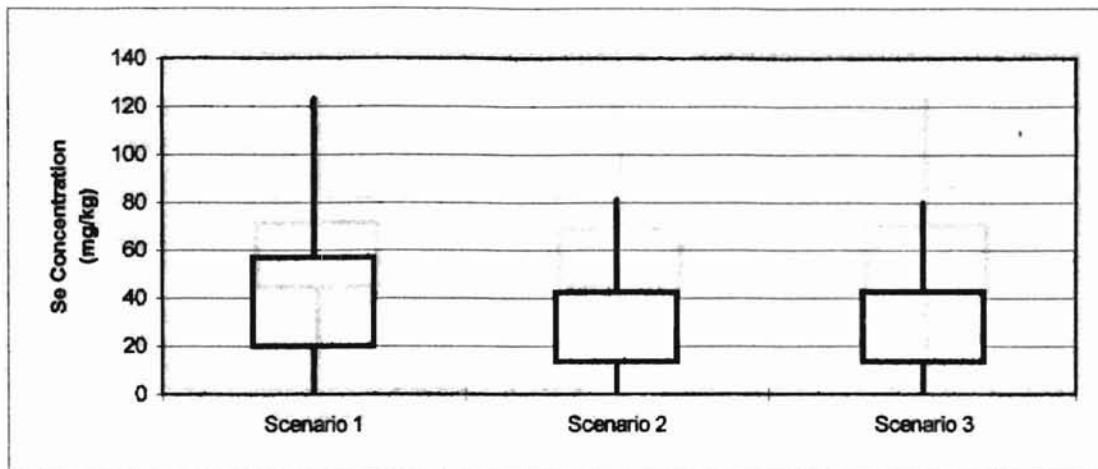


Figure 35. Probability distribution comparison for terrestrial plants compartment.

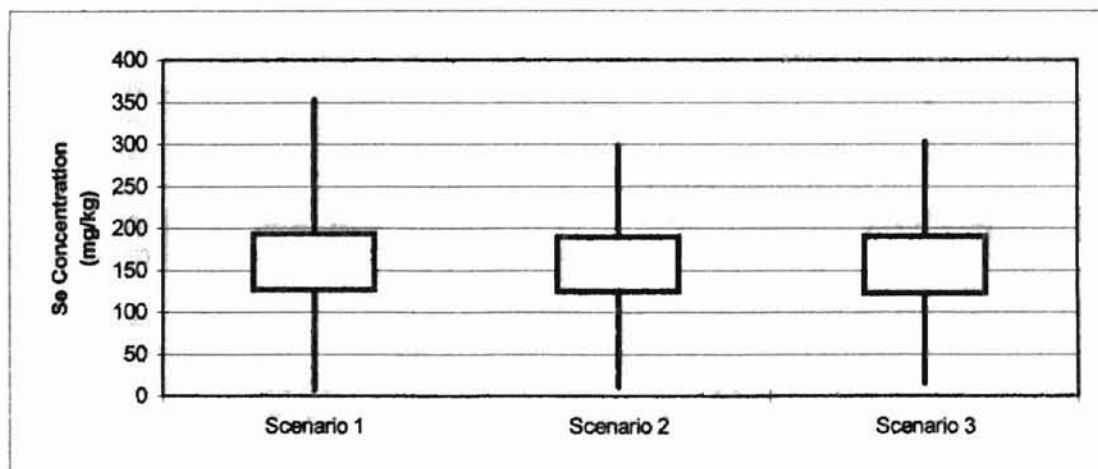


Figure 36. Probability distribution comparison for carnivorous fish compartment.

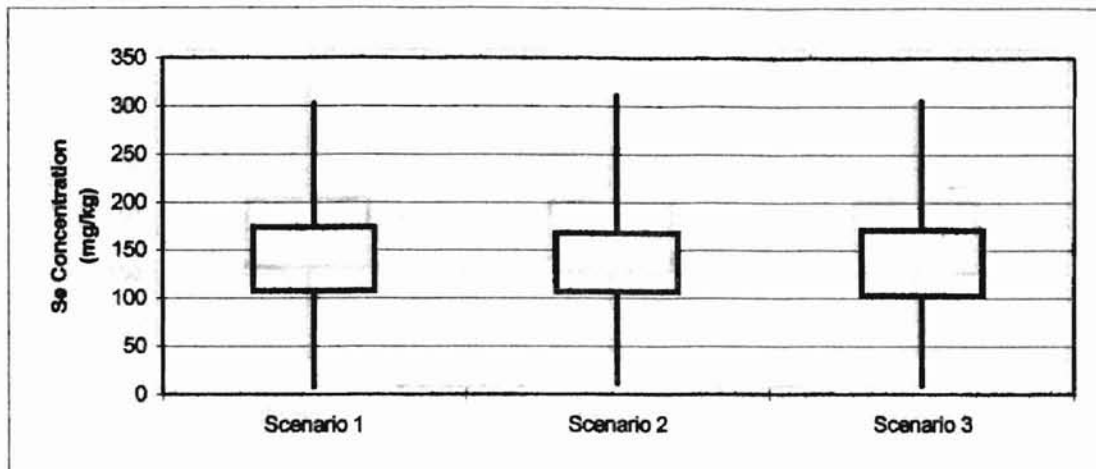


Figure 37. Probability distribution comparison for herbivorous fish compartment.

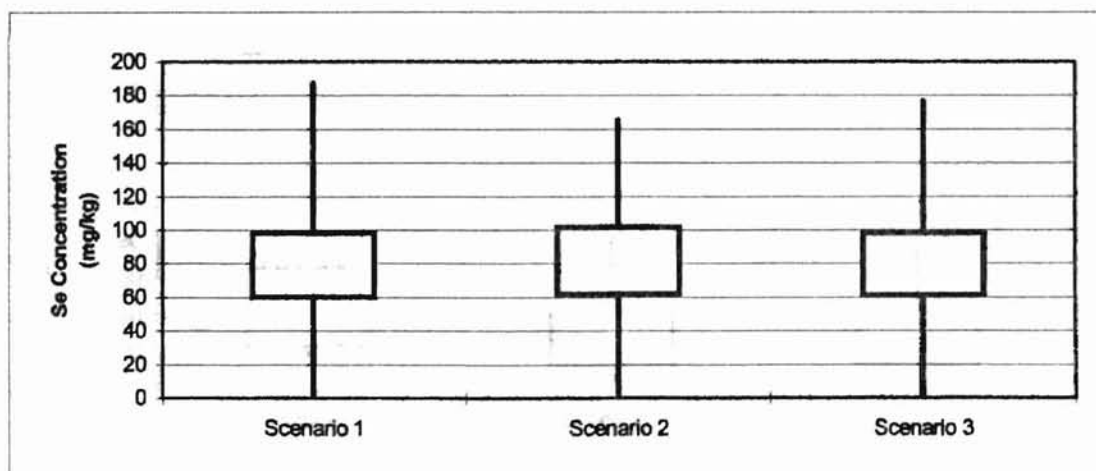


Figure 38. Probability distribution comparison for aquatic carnivorous invertebrates compartment.

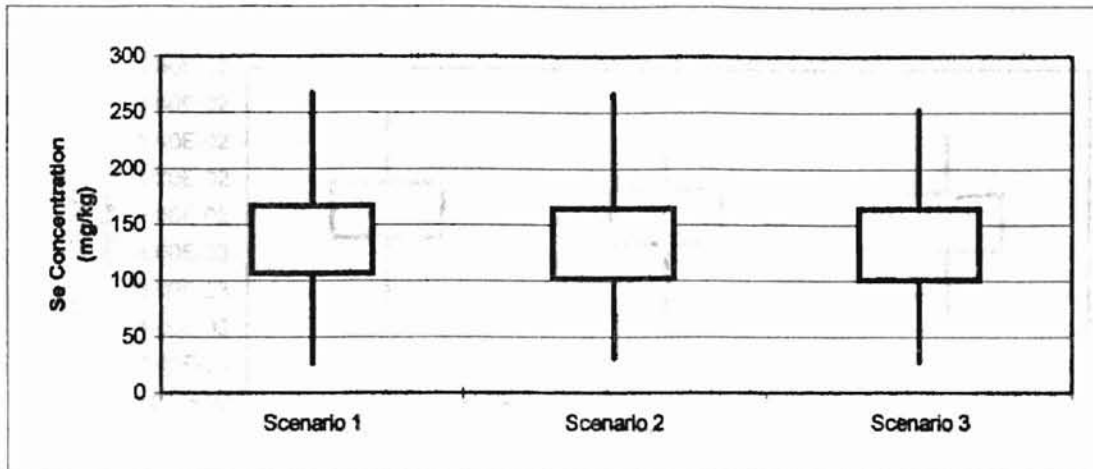


Figure 39. Probability distribution comparison for aquatic herbivorous invertebrates compartment.

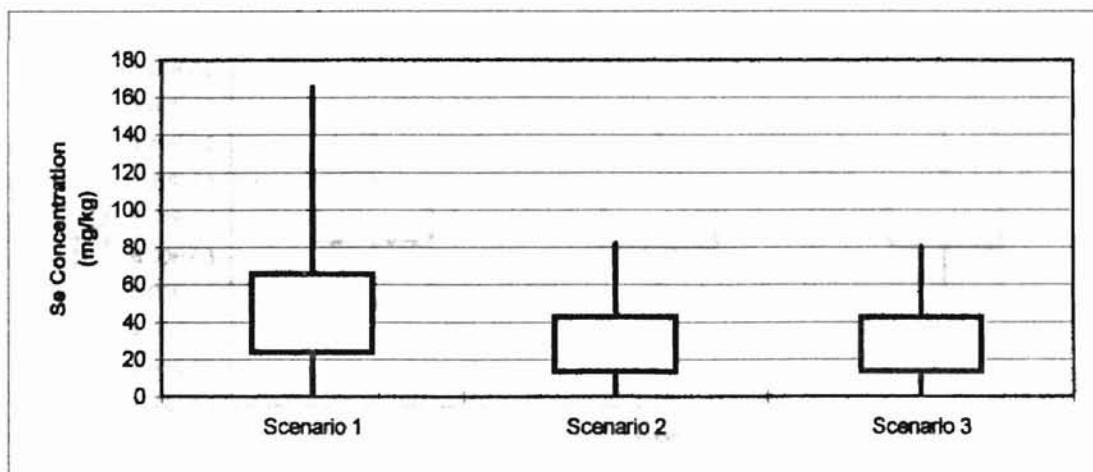


Figure 40. Probability distribution comparison for aquatic plants compartment.

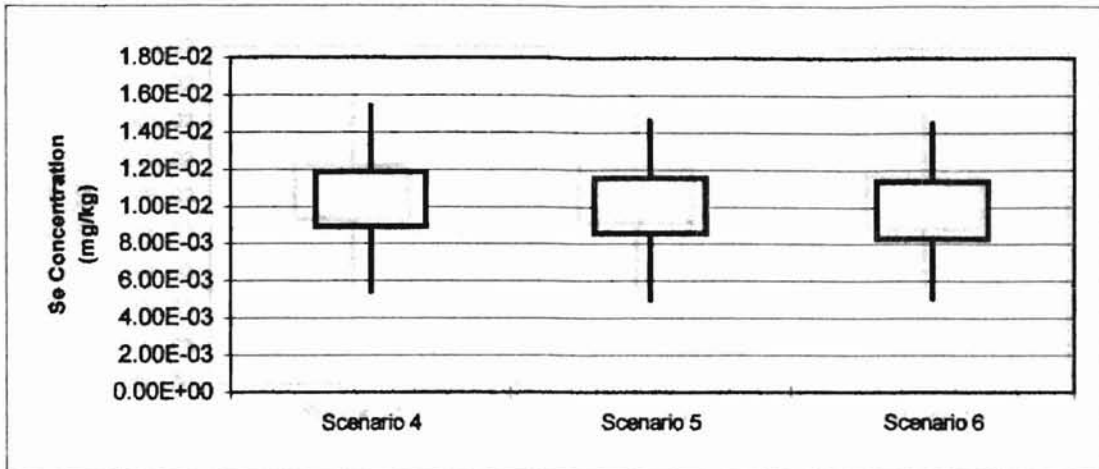


Figure 41. Probability distribution comparison for top carnivore compartment.

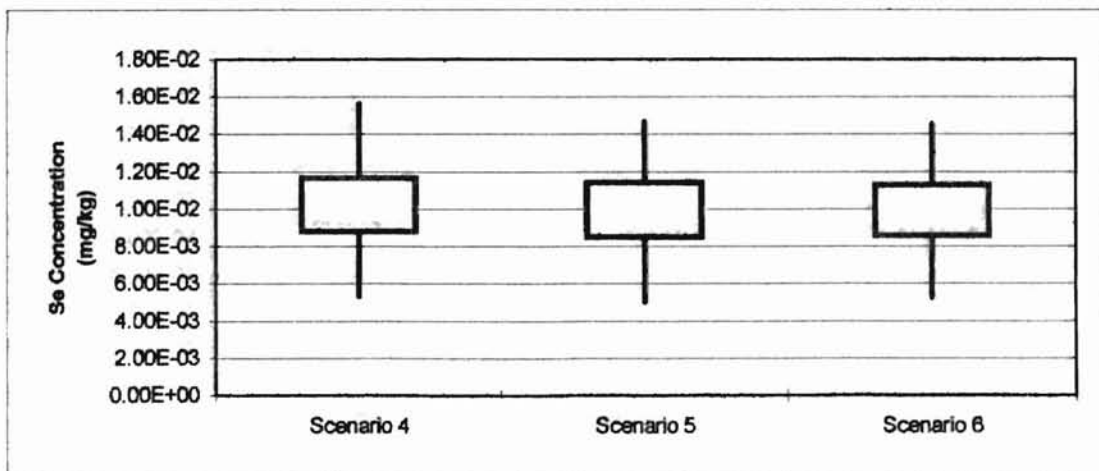


Figure 42. Probability distribution comparison for waterfowl compartment.

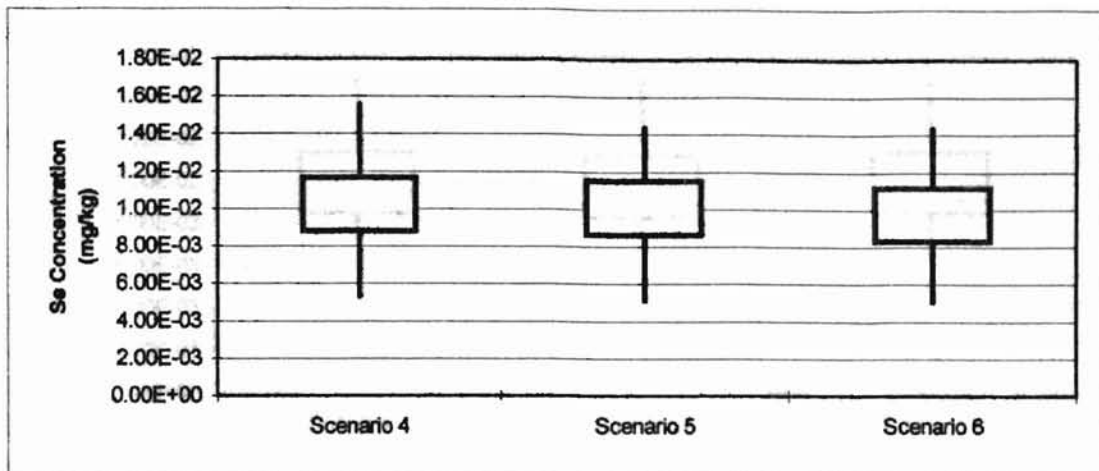


Figure 43. Probability distribution comparison for carnivorous mammals compartment.

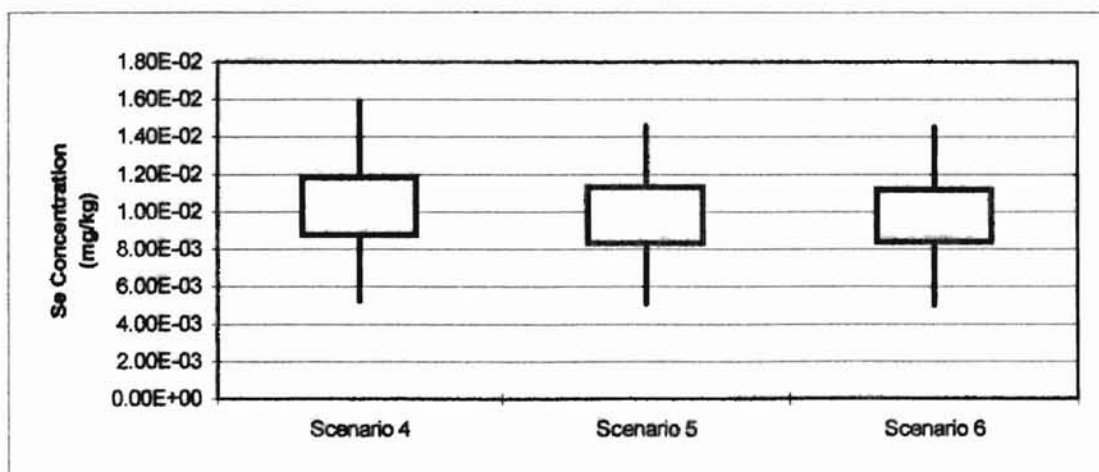


Figure 44. Probability distribution comparison for herbivorous mammals compartment.

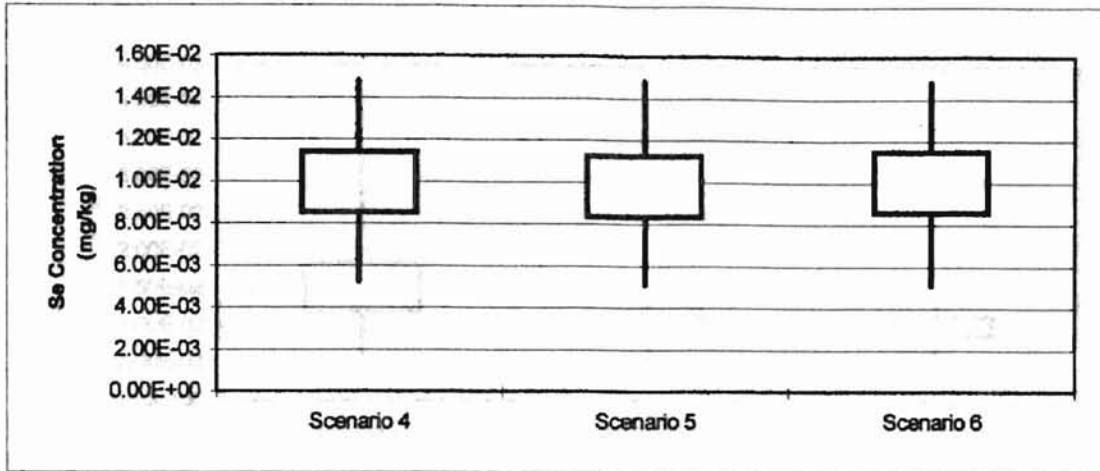


Figure 45. Probability distribution comparison for terrestrial carnivorous invertebrates compartment.

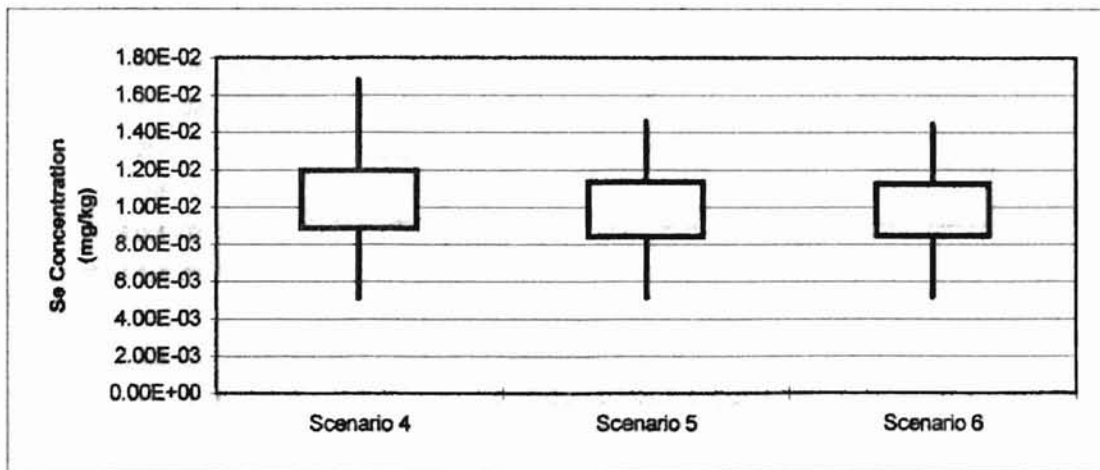


Figure 46. Probability distribution comparison for terrestrial herbivorous invertebrates compartment.

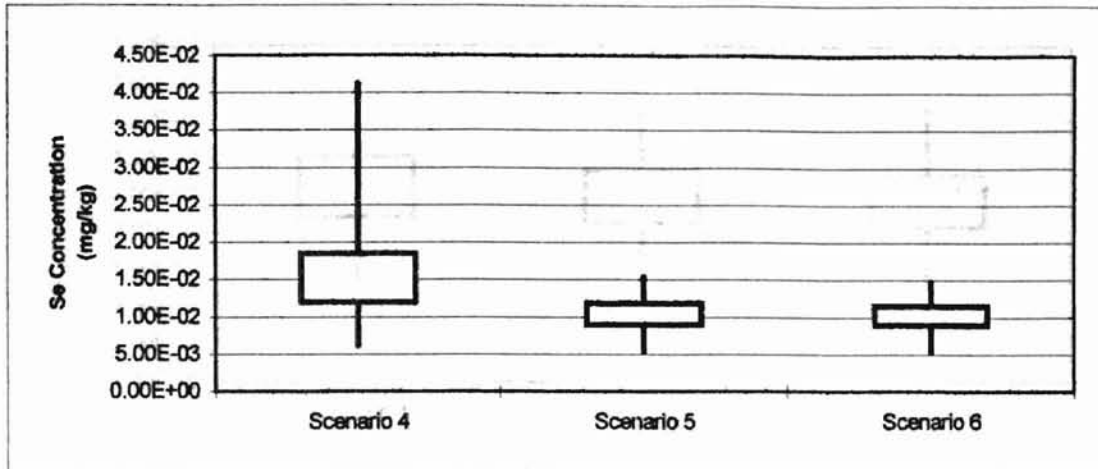


Figure 47. Probability distribution comparison for terrestrial plants compartment.

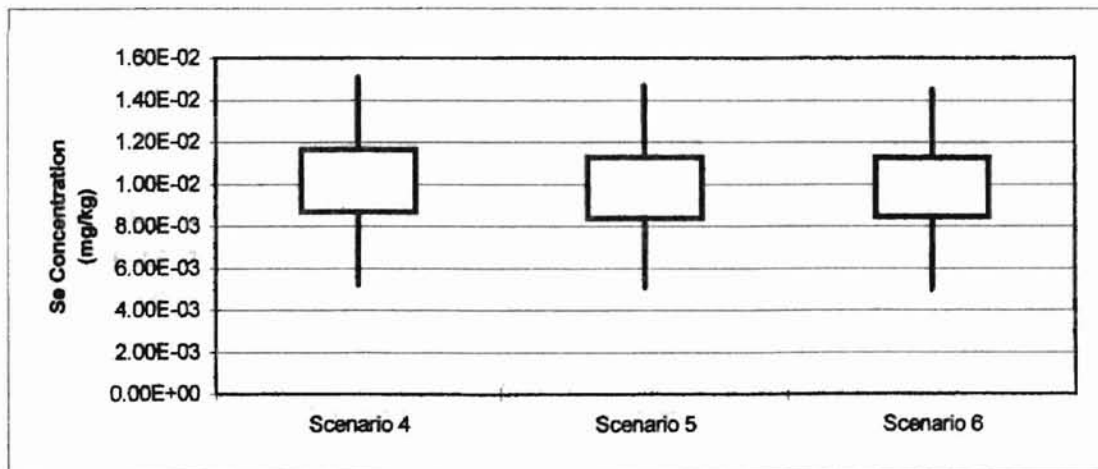


Figure 48. Probability distribution comparison for carnivorous fish compartment.

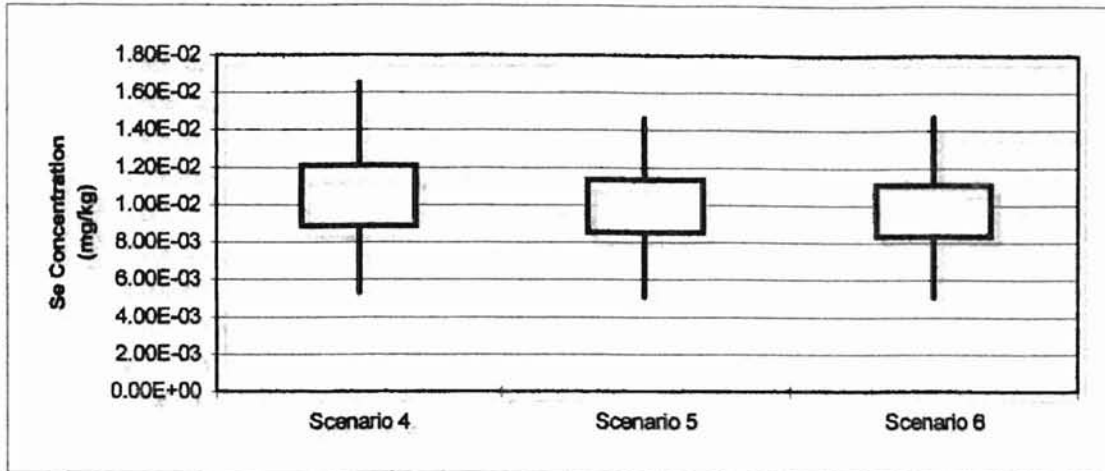


Figure 49. Probability distribution comparison for herbivorous fish compartment.

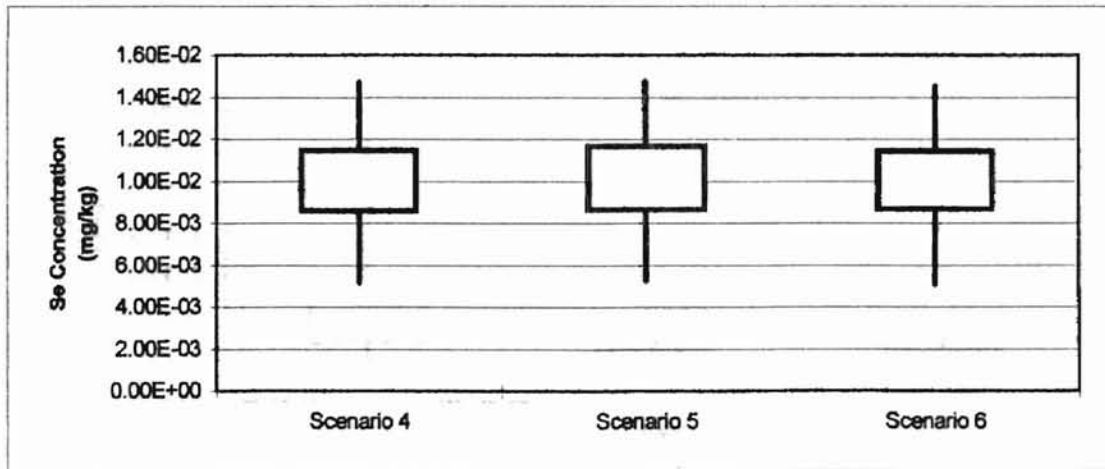


Figure 50. Probability distribution comparison for aquatic carnivorous invertebrates compartment.

4. DISCUSSION

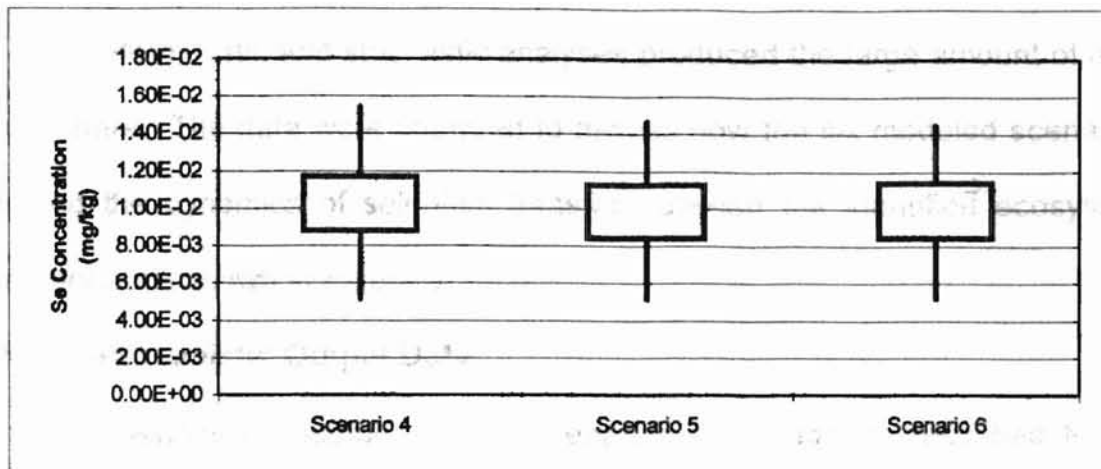


Figure 51. Probability distribution comparison for aquatic herbivorous invertebrates compartment.

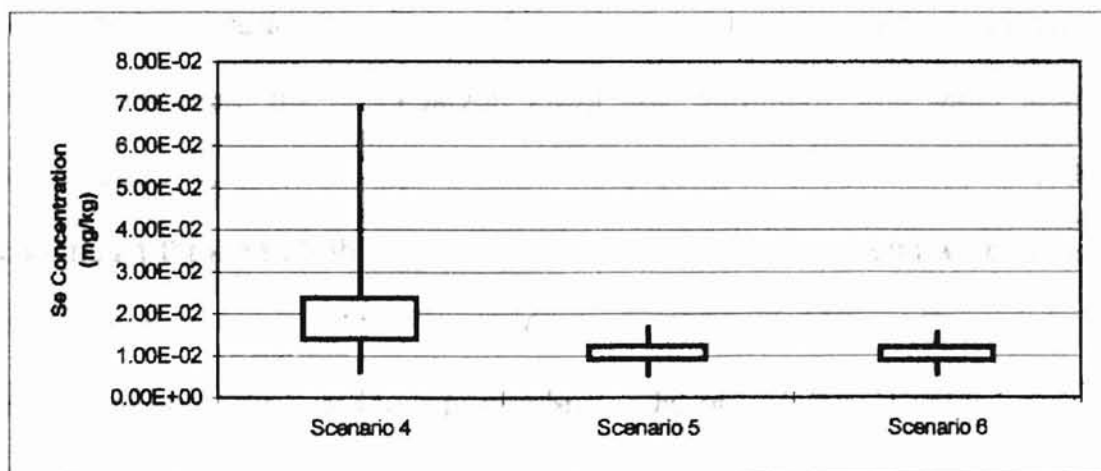


Figure 52. Probability distribution comparison for aquatic plants compartment.

is expected, the time required to reach the endpoint concentration increased
if the endpoint concentration within the three abiotic compartments
based on the differences were not realized in the to

4. DISCUSSION

The deterministic and stochastic analyses produced the large amount of data shown here. The data were analyzed to assess how the six modeled scenarios affected the dynamics of selenium transfer between the identified ecosystem compartments shown in Figure 3.

4.1 Deterministic Output Data

The deterministic output data for scenarios 1 through 3, described for all compartments in Figures 5 through 28, are represented by a fairly sigmoid curve which begins at the initial compartmental contaminant concentration (C_0) and then asymptotes toward 100% of the endpoint concentration (C_e). These data were used to compare the effects of the time required to reach C_e within each compartment under the successively lower soil, sediment, and water selenium concentrations suggested by CH2M Hill (1986). Under ASSUMPTION A (scenarios 1 through 3), the initial biotic selenium concentrations were set at the levels reported by CH2M Hill (1986) and USBR (1990). The underlying assumption here was that the reported levels did not reflect an equilibrium state between the biotic compartments and the ecosystem contamination. Under ASSUMPTION B (scenarios 4 through 6), a equilibrium state was assumed and the biotic C_e concentrations were set at the levels reported by CH2M Hill (1986) and USBR (1990).

As expected, the time required to reach the endpoint concentration increased for all compartments as the concentration within the three abiotic compartments decreased. However, appreciable differences were not realized in the top carnivore, waterfowl, carnivorous mammals, and aquatic carnivorous invertebrates compartments (see Figures 5, 6, 7, 12, and 14) under ASSUMPTION A criteria. A slight decrease in the time to C_e was calculated under ASSUMPTION A criteria for the terrestrial carnivorous and herbivorous invertebrates (Figures 9 and 10), herbivorous fish (Figure 13), and the aquatic herbivorous invertebrates (Figure 15) compartments. The most significant decreases for scenarios 1 through 3 were seen in the herbivorous mammals, terrestrial plants, and aquatic plants compartments as shown in Figures 8, 11, and 16 respectively. The ASSUMPTION B criteria results returned for scenarios 4 through 6 (Figures 17 through 28) provided the most striking stratification of time to C_e for all twelve biotic compartments.

In each case, the lower trophic levels generally appeared to be much more responsive to changes in the abiotic contaminant concentration than were the higher levels of the food web. Since increasingly higher trophic levels logically accumulate contaminant from increasingly more sources than the lower levels, this result would appear to be a plausible representation of natural food web dynamics.

4.2 Stochastic Output Data

As previously described, the stochastic analysis was twofold. First, a rank order correlation was performed which related model output results to the input

parameters. This portion of the stochastic analysis was done in order to measure the relative sensitivity the input parameters had on the final output. Secondly, the model was executed once for each of the five described scenarios in order to generate probability distribution functions for comparison of the various selenium contaminant levels at 1986 levels and the ONSITE-1 and ONSITE-2 cleanup goals.

4.2.1 Sensitivity of Outputs to Inputs

A rank order correlation analysis was performed for each compartment in an attempt to better understand the relative sensitivity each uncertain input variable had on the final output results. Not surprising, the initial contaminant concentration (C_0) had nearly a perfect positive correlation on the output results for all twelve biotic compartments. Unexpected however, was that this parameter was the only variable resulting in a strong correlation.

4.2.2 Stochastic Output Result Ranges and Probability Distributions

The Latin Hypercube analysis generated ranges and probability density functions (pdfs) for each compartmental contaminant concentration at the initial time step ($t = 1$). As before, the model was run once for each of the six scenarios. Summary results for scenarios 1 through 6 are presented in Tables 44 through 49. The intraquartile range and maximum and minimum values returned for each compartment under the six contaminant scenarios are compared in the box and whisker diagrams of Figures 29 through 52. Unlike the deterministic analysis, reducing the abiotic selenium concentrations did not universally reduce the modeled selenium concentration in all biotic

compartments under scenarios 1 through 6. In fact, for several compartments, the model returned high maximum values for scenarios 2 and 3 (ASSUMPTION A criteria) and scenarios 5 and 6 (ASSUMPTION B criteria) than for scenarios 1 and 4. The terrestrial and aquatic plants were the only two compartments which showed a significant downward shift in probability range (see Figures 35, 40, 47, and 52) under both assumption criteria. These two compartments represent the lowest trophic levels and could be assumed to reflect a decrease in soil, sediment, and water selenium concentrations prior to the other compartments.

storing the water selenium concentrations in the twelve biotic compartments. Selenium concentrations were not detectable in the lower biotic levels which is not surprising since the lower biotic elements of the modeled system are not directly exposed to the selenium source.

5. SUMMARY and CONCLUSIONS

Ecological risk assessment is a broad and complicated endeavor which, as described by Norton *et al.* (1992), requires analysts to measure the probability of ecological damage due to stressor contamination. In order to complete projects within generally acceptable timeframes and in an economically feasible manner, researchers must often times reduce complicated ecosystem interactions to simplified mathematical expressions (i.e., models). Unfortunately, no model will ever fully describe ecological interactions and very few input data sets are complete. For these reasons, risk assessors are faced with uncertainties surrounding not only the models chosen, but the input data used to calculate the output results. As such, the final results of an analysis must be interpreted with an understanding of the relative uncertainty inherent to the chosen model and input data. Stochastic modeling techniques can aid researchers in managing input parameter uncertainties. Selection of the proper model for a specific site or contaminant should be done very carefully so to reduce model uncertainties.

The model chosen for this work lent itself well to the compartment-based approach as well as incorporating first-order kinetics and equilibrium partitioning components into a single term. The modeled results under the six scenarios appear to make sense ecologically. For the deterministic assessment, as the soil, sediment, and water selenium concentrations were reduced, the model

returned decreased selenium concentrations in the twelve biotic compartments. As expected, decreases were most notable in the lower trophic levels which are more intimately related to the three abiotic elements of the modeled system. The stochastic analysis did not provide as clear a relationship between decreased abiotic selenium concentrations and the output data as did the point estimate approach. However, the two lowest trophic levels (terrestrial and aquatic plants) did show a marked decrease in compartmental selenium concentration as the abiotic compartments were reduced.

The results of the deterministic and stochastic analyses suggest that the ONSITE-1 and ONSITE-2 cleanup alternatives proposed by CH2M Hill (1986) would not likely benefit the current resident populations which are already "loaded" with selenium. However, future generations would enjoy a much reduced level of risk.

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**APPENDIX A – BESTFIT (1998) CURVE FITTING SUMMARY STATISTICS
FOR Se WATER CONTAMINANT CONCENTRATION
AT KESTERSON RESERVOIR**

Input Distribution	Normal	Weibull	Beta	Rayleigh	Logistic	Gamma	Erlang	PearsonVI	Lognorm	Lognorm2
Parameter 1	0.228	1.732	2.188	0.185	0.228	2.002	2.000	3.087	0.270	-1.744
Parameter 2	0.129	0.253	7.417		0.070	0.112	0.114	2.547	0.318	0.833
Parameter 3								0.112		
Minimum	0.002									
Maximum	0.540									
Mean	0.228	0.228	0.228	0.232	0.228	0.228	0.228	0.286	0.270	0.270
Mode	0.244	0.228	0.154	0.158	0.185	0.228	0.118	0.084	0.073	0.073
Median	0.230	0.228	0.205	0.208	0.218	0.228	0.192	0.185	0.175	0.175
Standard Deviation	0.129	0.129	0.134	0.129	0.121	0.128	0.180	0.161	0.318	0.318
Variance	0.017	0.017	0.018	0.017	0.015	0.016	0.028	0.026	0.101	0.101
Skewness	0.368	0.000	0.777	0.738	0.631	0.000	1.403	1.414	3.058	3.167
Kurtosis	2.422	3.000	3.354	4.024	3.245	4.200	5.953	8.000	20.888	23.700
Histogram										
Minimum	0.002	0.002	0.002	0.002	0.002	0.002	0.002	0.002	0.002	0.002
Maximum	0.540	0.540	0.540	0.540	0.540	0.540	0.540	0.540	0.540	0.540
P1	5.000	3.675	5.355	4.050	3.284	2.942	6.584	6.779	9.038	8.038
P2	10.000	6.438	10.248	10.110	8.007	6.578	12.058	12.100	14.381	14.724
P3	10.000	9.461	12.120	12.453	11.952	9.391	12.519	12.458	13.087	11.882
P4	10.000	11.874	11.840	12.211	12.881	12.980	10.921	10.834	9.808	8.790
P5	12.000	12.091	10.229	10.541	11.733	13.782	8.741	8.888	7.079	6.454
P6	10.000	10.512	8.043	8.304	9.342	11.002	6.647	6.599	5.121	4.788
P7	7.000	7.870	5.844	6.058	6.611	7.001	4.884	4.880	3.758	3.808
P8	5.000	4.998	3.980	4.112	4.187	3.641	3.502	3.485	2.807	2.757
P9	1.000	2.415	2.519	2.584	2.405	1.941	2.488	2.489	2.134	2.139
P10	3.000	1.042	1.510	1.511	1.248	0.940	1.711	1.651	1.681	1.681
Chi-Square										
Test Value	7.333	4.355	4.440	5.590	11.310	7.622	7.830	16.349	20.885	20.885
Confidence	>0.6	>0.88	>0.88	>0.78	>0.25	>0.15*	>0.55	>0.05	>0.01	>0.01
Rank	4	1	2	3	8	5	9	9	13	12
Kolmogorov-Smirnov										
Test Value	0.008	0.108	0.087	0.083	0.120	0.125	0.127	0.144	0.168	0.168
Confidence	>0.05	>0.025	>0.15*	>0.15*	>0.15*	>0.15*	>0.15*	>0.05*	>0.025*	>0.025*
Rank	2	4	3	1	6	7	8	9	10	11
Anderson-Darling										
Test Value	0.457	0.757	1.050	1.058	0.815	1.388	1.418	2.632	3.288	3.288
Confidence	>0.15	>0.025	>0.15*	>0.15*	>0.15*	>0.15*	>0.15*	>0.025*	>0.01*	>0.01*
Rank	1	2	4	5	3	7	8	9	10	11
Confidence										
Chi-Square										
Adjusted Value	7.333	4.355	4.440	5.590	11.310	7.622	7.830	16.349	20.885	20.885
Critical Value @ 0.750	5.899	5.899	5.899	5.899	5.899	5.899	5.899	5.899	5.899	5.899
Critical Value @ 0.800	8.343	8.343	8.343	8.343	8.343	8.343	8.343	8.343	8.343	8.343
Critical Value @ 0.850	11.389	11.389	11.389	11.389	11.389	11.389	11.389	11.389	11.389	11.389
Critical Value @ 0.900	14.884	14.884	14.884	14.884	14.884	14.884	14.884	14.884	14.884	14.884
Critical Value @ 0.950	18.919	18.919	18.919	18.919	18.919	18.919	18.919	18.919	18.919	18.919
Critical Value @ 0.995	19.023	19.023	19.023	19.023	19.023	19.023	19.023	19.023	19.023	19.023
Critical Value @ 0.999	21.889	21.889	21.889	21.889	21.889	21.889	21.889	21.889	21.889	21.889
Kolmogorov-Smirnov										
Adjusted Value	0.825	0.881	0.838	0.808	1.038	1.084	1.088	1.240	1.438	1.438
Critical Value @ 0.150	0.774		1.138	1.138	1.138	1.138	1.138	1.138	1.138	1.138
Critical Value @ 0.100	0.819		1.224	1.224	1.224	1.224	1.224	1.224	1.224	1.224
Critical Value @ 0.050	0.865		1.358	1.358	1.358	1.358	1.358	1.358	1.358	1.358
Critical Value @ 0.025	0.955		1.480	1.480	1.480	1.480	1.480	1.480	1.480	1.480
Critical Value @ 0.010	1.035		1.628	1.628	1.628	1.628	1.628	1.628	1.628	1.628
Anderson-Darling										
Adjusted Value	0.479	0.738	1.030	1.068	0.815	1.388	1.418	2.632	3.288	3.288
Critical Value @ 0.300	0.474									
Critical Value @ 0.100	0.578		1.610	1.610	1.610	1.610	1.610	1.610	1.610	1.610
Critical Value @ 0.050	0.658	0.637	1.933	1.933	1.933	1.933	1.933	1.933	1.933	1.933
Critical Value @ 0.025	0.787	0.757	2.462	2.462	2.462	2.462	2.462	2.462	2.462	2.462
Critical Value @ 0.010	0.918	0.877	3.070	3.070	3.070	3.070	3.070	3.070	3.070	3.070
Critical Value @ 0.005	1.062	1.038	3.857	3.857	3.857	3.857	3.857	3.857	3.857	3.857

**APPENDIX B – VISUAL BASIC (MACRO) CODE USED
FOR MODEL AUTOMATION PROCESSES**

```
'  
' RUN MODEL  
' Macro recorded 2/22/99 by Michael L. Thayer  
'  
Sub Run_Model()  
    Application.ScreenUpdating = False  
    Sheets("RAW").Select  
' Clear previous TOP CARNIVORE results  
    Range("B9").Select  
    Range(ActiveCell, ActiveCell.End(xlDown)).Select  
    Selection.Borders(xlLeft).LineStyle = xlNone  
    Selection.Borders(xlRight).LineStyle = xlNone  
    Selection.Borders(xlTop).LineStyle = xlNone  
    Selection.Borders(xlBottom).LineStyle = xlNone  
    Selection.BorderAround LineStyle:=xlNone  
    Selection.ClearContents  
    Range("C9").Select  
    Range(ActiveCell, ActiveCell.End(xlDown)).Select  
    Selection.Borders(xlLeft).LineStyle = xlNone  
    Selection.Borders(xlRight).LineStyle = xlNone  
    Selection.Borders(xlTop).LineStyle = xlNone  
    Selection.Borders(xlBottom).LineStyle = xlNone  
    Selection.BorderAround LineStyle:=xlNone  
    Selection.ClearContents  
    Range("D9").Select  
    Range(ActiveCell, ActiveCell.End(xlDown)).Select  
    Selection.Borders(xlLeft).LineStyle = xlNone  
    Selection.Borders(xlRight).LineStyle = xlNone  
    Selection.Borders(xlTop).LineStyle = xlNone  
    Selection.Borders(xlBottom).LineStyle = xlNone  
    Selection.BorderAround LineStyle:=xlNone  
    Selection.ClearContents  
' Clear previous WATERFOWL results  
    Range("F9").Select  
    Range(ActiveCell, ActiveCell.End(xlDown)).Select  
    Selection.Borders(xlLeft).LineStyle = xlNone  
    Selection.Borders(xlRight).LineStyle = xlNone  
    Selection.Borders(xlTop).LineStyle = xlNone  
    Selection.Borders(xlBottom).LineStyle = xlNone  
    Selection.BorderAround LineStyle:=xlNone  
    Selection.ClearContents
```

```

Range("G9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("H9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
'Clear previous CARNIVOROUS MAMMALS results
Range("J9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("K9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("L9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
'Clear previous HERBIVOROUS MAMMALS results
Range("N9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone

```

```

Selection.Borders(xlRight).LineStyle = xlNone
C Selection.Borders(xlTop).LineStyle = xlNone S INVERTEBRATES results
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone Select
Selection.ClearContents LineStyle = xlNone
Range("O9").Select No 9, LineStyle = xlNone
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone ne
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone 202
Selection.ClearContents
Range("P9").Select 202
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone 202
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents 202
'Clear previous TERRESTRIAL CARNIVOROUS INVERTEBRATES results
Range("R9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("S9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("T9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone

```

```

Selection.ClearContents
'Clear previous TERRESTRIAL HERBIVOROUS INVERTEBRATES results
Range("V9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("W9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("X9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
'Clear previous TERRESTRIAL PLANTS results
Range("Z9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AA9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AB9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select

```

```

Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
'Clear previous CARNIVOROUS FISH results
Range("AD9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AE9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AF9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
'Clear previous HERBIVOROUS FISH results
Range("AH9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AI9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone

```

```

Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AJ9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
'Clear previous AQUATIC CARNIVOROUS INVERTEBRATES results
Range("AL9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AM9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AN9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
'Clear previous AQUATIC HERBIVOROUS INVERTEBRATES results
Range("AP9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents

```

```

Range("AQ9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AR9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
'Clear previous AQUATIC PLANTS results
Range("AT9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AU9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
Range("AV9").Select
Range(ActiveCell, ActiveCell.End(xlDown)).Select
Selection.Borders(xlLeft).LineStyle = xlNone
Selection.Borders(xlRight).LineStyle = xlNone
Selection.Borders(xlTop).LineStyle = xlNone
Selection.Borders(xlBottom).LineStyle = xlNone
Selection.BorderAround LineStyle:=xlNone
Selection.ClearContents
'Calculate new results for TOP CARNIVORE
Range(Cells(8, 2), Cells(8, 4)).Select
Selection.Copy
Range(Cells(8 + 1, 2), Cells(8 + (Cells(4, 4).Value - 1), 4)).Select

```

```

ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 4).Value - 1), 2), Cells(8 + (Cells(4, 4).Value - 1),
4)).Select
With Selection.Borders(xlBottom)
    .Weight = xlMedium
    .ColorIndex = xlAutomatic
End With
'Calculate new results for WATERFOWL
Range(Cells(8, 6), Cells(8, 8)).Select
Selection.Copy
Range(Cells(8 + 1, 6), Cells(8 + (Cells(4, 8).Value - 1), 8)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 8).Value - 1), 6), Cells(8 + (Cells(4, 8).Value - 1),
8)).Select
With Selection.Borders(xlBottom)
    .Weight = xlMedium
    .ColorIndex = xlAutomatic
End With
'Calculate new results for CARNIVOROUS MAMMALS
Range(Cells(8, 10), Cells(8, 12)).Select
Selection.Copy
Range(Cells(8 + 1, 10), Cells(8 + (Cells(4, 12).Value - 1), 12)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 12).Value - 1), 10), Cells(8 + (Cells(4, 12).Value -
1), 12)).Select
With Selection.Borders(xlBottom)
    .Weight = xlMedium
    .ColorIndex = xlAutomatic
End With
'Calculate new results for HERBIVOROUS MAMMALS
Range(Cells(8, 14), Cells(8, 16)).Select
Selection.Copy
Range(Cells(8 + 1, 14), Cells(8 + (Cells(4, 16).Value - 1), 16)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 16).Value - 1), 14), Cells(8 + (Cells(4, 16).Value -
1), 16)).Select
With Selection.Borders(xlBottom)

```



```

.Weight = xlMedium
.ColorIndex = xlAutomatic
End With
'Calculate new results for TERRESTRIAL CARNIVOROUS INVERTEBRATES
Range(Cells(8, 18), Cells(8, 20)).Select
Selection.Copy
Range(Cells(8 + 1, 18), Cells(8 + (Cells(4, 20).Value - 1), 20)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 20).Value - 1), 18), Cells(8 + (Cells(4, 20).Value -
1), 20)).Select
With Selection.Borders(xlBottom)
.Weight = xlMedium
.ColorIndex = xlAutomatic
End With
'Calculate new results for TERRESTRIAL HERBIVOROUS INVERTEBRATES
Range(Cells(8, 22), Cells(8, 24)).Select
Selection.Copy
Range(Cells(8 + 1, 22), Cells(8 + (Cells(4, 24).Value - 1), 24)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 24).Value - 1), 22), Cells(8 + (Cells(4, 24).Value -
1), 24)).Select
With Selection.Borders(xlBottom)
.Weight = xlMedium
.ColorIndex = xlAutomatic
End With
'Calculate new results for TERRESTRIAL PLANTS
Range(Cells(8, 26), Cells(8, 28)).Select
Selection.Copy
Range(Cells(8 + 1, 26), Cells(8 + (Cells(4, 28).Value - 1), 28)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 28).Value - 1), 26), Cells(8 + (Cells(4, 28).Value -
1), 28)).Select
With Selection.Borders(xlBottom)
.Weight = xlMedium
.ColorIndex = xlAutomatic
End With
'Calculate new results for CARNIVOROUS FISH
Range(Cells(8, 30), Cells(8, 32)).Select
Selection.Copy

```

```

Range(Cells(8 + 1, 30), Cells(8 + (Cells(4, 32).Value - 1), 32)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 32).Value - 1), 30), Cells(8 + (Cells(4, 32).Value -
1), 32)).Select
With Selection.Borders(xlBottom)
    .Weight = xlMedium
    .ColorIndex = xlAutomatic
End With
'Calculate new results for HERBIVOROUS FISH
Range(Cells(8, 34), Cells(8, 36)).Select
Selection.Copy
Range(Cells(8 + 1, 34), Cells(8 + (Cells(4, 36).Value - 1), 36)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 36).Value - 1), 34), Cells(8 + (Cells(4, 36).Value -
1), 36)).Select
With Selection.Borders(xlBottom)
    .Weight = xlMedium
    .ColorIndex = xlAutomatic
End With
'Calculate new results for AQUATIC CARNIVOROUS INVERTEBRATES
Range(Cells(8, 38), Cells(8, 40)).Select
Selection.Copy
Range(Cells(8 + 1, 38), Cells(8 + (Cells(4, 40).Value - 1), 40)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 40).Value - 1), 38), Cells(8 + (Cells(4, 40).Value -
1), 40)).Select
With Selection.Borders(xlBottom)
    .Weight = xlMedium
    .ColorIndex = xlAutomatic
End With
'Calculate new results for AQUATIC HERBIVOROUS INVERTEBRATES
Range(Cells(8, 42), Cells(8, 44)).Select
Selection.Copy
Range(Cells(8 + 1, 42), Cells(8 + (Cells(4, 44).Value - 1), 44)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 44).Value - 1), 42), Cells(8 + (Cells(4, 44).Value -
1), 44)).Select

```

```

With Selection.Borders(xlBottom)
    .Weight = xlMedium
    .ColorIndex = xlAutomatic
End With
'Calculate new results for AQUATIC PLANTS
Range(Cells(8, 46), Cells(8, 48)).Select
Selection.Copy
Range(Cells(8 + 1, 46), Cells(8 + (Cells(4, 48).Value - 1), 48)).Select
ActiveSheet.Paste
Application.CutCopyMode = False
'Place bold line at bottom of Raw Output Results Table
Range(Cells(8 + (Cells(4, 48).Value - 1), 46), Cells(8 + (Cells(4, 48).Value -
1), 48)).Select
With Selection.Borders(xlBottom)
    .Weight = xlMedium
    .ColorIndex = xlAutomatic
End With
'Return to cell A1 of RAW DATA OUTPUT worksheet
Cells(1, 1).Select
'Return to cell A1 of OUTPUTS worksheet
Sheets("OUTPUTS").Select
Cells(1, 1).Select
End Sub

```

APPENDIX C – STOCHASTIC MODEL SIMULATION SUMMARY STATISTICS UNDER SCENARIO 1 CONDITIONS

Simulation Results for (INITIAL).xls

Iterations 1,000
 Simulations 1
 # Input Variables 141
 # Output Variables 12
 Sampling Type Latin Hypercube
 Parallel 80 PUS

Summary Statistics

	TOP CARBONS	WATERPOWL	CARBONICUS BUBBLES	HOMOGENEOUS BUBBLES	TERRESTRIAL CARBONICUS INVERTEBRATES	TERRESTRIAL HOMOGENEOUS INVERTEBRATES	TERRESTRIAL PLANTS	CARBONICUS PRRH	HOMOGENEOUS PRRH	TERRESTRIAL CARBONICUS PRRH	HOMOGENEOUS PRRH	AQUATIC CARBONICUS INVERTEBRATES	AQUATIC HOMOGENEOUS INVERTEBRATES	AQUATIC PLANTS
Minimum *	2.202	3.245	1.153	1.848	0.891	0.729	6.708	7.520	24.255	74.378	24.255	24.255	24.255	195.147
Maximum *	130.819	81.742	128.598	151.028	82.480	133.088	303.210	302.286	267.208	182.882	267.208	267.208	267.208	45.423
Mean *	54.271	20.698	80.275	54.478	21.239	29.287	157.208	158.395	138.391	79.741	138.391	138.391	138.391	27.217
Std. Deviation *	28.726	10.786	27.543	28.878	18.791	13.822	81.518	81.825	53.321	42.885	81.518	81.518	81.518	240.707
Variance *	828.108	115.809	759.653	833.717	352.729	194.085	2,667.005	2,482.024	2,843.341	1,838.471	2,667.005	2,667.005	2,667.005	58,960.860
Skewness *	0.348	0.642	0.291	0.109	0.854	0.407	-0.225	-0.225	0.048	-0.148	-0.225	-0.225	-0.225	0.800
Kurtosis *	2.179	2.913	2.118	2.098	2.378	2.344	3.004	2.704	2.618	4.714	3.004	3.004	3.004	3.100
Errors Calculated *	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
Std. Err.	58.207	9.254	27.857	64.422	9.821	11.727	92.028	98.278	65.828	41.033	92.028	92.028	92.028	21.887
95% Pers *	12.247	6.267	10.027	11.110	7.791	8.881	62.704	62.704	48.798	28.300	62.704	62.704	62.704	7.824
90% Pers *	16.170	7.735	14.081	15.167	9.811	7.703	84.088	84.088	61.462	44.334	84.088	84.088	84.088	11.034
85% Pers *	21.279	9.104	18.190	20.821	11.888	8.418	103.878	103.878	82.071	61.388	103.878	103.878	103.878	15.081
80% Pers *	25.421	10.295	22.388	26.347	13.541	9.757	115.822	115.822	92.388	64.794	115.822	115.822	115.822	18.678
75% Pers *	29.494	11.270	27.288	30.327	15.700	12.028	128.000	128.000	108.888	68.800	128.000	128.000	128.000	23.321
70% Pers *	34.477	12.883	35.149	35.871	18.389	13.382	133.421	133.421	118.549	74.185	133.421	133.421	133.421	28.882
65% Pers *	38.339	14.488	40.408	40.408	21.977	14.818	141.891	141.891	124.725	87.804	141.891	141.891	141.891	30.728
60% Pers *	44.200	16.073	44.028	44.028	23.271	16.818	148.337	148.337	132.809	101.091	148.337	148.337	148.337	34.028
55% Pers *	48.542	17.388	48.070	48.070	26.178	17.875	158.748	158.748	138.346	118.811	158.748	158.748	158.748	37.888
50% Pers *	51.084	18.084	51.388	51.388	28.488	18.888	162.842	162.842	142.846	124.811	162.842	162.842	162.842	41.811
45% Pers *	57.055	20.055	57.344	57.344	31.488	21.338	168.841	168.841	148.846	130.811	168.841	168.841	168.841	46.388
40% Pers *	61.671	22.488	64.418	64.418	34.028	23.133	172.744	172.744	152.846	134.811	172.744	172.744	172.744	50.427
35% Pers *	65.844	24.418	68.270	68.270	36.191	24.728	178.214	178.214	158.024	138.811	178.214	178.214	178.214	54.024
30% Pers *	71.058	26.391	72.787	72.787	41.871	28.422	188.317	188.317	168.088	148.811	188.317	188.317	188.317	58.424
25% Pers *	78.224	28.383	77.187	77.187	48.499	31.893	193.878	193.878	173.811	158.811	193.878	193.878	193.878	62.499
20% Pers *	81.805	30.532	81.980	81.980	51.289	34.185	203.877	203.877	183.811	168.811	203.877	203.877	203.877	66.499
15% Pers *	85.388	33.348	87.142	87.142	54.164	36.880	208.800	208.800	188.811	178.811	208.800	208.800	208.800	70.800
10% Pers *	91.278	36.182	91.278	91.278	58.481	41.811	218.811	218.811	198.811	188.811	218.811	218.811	218.811	74.811
5% Pers *	103.194	41.138	101.168	101.168	67.874	49.427	228.488	228.488	208.488	198.488	228.488	228.488	228.488	78.488

APPENDIX D - STOCHASTIC MODEL SIMULATION SUMMARY
STATISTICS UNDER SCENARIO 2 CONDITIONS

Simulation Results for CHSTE-14

Iterations 1,368
 Simulations 1
 # Input Variables 138
 # Output Variables 12
 Sampling Type Latin Hypercube
 Runtime 86:01:28

Summary Statistics

	TOP CARNIVORE	WATERFOWL	CARNIVOROUS MAMMALS	HERBIVOROUS MAMMALS	TERRESTRIAL CARNIVOROUS INVERTEBRATES	TERRESTRIAL HERBIVOROUS INVERTEBRATES	TERRESTRIAL PLANTS	CARNIVOROUS FISH	HERBIVOROUS FISH	AQUATIC CARNIVOROUS INVERTEBRATES	AQUATIC HERBIVOROUS INVERTEBRATES	AQUATIC PLANTS
Minimum *	-2.528	2.214	1.241	1.021	2.643	8.845	0.862	10.021	11.643	-118.727	30.892	0.482
Maximum *	132.098	88.154	131.548	117.822	83.084	47.874	81.032	288.813	311.234	185.348	288.807	81.408
Mean *	35.840	20.482	53.004	48.628	32.405	18.088	28.185	154.854	135.088	81.088	133.190	28.402
Std Deviation *	27.627	10.480	28.088	27.920	18.534	10.019	18.829	90.180	47.888	21.382	43.887	18.041
Variance *	778.813	108.211	787.810	781.204	343.510	100.377	347.088	2,817.888	2,303.844	884.840	1,808.480	362.880
Skewness *	0.127	0.541	0.180	0.307	0.638	0.488	0.517	-0.237	-0.087	-0.314	0.078	0.548
Kurtosis *	2.158	2.623	2.108	2.101	2.418	2.420	2.330	3.008	3.144	4.730	2.548	2.301
Errors Calculated *	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000	0.000
Mode *	80.888	13.175	38.843	73.818	8.154	22.080	8.745	148.428	191.888	88.023	183.484	8.888
5% Perc *	13.388	8.445	10.788	10.288	8.015	5.287	4.790	83.888	48.903	33.241	81.827	4.214
10% Perc *	18.210	7.851	15.319	13.154	10.218	7.379	6.677	88.880	88.888	42.830	74.014	8.774
15% Perc *	24.178	8.227	20.132	17.781	12.593	8.488	8.248	103.743	84.078	85.083	88.125	8.880
20% Perc *	28.878	10.420	24.203	21.088	14.405	8.771	11.085	114.127	88.887	87.314	82.487	10.820
25% Perc *	34.833	11.548	28.224	25.234	16.827	10.888	13.322	123.888	108.888	81.304	101.121	13.114
30% Perc *	38.255	13.088	34.210	30.880	18.087	12.188	15.535	131.728	113.883	88.844	108.884	15.741
35% Perc *	43.081	14.482	38.883	34.883	21.910	13.820	17.881	138.731	120.188	88.878	118.780	18.384
40% Perc *	47.275	15.883	43.324	38.351	24.081	14.881	20.477	145.087	128.375	72.824	121.310	21.088
45% Perc *	50.788	17.632	48.145	42.188	26.828	16.181	23.284	150.710	132.820	78.788	128.884	23.878
50% Perc *	54.751	18.382	52.445	48.331	28.548	17.308	25.138	158.828	142.287	80.728	133.885	26.488
55% Perc *	58.737	20.884	57.488	50.331	32.812	18.238	28.319	162.888	148.182	84.803	138.888	28.738
60% Perc *	64.900	22.381	61.804	55.782	35.081	20.670	32.014	168.088	158.513	87.440	145.848	32.088
65% Perc *	68.888	24.043	65.829	61.288	38.253	22.170	35.882	175.887	165.804	81.427	150.732	34.881
70% Perc *	74.281	25.878	68.813	68.001	41.925	24.008	38.113	182.412	180.788	88.810	158.748	38.307
75% Perc *	78.828	27.320	74.878	71.548	48.487	25.933	42.480	188.888	187.887	101.541	164.221	42.888
80% Perc *	83.013	28.738	78.787	76.841	48.888	28.888	48.881	197.234	173.787	108.728	171.870	47.008
85% Perc *	88.115	31.848	85.070	82.738	54.271	30.882	51.375	208.418	182.343	112.888	180.828	51.774
90% Perc *	95.113	35.828	81.308	88.548	58.888	33.813	57.278	218.887	182.112	118.884	181.788	57.887
95% Perc *	102.881	38.883	100.484	97.487	67.257	37.878	63.881	233.075	208.485	138.512	204.228	63.812

APPENDIX E – STOCHASTIC MODEL SIMULATION SUMMARY STATISTICS UNDER SCENARIO 3 CONDITIONS

Simulation Results for OARTEC-34

Iterations 1,000
 Simulations 1
 # Input Variables 138
 # Output Variables 12
 Sampling Type Latin Hypercube
 Random Seed 882134

Summary Statistics

	TOP CARIBBEAN	WATERPONT	CARIBBEAN MAMMALS	NONPOUOUS MAMMALS	TERRISTRIAL CARIBBEAN BIRDTERRITORIES	TERRISTRIAL NONPOUOUS BIRDTERRITORIES	TERRISTRIAL PLANTS	CARIBBEAN FISH	NONPOUOUS FISH	AQUATIC CARIBBEAN BIRDTERRITORIES	AQUATIC NONPOUOUS BIRDTERRITORIES	AQUATIC PLANTS
Minimum =	2,852	2,812	1,528	1,200	2,483	1,328	0,307	14,094	8,820	-176,718	27,198	0,000
Maximum =	148,328	85,911	130,381	117,848	81,187	68,782	78,878	302,887	305,181	578,071	282,383	80,027
Mean =	56,801	20,088	64,178	80,807	21,542	18,463	23,008	106,321	138,741	80,135	132,388	28,173
Std Deviation =	28,490	10,085	20,353	28,871	10,549	10,351	10,549	48,845	48,845	30,048	49,824	18,000
Variance =	810,400	112,381	403,800	835,277	111,248	108,827	111,248	2,384,488	2,384,488	902,887	2,483,888	325,000
Skewness =	0,179	0,090	0,148	0,228	0,141	0,094	0,073	-0,205	-0,202	-0,079	-0,103	0,091
Kurtosis =	2,205	2,782	2,024	2,021	2,400	2,090	2,400	2,800	2,800	2,898	2,408	2,368
Entropy Coefficient =	0,000	0,000	0,000	0,000	0,000	0,000	0,000	0,000	0,000	0,000	0,000	0,000
Mode =	74,137	7,421	47,388	18,383	27,859	10,370	28,488	172,808	143,874	74,081	188,174	11,027
90% Perc =	11,884	8,619	12,003	8,760	7,688	4,807	5,009	65,091	48,942	32,878	68,304	4,051
95% Perc =	16,824	7,478	18,401	13,983	9,389	6,089	7,235	88,887	64,810	43,071	72,048	7,441
99% Perc =	28,888	8,083	20,828	17,721	11,720	8,578	8,337	108,887	81,254	51,467	83,888	8,473
99% Perc =	27,478	10,178	28,443	21,728	13,900	9,784	11,008	110,280	82,889	58,280	81,187	11,132
99% Perc =	33,350	11,595	28,804	25,442	15,821	10,778	13,277	121,428	102,448	60,543	88,917	13,000
99% Perc =	37,788	12,887	34,023	28,884	18,108	12,191	15,481	132,788	113,851	64,710	103,821	18,078
99% Perc =	42,177	14,388	40,874	35,215	20,341	13,848	17,800	142,891	121,380	68,324	119,888	19,848
99% Perc =	47,383	15,888	44,878	38,382	23,141	15,288	20,882	147,887	128,888	72,143	121,712	20,888
99% Perc =	52,108	18,003	48,878	44,091	25,882	18,888	23,888	153,888	138,888	78,888	127,848	22,888
99% Perc =	58,134	18,888	52,788	48,887	28,877	19,378	25,888	158,484	141,713	81,713	132,888	24,888
99% Perc =	63,382	19,881	58,108	54,322	31,888	19,888	28,784	165,288	147,514	84,322	143,388	26,888
99% Perc =	64,334	21,088	62,518	58,288	34,281	21,321	31,328	171,348	152,887	87,548	148,881	28,888
99% Perc =	68,570	23,323	68,000	68,000	38,229	23,148	34,008	177,288	167,842	91,820	151,821	30,888
99% Perc =	71,228	28,422	71,285	68,781	41,014	24,803	38,784	183,288	184,442	94,872	157,882	32,888
99% Perc =	76,878	27,707	76,888	72,748	44,173	26,813	42,788	193,341	193,888	97,814	164,488	34,888
99% Perc =	81,388	28,388	81,388	77,388	48,382	28,514	48,712	202,888	202,888	102,888	173,381	36,888
99% Perc =	87,088	32,811	87,088	81,488	52,888	31,888	52,888	208,881	208,881	108,888	182,888	38,888
99% Perc =	93,888	38,482	93,888	81,182	58,888	34,888	58,888	215,888	215,888	118,442	187,488	40,888
99% Perc =	100,000	48,324	100,000	98,873	68,882	37,384	64,173	228,788	228,788	128,117	203,773	42,888

APPENDIX G – STOCHASTIC MODEL SIMULATION SUMMARY STATISTICS UNDER SCENARIO 5 CONDITIONS

Simulation Results for CHMTE-1.1

Iterations = 1,000
 Simulations = 1
 # Input Variables = 19
 # Output Variables = 13
 Sampling Type = Latin Hypercube
 Returns = 100,000

Summary Statistics

	TOP CARPPORE	WATERFOWL	CARNIVOROUS MAMMALS	HERBIVOROUS MAMMALS	TERRESTRIAL CARNIVOROUS INVERTEBRATES	TERRESTRIAL HERBIVOROUS INVERTEBRATES	TERRESTRIAL PLANTS	CARNIVOROUS FISH	HERBIVOROUS FISH	AQUATIC CARNIVOROUS INVERTEBRATES	AQUATIC HERBIVOROUS INVERTEBRATES	AQUATIC PLANTS
Minimum *	4,082-03	4,082-03	5,032-03	5,032-03	5,112-03	5,112-03	6,422-03	6,082-03	6,082-03	6,382-03	6,382-03	6,282-03
Maximum *	1,482-02	1,482-02	1,482-02	1,482-02	1,482-02	1,482-02	1,482-02	1,482-02	1,482-02	1,482-02	1,482-02	1,482-02
Mean *	9,812-03	9,812-03	9,812-03	9,812-03	9,812-03	9,812-03	1,032-02	9,812-03	9,812-03	9,812-03	9,812-03	9,812-03
Std Deviation *	2,082-03	2,082-03	2,082-03	2,082-03	2,082-03	2,082-03	2,082-03	2,082-03	2,082-03	2,082-03	2,082-03	2,082-03
Variance *	4,332-06	4,332-06	4,332-06	4,332-06	4,332-06	4,332-06	4,332-06	4,332-06	4,332-06	4,332-06	4,332-06	4,332-06
Skewness *	-0.722-01	-0.722-01	-0.722-01	-0.722-01	-0.722-01	-0.722-01	-0.722-01	-0.722-01	-0.722-01	-0.722-01	-0.722-01	-0.722-01
Kurtosis *	2.372-00	2.372-00	2.372-00	2.372-00	2.372-00	2.372-00	2.372-00	2.372-00	2.372-00	2.372-00	2.372-00	2.372-00
Errors Calculated *	0.022-00	0.022-00	0.022-00	0.022-00	0.022-00	0.022-00	0.022-00	0.022-00	0.022-00	0.022-00	0.022-00	0.022-00
Mean *	8,082-03	8,082-03	8,082-03	8,082-03	8,082-03	8,082-03	8,082-03	8,082-03	8,082-03	8,082-03	8,082-03	8,082-03
9% Perc *	6,482-03	6,482-03	6,482-03	6,482-03	6,482-03	6,482-03	6,482-03	6,482-03	6,482-03	6,482-03	6,482-03	6,482-03
10% Perc *	7,082-03	7,082-03	7,082-03	7,082-03	7,082-03	7,082-03	7,082-03	7,082-03	7,082-03	7,082-03	7,082-03	7,082-03
15% Perc *	7,782-03	7,782-03	7,782-03	7,782-03	7,782-03	7,782-03	7,782-03	7,782-03	7,782-03	7,782-03	7,782-03	7,782-03
20% Perc *	8,132-03	8,132-03	8,132-03	8,132-03	8,132-03	8,132-03	8,132-03	8,132-03	8,132-03	8,132-03	8,132-03	8,132-03
25% Perc *	8,482-03	8,482-03	8,482-03	8,482-03	8,482-03	8,482-03	8,482-03	8,482-03	8,482-03	8,482-03	8,482-03	8,482-03
30% Perc *	8,832-03	8,832-03	8,832-03	8,832-03	8,832-03	8,832-03	8,832-03	8,832-03	8,832-03	8,832-03	8,832-03	8,832-03
35% Perc *	9,182-03	9,182-03	9,182-03	9,182-03	9,182-03	9,182-03	9,182-03	9,182-03	9,182-03	9,182-03	9,182-03	9,182-03
40% Perc *	9,532-03	9,532-03	9,532-03	9,532-03	9,532-03	9,532-03	9,532-03	9,532-03	9,532-03	9,532-03	9,532-03	9,532-03
45% Perc *	9,882-03	9,882-03	9,882-03	9,882-03	9,882-03	9,882-03	9,882-03	9,882-03	9,882-03	9,882-03	9,882-03	9,882-03
50% Perc *	1,022-02	1,022-02	1,022-02	1,022-02	1,022-02	1,022-02	1,022-02	1,022-02	1,022-02	1,022-02	1,022-02	1,022-02
55% Perc *	1,062-02	1,062-02	1,062-02	1,062-02	1,062-02	1,062-02	1,062-02	1,062-02	1,062-02	1,062-02	1,062-02	1,062-02
60% Perc *	1,102-02	1,102-02	1,102-02	1,102-02	1,102-02	1,102-02	1,102-02	1,102-02	1,102-02	1,102-02	1,102-02	1,102-02
65% Perc *	1,142-02	1,142-02	1,142-02	1,142-02	1,142-02	1,142-02	1,142-02	1,142-02	1,142-02	1,142-02	1,142-02	1,142-02
70% Perc *	1,182-02	1,182-02	1,182-02	1,182-02	1,182-02	1,182-02	1,182-02	1,182-02	1,182-02	1,182-02	1,182-02	1,182-02
75% Perc *	1,222-02	1,222-02	1,222-02	1,222-02	1,222-02	1,222-02	1,222-02	1,222-02	1,222-02	1,222-02	1,222-02	1,222-02
80% Perc *	1,262-02	1,262-02	1,262-02	1,262-02	1,262-02	1,262-02	1,262-02	1,262-02	1,262-02	1,262-02	1,262-02	1,262-02
85% Perc *	1,302-02	1,302-02	1,302-02	1,302-02	1,302-02	1,302-02	1,302-02	1,302-02	1,302-02	1,302-02	1,302-02	1,302-02
90% Perc *	1,342-02	1,342-02	1,342-02	1,342-02	1,342-02	1,342-02	1,342-02	1,342-02	1,342-02	1,342-02	1,342-02	1,342-02

VITA

Michael L. Thayer

Candidate for the Degree of

Master of Science

Thesis: COMPARTMENT-BASED WETLAND ECOSYSTEM CONTAMINANT TRANSFER MODEL FOR ASSESSING ECOSYSTEM RISK

Major Field: Environmental Science

Biographical:

Personal Data: Born in Tulsa, Oklahoma on 13 April 1970, the son of David and Ann Thayer. Husband of Lisa Thayer and father of Sarah and David Thayer.

Education: Graduated from Nathan Hale High School, Tulsa, Oklahoma in May 1988; received Associate of Science degree in Biology from Tulsa Community College in May 1991; received Bachelor of Science degree in Wildlife Biology from Northeastern State University, Tahlequah, Oklahoma in May 1994. Completed the requirements for the Master of Science degree with a major in Environmental Science at Oklahoma State University, Stillwater, Oklahoma in May 2000.

Experience: Laboratory Technician for ICI Explosives, Inc. in Joplin, Missouri, January 1992 to August 1992; worked two summers (1993 and 1994) with the U.S. Fish and Wildlife Service restoring wetland ecosystem habitats throughout the State of Oklahoma; employed as Physiology Laboratory Assistant at Northeastern State University during Fall (1993) and Spring (1994) semesters; employed by Fibercast Company, Sand Springs, Oklahoma as Environmental Coordinator, 1996 to present.